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1	Chapter 13
2	Energy production in anaerobic membrane bioreactors:
3	Opportunities and challenges
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18 Abstract

Anaerobic Membrane Bioreactors (AnMBRs) have been favored over their aerobic counterparts 19 20 because the former have the potential to recover energy from different types of wastewater streams. Currently, the majority of these AnMBR operations involve big improvements in methane 21 production. Technical studies have been undertaken showing that biohydrogen and Volatile Fatty 22 Acid (VFA) can also be a potential source of energy recovered from the AnMBR. Recent 23 developments in AnMBR design have made possible the production of methane and biohydrogen 24 or VFA simultaneously in various stages of the bioreactor. Controlling process inhibition and 25 26 improving the energy density through post-treatment can be challenging for sustainable energy 27 recovery from this system. There are challenges in designing a safe and reliable storage system for biohydrogen. Economically, VFA has not yet been considered a viable option due to the costintensive separation and purification process. Nonetheless, overcoming these challenges can lead
to a potential solution as a way to maximize energy recovery from AnMBRs.

31 Keywords: Anaerobic, fouling, methane, biohydrogen, energy

32 1. Introduction

33

Anaerobic Membrane Bioreactors (AnMBRs) can recover energy from different wastewater streams through the degradation of organic waste. In waste management, the anaerobic process has been favored over the aerobic process as it offers an energy recovery option through the production of methane-containing biogas. Furthermore, any full-scale operations of the anaerobic process are not yet possible in the waste management industry due to a low energy density, poor cost effectiveness and technical issues in the process operations as well [1].

40

Over the past few years, energy recovery has been mainly confined to generating methane which is the final product of anaerobic digestion. Anaerobic digestion is a combination of four major biochemical steps: bacterial hydrolysis, acidogenesis, acetogenesis, and methanogenesis. Methanogenesis has been identified as the most critical step as it is the slowest among all other stages in anaerobic digestion (AD). It is also dependent under strict operating conditions. As a result, improving methane production using the AD process was extensively studied in order to improve its energy efficiency [2].

48

The opportunities available in energy recovery from the AnMBR can be divided into two majorstreams. Firstly, the operating conditions of AnMBR can be optimized for final product recovery

without a change in the AnMBR design. This approach also involves integrating pre- and posttreatment processes of wastewater streams to improve product yields. Secondly, the product spectrum can be controlled for an AnMBR to produce Volatile Fatty Acid (VFA), and biohydrogen instead of methane. The second approach includes design modification, optimizing the process parameters, coupled with the inhibition of the methanogenesis process [3, 4].

56

57 With the aim of producing methane, AnMBR technology is now at a more mature level of 58 development. Studies have shown that the growth rate for methanogenic microorganisms is higher in thermophilic conditions (50-70 °C) compared to the rate in mesophilic or psychrophilic 59 60 conditions. A pH range of 6.8-7.4 has been identified as ideal for the production of methane using the anaerobic process. Unlike temperature and pH, the optimum value of Hydraulic Retention 61 Time (HRT) and Solid Retention Time (SRT) cannot be defined universally as both depend on the 62 63 design of the bioreactor, as well as the composition of feed material, including the additional operating conditions like temperature and pH [5]. Apart from the process parameters, chemical 64 additives like nanoparticles containing Co, Ni, FE, Fe₃O₄, biochar, ozone, etc., and treatment 65 processes like high pressure, ammonia soaking, alkaline pre-treatment have also been applied to 66 improve the production of methane [5-7]. Despite these improvements, methane production has 67 still not resulted in a net overall revenue earned from using the AnMBR. Hence, additional 68 challenges lie in the percentage of methane actually produced from biogas in the AnMBR. Pipeline 69 70 quality bio-methane production, and CO₂ removal from produced biogas are considered the major 71 challenges of energy recovery from the AnMBR.

VFAs have been identified as the potential raw materials of Polyhydroxyalkanoates (PHA), and 73 these include aldehydes, ketones, alcohols, and biogas. As a result, these are a potential resource 74 of energy recovery from the AnMBR. Economically, the production of VFA can be beneficial 75 compared to the revenue earned from the production of methane through anaerobic digestion. 76 Kleerebezem et al. (2015) compared the revenue earned from cardboard wastewater. The 77 comparison showed that the daily revenues earnt from methane and PHA were 3.6 and 20.2 k€ 78 79 respectively [8]. At the same time recovering VFA helps to avoid the problem of VFA 80 accumulation in the AnMBR. This helps to maintain a stable level of pH inside the reactor [9]. Therefore, producing VFA from an anaerobic process can be beneficial from both technical and 81 82 economic viewpoints. Challenges still lie ahead for the separation and purification technologies involved in VFA recovery. For an anaerobic VFA production process, the costs associated with 83 extracting VFA are a major challenge for sustainable VFA production using the AnMBR. 84

85

Production of biohydrogen using the anaerobic process is favored for methane production due to 86 a number of reasons. For example, the common fuel properties such as energy density, wide 87 flammability limit, and environment-friendly combustion have made biohydrogen a better energy 88 source over methane. Whilst AnMBRs are designed to produce biohydrogen instead of methane 89 this does not have the negative environmental impact caused by the dissolved methane. 90 91 Simultaneously, production of hydrogen and methane can improve the overall product revenue of the AnMBR. However, the biggest challenge in biohydrogen production is in designing a safe 92 storage and transportation system for biohydrogen. Low storage density, hydrogen loss due to the 93 boiling-off phenomenon, and methane embrittlement due to hydrogen penetration are the major 94 issues in biohydrogen storage and transportation [4]. 95

96

97 This chapter includes the most recent opportunities that AnMBR technology can offer in terms of 98 energy recovery. Although the different designs and multiple product recovery solutions do offer 99 an improved energy recovery from the AnMBR. The current technical and economic challenges 100 must be overcome to replicate the research successes in industrial practical applications.

101

- 102 2. Current energy production scenario
- 103

The current status of energy production in an AnMBR is mainly dependent on the production rate and yield of methane. Although VFA and biohydrogen are considered to be useful resources, they have been recovered with methane as a by-product. The amount of energy recovered from an anaerobic process depends on feed composition, bioreactor design and arrangement, type of preand post-treatment processes and above all on the microbial activity. Table 1 lists some examples of anaerobic membrane bioreactors, and their energy production in terms of methane, biohydrogen, and VFA production.

111

insert Table 1

112

Table 1 shows a minimum of 86% and a maximum of 99% COD removal for different wastewater streams. Landfill leachate treated in the cross-flow ultrafiltration AnMBR indicated the highest methane production of 460 L/kg COD. The potential of AnMBR in energy production can be evaluated through COD mass balance, and biogas production in terms of methane. The COD fed into the system is usually divided considering COD present in the bioreactor. For example, Gianico et al. (2013) have identified a consumption amount of 1.42 g COD for 1 g of biomass, and the total amount is converted into 0.35 L of methane under standard temperatures and pressure. Therefore,
for a given anaerobic process the potential energy production in the form of methane can be
calculated through the amount of COD consumed by the system [22, 23].

122

However, energy recovery from the AnMBR through the production of methane needs to address 123 the technical, economic and environmental issues. Since the final stage of anaerobic methane 124 125 production is the slowest (AD stage), the production of methane containing biogas is, in fact, a 126 very slow process. Different pre- and post-treatment processes integrated with the AnMBR technology have been effective to accelerate the initial hydrolysis or acidogenesis process, but it 127 128 cannot necessarily increase the speed of methane production to a great extent [1]. Currently, the percentage of methane in the produced biogas has become a higher concern. For different 129 AnMBRs, the composition of produced biogas depends on the bacterial community, substrate, feed 130 131 composition, and operating conditions [3, 24, 25]. For industrial application, it is a challenge to maintain the same feed composition at long-term AnMBR operations. Consequently, the methane 132 composition in the produced biogas cannot be maintained at a fixed value. 133

134

Energy recovery from the AnMBR through methane is also largely affected by the possible inhibition of methanogenesis due to the chemicals produced in the intermediate stages of anaerobic digestion. AnMBR operating in a high Organic Loading Rate (OLR) or short Hydraulic Retention Time (HRT) experiences a fast hydrolysis process that eventually leads to a higher rate of VFA production. The produced fatty acids can accumulate inside the reactor as they are consumed at a slower rate by the methanogens. The accumulated fatty acids can be responsible to cause a sharp decrease in the reactor pH and make the operation unstable. 142

Production of methane involves environmental outcomes like aquatic eco-toxicity, human toxicity, 143 abiotic depletion and above all global warming [26-28]. The combustion product of methane 144 includes CO₂ and CO based on the oxygen supply. Both these combustion products are potentially 145 146 responsible for increasing the effects of global warming from using the AnMBR. At room temperature, about 22.7 mg of methane can be present in each litre of bioreactor effluent as 147 148 dissolved methane. The dissolved methane in the AnMBR effluent is directly responsible for causing adverse impacts on the environment, such as freshwater eco-toxicity and human toxicity. 149 Furthermore, the net energy production from the AnMBR depends on the amount of energy 150 required in AnMBR operations. The current mode of biogas production requires biogas scouring 151 but this consumes most of the energy that an AnMBR requires for its operation. Khan et al. (2016) 152 in their study reported that the gas scouring in a AnMBRs treated a volume of 20,000 m³ municipal 153 154 wastewater. This required 46.7% of the total energy cost. Their study also noted that 73.5% of the total energy was consumed by a biogas recycle blower [recycling blower?] in an AnMBR treating 155 $(3.2 \pm 0.7 \text{ m}^3/\text{day})$ sulphate-rich urban wastewater [9]. 156

157

The concept of recovering biohydrogen and VFA from AnMBR is relatively new. Only a few studies have so far been published on producing biohydrogen and VFA from AnMBRs. Table 2 lists some commonly used AnMBR configurations along with the production rates, and yields of biohydrogen for each system.

162

163

insert Table 2

From Table 2, it is evident that only a few research studies have been done on producing 165 biohydrogen from AnMBR. This suggests there is not enough data in these research studies that 166 actually compares the expected revenues and operating costs. Therefore, it is difficult to 167 demonstrate the feasibility of biohydrogen production form an economic viewpoint. Apart from 168 biogas and biohydrogen production, extracting VFA from AnMBR has been a major challenge as 169 this actually limits the economic feasibility of VFA production. Production of VFA from AnMBR 170 171 has not been widely accepted for many reasons. One of the primary concerns is associated with 172 the cost involved in separation, and post-treatment of VFA. The production of VFA can be maximized at low HRT and high OLR. Both these operating conditions are responsible for causing 173 174 severe membrane fouling. The fouling layer can eventually retain VFA inside the anaerobic bioreactor, and make the separation and purification processes even more challenging [38, 39]. On 175 this issue, Khan et al. (2019) carried out an experiment to produce VFA from low strength synthetic 176 177 wastewater using a single stage AnMBR [3]. There results showed the highest VFA yield of $48.20 \pm 1.21\%$ (mg VFA/mg COD_{feed}) at HRT lasting 8 hrs. The corresponding membrane fouling 178 rate was higher compared to the rates observed at HRT lasting 48, 24, 18, and 12 hrs. The same 179 study revealed an overall reduction in the VFA yield due to COD removal efficiency being at 180 35.39% when the OLR rose from 68.75 to 89.38 mg COD/L.h. 181

182

The present scenario of energy recovery from the AnMBR demonstrates not enough research has been undertaken to extract VFA and biohydrogen. The issues involved in bioreactor operation along with the costs involved in product recovery have for this reason made AnMBR a less attractive option in energy recovery.

188 3. Energy production opportunities

189 3.1 Production of biohydrogen

Biohydrogen is produced from the volatile fatty acids during acidogenesis, and acetogenesis stages
of anaerobic digestion [4]. Compared to methane, hydrogen has potential advantages, for instance,
higher energy density and clean combustion production. Additional fuel properties of hydrogen
have been compared with methane and these are summarized in Table 3 below.

- 194
- 195

insert Table 3

196

From Table 3 it is evident that, besides energy density, hydrogen has significantly higher specific 197 heat constant, wider flammability limit in air (%), higher flame temperature (K) and greater 198 199 explosion limit. As a result, biohydrogen can be considered a better alternative than methane when fuel properties are taken into consideration. During the acidogenic phase, production of 200 biohydrogen can be divided into three different types. H₂ and CO₂ are produced with acetic and 201 butyric acid during butyrate-type fermentation. Propionate-type fermentation does not contribute 202 203 significantly to biohydrogen and CO₂ production. Acetic, propionic and valeric acids are the major products derived from this type of fermentation. Ethanol-type fermentation involves the 204 production of ethanol, and acetic acid with biohydrogen and CO₂. VFA produced during this stage 205 goes through the third anaerobic stage called acetogenesis. During this stage, the produced volatile 206 fatty acid components are again converted to acetates and biohydrogen. The amount of 207 biohydrogen produced throughout the third acetogenic stage is significantly larger compared to 208 the amounts generated during the second stage. The resulting biohydrogen from the second and 209 third anaerobic stages is directly consumed in the final stage of anaerobic digestion. At this point, 210

hydrogenotrophic methanogens consume the available biohydrogen to produced methane and
carbon dioxide. Biohydrogen acts as a proton donor at this stage. Consequently, it is important to
inhibit the activities of methanogens for the production of biohydrogen [4].

214

Type of substrates plays a very important role in improving the yield of biohydrogen produced from the anaerobic process. Although only very few research studies have been done on generating biohydrogen from the AnMBR, different anaerobic digestion processes have demonstrated promising results. Table 4 shows the highest biohydrogen yield achieved from a range of different substrates and bioreactor configurations. Findings from these studies can be applied to AnMBRs, and it can eventually contribute to improving the amount of energy recovered from it.

221

insert Table 4

222

223 Table 4 shows that a fluidized bed reactor produces the highest biohydrogen yield (4.26 mol H₂/mol sucrose). It is significantly larger than the other biohydrogen production processes. 224 Continuous stirred-tank and fluidized bed reactors have proved to be efficient in biohydrogen 225 production. Modified designs can be applied to existing AnMBR technology to improve 226 biohydrogen production. Hexose and glucose are two major carbon sources that have shown a high 227 conversion rate to biohydrogen. Some research studies have set out to optimize the production of 228 229 biohydrogen. Temperature, pH, Hydraulic Retention Time (HRT), Solid Retention Time (SRT) and Organic Loading Rate (OLR) have been further refined for different processes involved in 230 generating biohydrogen (both HRT and SRT), organic loading rate and specific chemical additives 231 that enhance biohydrogen production. 232

Temperature has a positive effect on the production of biohydrogen through the anaerobic process. 234 Since hydrogen is produced at the second and third anaerobic stages, a high rate of initial 235 hydrolysis and biomass acclimatization can increase the rate of biohydrogen being produced. In 236 general, high temperature favors the biomass acclimatization and the rate of initial hydrolysis. For 237 example, Zhong et al. (2015) revealed that the biohydrogen production rate increased from 116.5 238 to 131.5 ml/g-COD when the temperature rose from 40 to 60 °C [55]. High temperatures can also 239 240 be effective in inhibiting the microbial activity of the methanogens. According to the findings 241 documented in Jariyaboon et al. (2015), anaerobic sludge treated at 100 °C for 30 minutes can inhibit the activity of the methanogens. Unfortunately, the findings from this research are still 242 243 confined to laboratory scale applications [56]. For industrial processes, the feasibility of using high temperature can be assessed based on the expected higher revenue generated from the 244 biohydrogen. 245

246

pH is the second operating condition that can be optimized so that the ability to produce biohydrogen is maximized. Biohydrogen is favored within a pH range between 5.5-6.8, whereas a pH level below 4.5 is said to be inhibitory. Most of the research findings have identified pH 5.5 to be the optimum value for the production of biohydrogen [4].

251

HRT, SRT and OLR are the operating conditions depending on the design and arrangement of the bioreactor, including the type and composition of the substrate. Hence, it is important to understand these changes in biohydrogen production when the parameters change for different anaerobic hydrogen production processes, as a change in HRT or SRT actually changes the optimum OLR in an anaerobic process. Technically, the HRT of a bioreactor is designed to inhibit the activity of hydrogen consuming hydrogenotrophic methanogens whilst simultaneously preventing the washout of hydrogen produced bacteria. In general, a smaller HRT can increase the production rate of biohydrogen [5]. Angeriz-Campoy et al. (2015) demonstrated that HRT values between 3 to 6 hrs can maximize the biohydrogen production rate (25.9 L H₂/L-d) from granular hydrogen producing mixed cultures [57]. An experiment by Kumar et al. (2014) reveals that the highest biohydrogen production rate (4.49 L/L/d) was attained at a HRT range between 6-18 hrs [58].

263

The OLR and HRT share a unique relationship to maximize the production of biohydrogen in an anaerobic process. For AnMBRs, it is particularly challenging as a high organic loading rate can be responsible for severe membrane fouling and could make the operation unstable through having a high production rate of VFA. Therefore, it is rather practical to ascertain the optimum pair of HRT and OLR rather than optimizing them individually. For example, for a given anaerobic process, a particular HRT might involve an OLR high enough to make the operation of AnMBR unstable.

271

Several research studies have been conducted to carry out the influence of HRTs and OLRs on biohydrogen production. According to Zhang et al. (2013) the production of hydrogen in an anaerobic CSTR rose when the operating conditions were adjusted to 60 g-glucose/L-reactor/day of 6 hrs HRT using 20 g-glucose/L/day, and 12 hrs of HRT [59]. This was then followed by another experiment to optimize the HRT, and OLR simultaneously. Four different OLRs (10, 15, 20 and 40 g/L/day) were applied at 6, 12 and 24 hours of HRT. These results determined that a combination of 40 g/L/day and HRT of 12 hrs provides the maximum yield of hydrogen [60].

The opportunities in energy recovery through the production of biohydrogen still require extensive research and new initiatives to produce feasible results from both a technical and economic perspective. Different types of substrates, design modifications, and ensuring process optimization are evident approaches to increase the production of biohydrogen using AnMBRs.

284

285 3.2 VFA Recovery

286 The products of VFA include biogas and alcohol which are deemed to be a useful source of energy. This recovery of VFA could then contribute to the improvement of the energy recovery from the 287 AnMBR. During anaerobic digestion, the production of VFA can occur in two different ways. 288 Initially, the complex organic compounds present in the feed are converted to soluble 289 carbohydrates, amino acids and fatty acids through the process of hydrolysis. VFA is also produced 290 at the second anaerobic stage when the products in the initial stage are converted again to produce 291 292 VFA. As VFA is consumed to produce methane at the final anaerobic stage, a very low rate in VFA production can result in a low production rate of biogas. In contrast, an excessive production 293 of VFA can have adverse effects on the total production of (final) biogas product as the rate of 294 consumption of VFA is slower compared to the initial anaerobic stages of VFA production. In fact, 295 a major problem in the AnMBR is the accumulation of VFAs inside the reactor. The accumulation 296 triggers a sharp pH drop, and this disrupts the stability of AnMBR operations [3, 61]. 297

298

Recovering VFA can actually improve product revenues that eventually improve the overall income earned from the AnMBR, and this in turn improves the energy efficiency of the system. Although different organic components like alcohols, aldehydes, ketones, and organic acids can be synthesized from VFA, it has also been used a potential precursor of biodegradable polymers

and biogas. From a technical perspective recovering VFA helps to maintain a stable pH in the 303 AnMBR especially at high organic loading rates and lower HRTs. Consequently, the capacity of 304 a given AnMBR can be increased when it is designed for VFA and methane recovery. Hence, VFA 305 can be a favorable AnMBR economically as compared to the traditional methane. AnMBRs 306 designed to produce VFA need to apply the selective inhibition of methanogens which are the 307 major VFA consumers during anaerobic digestion. Heat shock and load shock are two common 308 309 methods that can inhibit the activity of the VFA-consuming microbes. Besides selective microbial 310 inhibition, the common process conditions can be optimized to maximize the production of VFA.

311

Generally, an increase in temperature has a positive impact on VFA production. Yuan et al. (2011) 312 carried out an experiment to observe the effect of temperature on VFA production. In this 313 observation three different temperatures (24.6, 14 and 4 °C) were applied to obtain the effects on 314 VFA production from waste activated sludge, and this revealed that the highest temperature 315 resulted in maximum VFA production (2154 mg L^{-1}) [62]. Several other studies have reported an 316 increase in temperature within the psychrophilic (4-20 °C) and mesophilic (20-50 °C) range 317 increases VFA production. This increment is associated with the fact that the rate of hydrolysis in 318 an anaerobic process generally increases at high temperature which results in a higher production 319 rate of VFA [62, 63]. 320

321

Besides temperature, pH affects the overall VFA production in a more complex way. The components present in the overall VFA mixture requires different optimum pH values. For simplicity, the current section will focus on the overall VFA yield, and production rate only.

Research studies have shown that between pH 5.0 and 6.0, the highest overall VFA concentration 325 is observed regardless of the type of which inoculum was used while producing VFA from food 326 waste [64]. However, Khan et al. (2019) reported that the highest VFA yield from low-strength 327 synthetic wastewater recorded 48.74 ± 1.5 mg VFA/ 100 mg COD_{feed} at pH 7.0, whereas Jie et al. 328 (2014) documented at pH 10.0 the accumulation of VFA reached the maximum limit during 329 anaerobic digestion of excess sludge (ES) [61, 65]. As different types of microbial species are 330 present in different inoculums, their optimum growth rates vary at different pH levels. Therefore, 331 332 it is more practical to analyze the microbial species first rather than generalizing an optimum pH level for VFA production. It is evident that operational parameters like HRT and OLR rely on the 333 334 design of the bioreactor. Between these two, optimum HRT can be generalized for VFA production from an anaerobic process. A general decrease in HRT of an anaerobic process results in increased 335 VFA production [66]. Khan et al. (2019) demonstrated that for an AnMBR treating low-strength 336 337 synthetic wastewater, 8 hrs of HRT provide the highest VFA production rate and yield [3]. However, it is also important to consider the membrane fouling, nutrients, and removal efficiency 338 of COD in the AnMBR at low HRTs [9, 24]. 339

340

341 3.3 Two-stage AnMBRs for multiple AnMBR product recovery

Two-stage AnMBRs have an interesting design that separates the initial stage of acidogenesis and final stage of methanogenesis [67, 68]. The produced VFA accumulated inside an AnMBR can inhibit methane production by sharply decreasing the level of pH. As a result, two-stage AnMBR design offers the advantage of producing VFA, biohydrogen and methane simultaneously. In addition to this, extracting multiple AnMBR products simultaneously can improve product revenues earned from an AnMBR, and improve energy efficiency. However, the two-stage design has not yet been popular due to the high cost in initial installation and the high energy requirements
in the process operation [38, 68-70]. Two-stage AnMBR designs have been particularly favorable
for extracting multiple products from the AnMBR. However, much research needs to be
undertaken to compare the increase in product revenues, added costs in installation and operations
for the two-stage design. Table 5 summarizes the features of some two-stage anaerobic bioreactors
that have been employed for multiple product recovery.

354 # insert Table 5

355

The information provided in Table 5 when analyzed provides two different viewpoints. Firstly, 356 there is the lack of research done on how to produce VFA and methane simultaneously from the 357 anaerobic process. Recovering VFA can actually improve the production of methane at the final 358 359 stage of an anaerobic process. At the same time, it reduces potential hazards of process inhibition due to a sudden drop in pH [55]. Secondly, the potential of the two-stage AnMBRs has not been 360 completely discovered for multiple product recovery. The reasons are mainly associated with 361 severe membrane fouling at low HRT, and high OLR, a high initial cost of installation and high 362 energy requirements in the bioreactor operation. 363

364

365 4. Challenges

366

367 Various challenges lie ahead to improve energy recovery from the AnMBR. They mainly include368 controlling process inhibition, storage and transportation of biohydrogen, recovery of dissolved

methane, VFA extraction and purification, and finally the need to reduce costs when operating theAnMBR.

371

372

4.1 Controlling process inhibition

Recovering energy from the AnMBR can be particularly challenging because of potential process inhibitions due to the compounds produced at different stages of anaerobic digestion. The major inhibitors of the anaerobic process include five major categories: ammonia; VFA; organic toxicants; heavy metals; and sulphides. The following sub-section describes the challenge involved in controlling the concentration of inhibitory components during anaerobic digestion.

378

379 4.1.1 Ammonia

Ammonia is one of the most common process inhibitors in anaerobic digestion. AnMBRs treating wastewater and wastewater streams with a high amount of nitrogen can release ammonia nitrogen in the form of NH4⁺. Ammonia can also be present in the AnMBR in the form of free ammonia (NH₃) which is responsible for inhibiting methane production.

384

Different studies have shown a reduction of 50% in methanogenic activity in upflow anaerobic sludge bed (UASB), and expanded granular sludge bed (EGSB) reactors at 2.48 and 2.89 g/L concentrations of ammonia nitrogen, respectively. Free [Free radicals derived from this?] ammonia can rapidly diffuse into the cell membrane and cause proton imbalance whereas ammonium ions can directly inhibit the methane synthesizing enzyme [81].

Controlling ammonia inhibition can be challenging, current approaches include increasing the SRT, dilution, and reducing the bioreactor pH. Few studies have proven that the addition of zeolite along with humic acid can reduce ammonia inhibition in the anaerobic process. Although certain experimental initiatives have proven to be effective, controlling ammonia inhibition still remains a challenge for large-scale operations.

396

397

398 **4.1.2 VFA**

Controlling the accumulation of VFA can be a challenge for single stage AnMBRs. VFAs can be 399 accumulated inside a reactor for many reasons: high OLR, low HRT, and severely fouled 400 membrane layers. High organic loading rate or low HRT can increase the rate of initial hydrolysis 401 in an anaerobic process whereas fouled layer on a membrane can reject the produced VFA inside 402 403 an AnMBR. Of the major VFA components, acetic and butyric acid can be easily consumed by the methanogen. About 60% of the biogas is produced through the conversion of acetic and butyric 404 acid. The conversion of another major VFA component (propanoic acid) is thermodynamically not 405 favorable. Therefore, propanoic acid is mainly responsible for reducing the level of pH in an 406 AnMBR [3, 4]. 407

408

VFAs can be equally damaging for biohydrogen production. They can be responsible for possible deviation of the metabolic pathway from acidogenesis to solventogenesis. In this process, organic solvents such as alcohols and acetone are produced by VFA. The accumulated VFA penetrates the cell membrane, dissociates and increases the H⁺ ion concentration. A pH imbalance caused by high H⁺ concentration can cause cell death and suppression. A combination of these events can inhibit
biohydrogen production [4].

415

416 4.1.3 Organic toxicants

417 The feedstock used in anaerobic digestion sometimes goes through different pre-treatment processes, i.e. heat treatment, alkaline or acidic pre-treatment and these can speed up the hydrolysis 418 419 process. The pre-treatment processes can produce organic compounds that are toxic to the anaerobic process [82]. Organic toxicants such as chlorophenol and halogenated open-chain 420 hydrocarbons can inhibit the overall anaerobic process. Chlorophenols directly disrupt the proton 421 gradient through the membrane and negatively affect the cellular energy transduction. This 422 outcome can in turn can cause process inhibition through a reduction in cell growth. Additionally, 423 open chain halogenated hydrocarbons like chloroform, trichloroethylene, and perchloroethylene 424 425 can also wield an inhibitory effect on methanogenesis. Furan, phenol derivatives such as Furfural, Phenol, and Vanillin also exert an inhibitory effect on the production of biohydrogen [4, 81]. 426

427

428 **4.1.4 Sulphides**

Different industrial effluents, such as those produced by tanneries, pharmaceuticals, and coalbased power plants can produce wastewater streams that are rich in sulfide. The activities of Sulphate Reducing Bacteria (SRB) and methane-producing bacteria are significantly affected by the high sulfide concentration present in the bioreactor feed. In this case, a general decrease in methane production was observed as it competed for the available carbon and hydrogen. Studies have identified that H₂S concentrations of 160 and 220 mg/L can completely inhibit the microbial activity of acetotrophic and hydrogenotrophic methanogens. A possible solution for solving this 436 problem is to incorporate a sulfide removal stage within the AnMBR design. For membrane 437 bioreactors, the biomass is retained inside the reactors for a longer period of time. In this case, the 438 adaptation of methane-producing bacteria to free H₂S can improve the methanogens' tolerance to 439 sulfides [81].

440

441 **4.1.5 Metal ions**

442 Light and heavy metal ions contribute to increase enzymic activity, cell metabolism and finally promote bacterial growth. For instance, iron supports bacterial growth, stimulates enzymic 443 activities, and helps to eliminate sulfide inhibition during biohydrogen production. Yet, an iron 444 concentration above 100 mg/L can have an inhibitory effect on hydrogen production. The 445 inhibitory effect of Mg²⁺ ions has been observed above 20.0 mg/L although it works as a cellular 446 protein builder at smaller concentrations. Two other micronutrients, Na⁺ and Ca²⁺ show an 447 inhibitory effect above 2000 mg/L and 100 mg/L, respectively. Similarly, 1600,15, 3 and 0 mg/L 448 are the threshold limits for Ni, Cr, Cu and Zn ions, respectively. Precipitation, coagulation and 449 sorption methods can be applied to remove or reduce the concentration of different metal ions 450 during anaerobic digestion [4]. 451

452

453 *4.2 Dissolved Methane recovery*

454 Methane is a greenhouse gas and the global warming potential is 34 times greater compared to 455 CO₂ over a 100-year period. Nearly 2.8% of the total GHG emissions is related to anaerobic waste 456 and wastewater treatment processes that occur worldwide. Dissolved methane in the AnMBR 457 effluent is one of the main causes of destruction to the environment. At the same time, it reduces the amount of recoverable energy from AnMBR. At atmospheric pressure [what specific pressure amount are you referring to?] and a temperature of 25 ° C, 21.6 mg methane can be present in each litre of water. The solubility of methane increases when the temperature is decreased. Anaerobic processes operated at low temperatures can lead to 60% of the total methane being dissolved in the bioreactor effluent whereas the percentage varies between 20 to 60% for sewage treatment. Therefore, recovery of dissolved methane not only improves the energy recovery, but it also reduces the negative environmental impacts of the AnMBR [83].

465

466 One common strategy for methane recovery is to integrate a post-treatment process for an 467 anaerobic bioreactor effluent. Aeration, air stripping, catalytic methane oxidation and membrane-468 based separation have been used to remove dissolved methane. Membrane-based removal of 469 dissolved methane from liquid streams is the most effective and commonly used technology of 470 dissolved methane removal. Table 6 lists the common methane removal technologies for dissolved 471 methane removal from municipal wastewater.

472

473

insert Table 6

474

The issues that are involved in dissolved methane recovery are primarily technical and economical. At the same time, dissolved methane can be responsible for creating an explosive atmosphere when the concentration exceeds 0.14 g/L in a closed conduit. Typically, the energy demand for an AnMBR process compared to other competitive anaerobic bioreactors is high. During municipal wastewater treatment, a typical AnMBR can have an energy demand ranging between 0.03–16.52 480 kWh/m³ compared to 0.133–0.227 kWh/m³ of an AFBR- AFMBR system. The reasons for this are
481 linked to biogas spurging and the degree of supersaturation.

482

A novel process was designed using a down-flow hanging sponge in anaerobic domestic 483 wastewater treatment. Methane Oxidizing Bacteria (MOB) was used in this process to remove 484 dissolved methane from the effluent through oxidation. The highest methane removal rate was 485 2.2 kg-COD m^{-3} day⁻¹ but the study did not include the effect on sulfide and organic 486 carbon oxidation. A follow up study showed that above 90% removal rate for COD, sulfide, 487 methane, and ammonium can be achieved by integrating down-flow hanging sponge unit installed 488 in a UASB reactor. Although the findings show the technical feasibility of methane removal, it 489 does not provide any estimation of energy requirement in full-scale application [95]. 490

491

492 Recovery of methane-using membranes involves different technical issues like the efficiency of recovery, limitation in mass transfer and above all the energy required for the recovery process. 493 At the same time, controlling membrane fouling can also be a technical issue. Membrane modules 494 operated at a lower temperature can encounter higher resistance if and when the liquid viscosity 495 increases and diffusivity decreases. The energy efficiency of this recovery process becomes more 496 complex for AnMBRs operated below mesophilic and thermophilic conditions. At low 497 498 temperature, the solubility of methane in water increases and this in turn increases the energy requirement for methane recovery. Therefore, from the perspective of improving energy 499 efficiency, methane recovery is not a feasible option at low temperatures. Optimization of process 500 conditions is another area for future research to improve the overall energy requirement in a 501 502 methane recovery process.

504 4.3 Storage and transportation of biohydrogen

The challenges involved in biohydrogen storage directly affect the initiatives for energy recovery 505 from AnMBR and made the energy recovery through hydrogen production less feasible for 506 507 industrial application. Storage of biohydrogen poses a greater challenge as it has wider 508 flammability (4 -75%) and explosion limit in air (13.0–59.0%) compared to methane. Although 509 different types of hydrogen storage systems are currently in operation in the industry, the type of 510 storage depends largely on the type of use. For automobile applications, gravimetric density is the main design consideration for hydrogen storage. However, for hydrogen transportation, process 511 safety is the primary area of concern along with the density of hydrogen. The currently used 512 hydrogen storage system suffers from different technical and safety issues. They include: low 513 gravimetric density, evaporative loss, boiling-off phenomenon, hydrogen embrittlement, etc. The 514 515 following sub-section details the challenges involved in different hydrogen storage systems.

516

517

518 4.3.1 Challenges in physical storage

The physical storage of biohydrogen mainly includes storage in two forms, compressed gas and cryogenic liquid hydrogen. Recently developed technologies in physical storage also include adsorption on solid materials such as carbon nanostructure, Borophene, etc., and solid storage. The most common physical storage involves pressurization up to 700 bars. Low storage density is a major problem of this method. At 350 and 700 bars the density can be only up to 5.5 and 4.6% (w/v) which is very low considering automobile applications. Recently developed storage material

based on carbon fiber can store 6-10 wt% H₂ at 350–700 bars but the energy needed for compression and volume reduction still remains a burning issue for this process [4, 96].

527

Storing hydrogen as cryogenic liquid at low pressure eliminates the issue of low gravimetric density; the density can be up to 20% wt at 1 bar and -253°C during this process. At the same time, storing hydrogen as liquid introduces a new problem – evaporative loss. Reducing the cost involved in the liquefaction process is also a major challenge in this process.

532

533 Different porous solid materials can accommodate molecular hydrogen in molecular state.

Clathrate compounds such as hydrates, fullerenes, graphene, etc., have served for the physical storage of hydrogen. The storage capacity of each component depends on the porous structure, available surface area and stability. Metal-organic frameworks (MOFs), CMPs (conjugated microporous polymers), and HCPs (hyper-cross-linked polymers) have been extensively studied in this process. Subsequently, the results have shown that the applicability of each type is limited at a specific temperature. Research studies need to be conducted at different temperatures to improve the application range of different adsorption components.

541

542 **4.3.2** Challenges in chemical storage

The most recent studies have shown that metal hydrides can offer high storage capacity of hydrogen and can release it at different temperatures and pressures. AB type alloys show high abruption/desorption capacity usually containing high hydride equilibrium pressure. For example, TiFe alloys have high hydride equilibrium pressure and low gravimetric storage capacity. To resolve this issue, hydrides of sodium and lithium have been employed but using sodium andlithium still remains expensive and thermodynamically unstable.

549

550 Metal borohydrides can be a possible solution in this case where both stability and high storage 551 capacity could be problematic. For example, LiBH4 shows a hydrogen storage capacity of 552 18.5 wt% when the temperature for decomposition is 673K. When a catalyst (SiO₂) was added to 553 reduce the decomposition temperature, the overall storage capacity was reduced to 13.5 wt%. 554 Consequently, it is particularly challenging to design a borohydride alloy that offers both high 555 storage capacity and low temperature for decomposition.

556

Organic compounds such as Liquid Organic Heterocycles (LOH), hydrazines, organic acids, and alcohols have been employed for hydrogen storage due to not much energy being required for hydrogenation. To improve hydrogen storage capacity, solid catalysts have been used in this case. Silver nanoparticles with pd coating have been used to decompose the stored hydrogen in formic acid. Ionic liquids such as methyl-guanidinium borohydride can contribute to increasing hydrogen release when added with chemical hydrides. However, they have also demonstrated synergistic effects [96].

564

Hydrogen can be stored electrochemically where it is adsorbed on substances in aqueous solution.
The effectiveness of a particular material in this process depends on the discharge rate of hydrogen and the storage capacity. In some cases, life cycle behavior is important because the material loses the adsorption capacity after a certain number of adsorption/desorption cycles. In this scenario, the

properties of TiNi have been studied and the results showed storage capacity of 230 mAhg⁻¹ but the rate of hydrogen discharge was very slow. AB₃ type alloys such as LaNi₃, CaNi₃, etc., have been investigated to increase the storage capacity of hydrogen through the electrochemical process. However, improving the capacity again triggers a drop in the recyclability of this storage system. Much more research needs to be done on discovering the intermetallic properties of these alloys.

574

575 The challenges involved in chemical and electrochemical storage of biohydrogen are many. The catalysts synthesized for hydrogen storage in formic acid show improved performance at a certain 576 temperature and pressure. New research initiatives are needed to synthesize new catalysts for 577 biohydrogen storage in organic liquids. Improved rates of hydrogenation and dehydrogenation at 578 different temperatures and pressures are another concern in LOHCs. Most importantly, the 579 economic aspects of hydrogen storage through different chemical and electrochemical processes 580 have been neglected. This is potentially a serious issue when the storage system is subjected to 581 full-scale industrial application. 582

583

584 4.4 Additional challenges

Separation and purification of VFA from different anaerobic processes have not yet been assessed for their economic feasibility. Although currently available research shows the recovery of VFA can be technically feasible, the cost involved in full-scale operation has not been addressed. Currently extracting VFA using organic solvents is the most common separation process for labscale operations. Reducing the cost of organic solvents for VFA recovery can be very challenging in the VFA recovery process. Reduction in operating and maintenance costs is one of the most widely researched areas in AnMBR technology. A typical AnMBR requires a lot of energy for gas
spurging, sludge feeder, stirring, permeate pump and membrane tank feeding pump. Additionally,
membrane fouling control, membrane cleaning and membrane replacement costs are essential
maintenance aspects of AnMBR operations.

595

Over the past few years, there have been significant improvements in membrane fouling control 596 597 and membrane fabrication processes. The costs involved in membrane fabrication have been 598 reduced and different physical and chemical processes for membrane fouling control have been applied. However, only a few studies have characterized the problem of membrane fouling when 599 600 AnMBR is being operated to produce alternative AnMBR products like VFA and biohydrogen. Recently developed membrane modules have a higher surface area, low energy requirement and 601 are more resistant to membrane fouling. For instance, a recently developed membrane module by 602 603 Pentair has 40% more surface area and requires 35% less energy during cross-low operating mode [97]. AnMBR systems that integrates Microbial Fuel Cells (MFCs) and Microbial Electrolysis 604 Cells (MECs) have shown that the voltage generated can be effective in controlling the fouling 605 through a reduction in foulants' deposition. However, the lab-scale successes have to be applied 606 607 in pilot/full-scale operations so that we know what the challenges are when attempting industrial applications. 608

609

Pretel et al. (2014) [98] investigated the energy requirement of an AnMBR treating sulphate-rich urban wastewater. According to their study, biogas spurging, membrane tank sludge feeding pump, feeding and permeate pumps are the additional energy consumers in AnMBR operation. The minimum energy requirement was 0.07 kW h/m3[•] A follow-up study [99] on this topic calculated the minimum energy requirement for a system treating 50,000m³/d. The results showed the minimum energy was in fact 0.04 kWh/m³ which is lower compared to their previous study. The cost reduction opportunities still need to be explored for AnMBR treating different wastewater streams. Challenges lie ahead to assess the energy consumption when product type, scale of operation, and operating conditions are changed in an AnMBR.

619

620 **5.** Conclusions and Perspectives

621

The energy recovery options from AnMBR have not yet been completely discovered. Although the currently available options mainly focus on methane, recovering VFA and biohydrogen from AnMBR can actually improve the energy efficiency of this system. Challenges like controlling process inhibition, recovering dissolved methane, storage and transportation of biohydrogen, and a reduction in operating and maintenance cost have to be resolved before the full-scale application of AnMBR.

628

629

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Feed wastewater	Bioreactor configuration, membrane type	COD removal rate (%)	Energy Production (Methane) (L/kg COD)	Methane Conversion rate (%)	Reference
Municipal Wastewater	UASB, HFM PVDF	91	160.5 ± 5.6	45.3	[12]
Synthetic Municipal wastewater	CSTR, PVDF	90-96	338	96.5	[13]
Kraft evaporator condensate	UASB, submerged MF	97–99	290–310	-	[14]
Landfill leachate	Cross-flow UF MBR	90	460	-	[15]
Real municipal	CSTR, ceramic (Al ₂ O ₃) membranes	86–88	300	-	[16]
Meat packing wastewater	AnMBR, hollow-fibre UF	88–98	130–180	-	[17]
Synthetic wastewater	CSTR, FSM PVDF	94.3–95.5	326	93%	[18]
Domestic and food waste-recycling wastewater	AnCMBR, N/A	97.9–99.3	200–220	-	[19]
Synthetic wastewater	Semi-continuous SAnMBR, flat sheet PVDF	>98	290	-	[20]
Synthetic wastewater	Granular AnMBR, PVDF HFM	$93.7\pm1.7\%$	156.3 ± 5.8	-	[21]

Table 1: Current Status of energy production in AnMBR [10, 11]

949 UASB - Upflow Anaerobic Sludge Blanket; HFM – Hollow Fibre Membrane; PVDF - Polyvinylidene

950 Difluoride; MF – Microfiltration; UF - Ultrafiltration; CSTR - Continuous Stirred Tank Reactor; AnCMBR

951 - Anaerobic Ceramic Membrane Bioreactor; SAnMBR – Submerged anaerobic Membrane Bioreactor.

Substrate	AnMBR type, membrane, surface area	Operating Condition(s)	Highest biohydrogen yield	Maximum production rate	Reference
Tofu	CSTR,	HRT	42.4 L	19.8 L	[29]
processing	hollow fibre,	2–8 h	H ₂ /mol	$H_2/L-d$	
waste	0.025 m^2	$pH \; 5.5 \pm 0.1$	hexose		
		T 60 °C \pm 0.1			
Glucose	H ₂ producing	23.0 ± 0.1 °C pH	-	4.4 L H ₂ /L-	[30]
	AnMBR,	$5.5\pm0.1.~HRT$ - $8~h$		d	
	hollow-fibre,	SRT – 24 d			
	0.047 m^2				
Glucose	SAnMBR,	35 ± 0.5 °C	27 L H ₂ /mol	5.8 L H ₂ /L-	[31]
	Plate-flame	HRT - 9h	glucose	d	
	type, 0.1 m^2	SRT – 2–90 d			
Glucose	H ₂ producing	HRT 8h	40.2 L	4.5 L H ₂ /L-	[32]
	AnMBR,	SRT24h	H ₂ /mol	d	
	hollow-fibre	23 ± 1 °C	glucose		
		5.5 ± 0.1			
Glucose	SAnMBR,	SRT 90 d		2.5 L H ₂ /L-	[33]
	plate flame (flat	35 ± 0.5 °C	19.5 L	d	
	sheet) type,	5.5 ± 0.05	H ₂ /mol		
	0.1 m^2		glucose		
Glucose	AnMBR with	SRT 12.5 h	35.4 L	5.9 L H2/L-	[34]
	magnetic	HRT 9h	H ₂ /mol	d	
	stirring,	35 ± 0.5 °C	glucose		
	microfiltration,	$pH \; 5.5 \pm 0.1$			
	0.1 m ²				
Glucose	UASB, tubular	$35 \pm 1 \ ^{\circ}\text{C}$	-	$475 \ mL \ H_2$	[35]
	ultrafiltration,	4.5 ± 0.1		L-1 h-1	
	0.04 m^2	HRT of 4 h			
Glucose	External	37 °С.	2.98 mol	51.38 L	[36]
	dynamic MBR	HRT 12,6,3, and 2 h	H ₂ /mol	$H_2/L-d$	
		рН 7.0	glucose		
Cheese	Ceramic-	SRT 50 days	3087.57	-	[37]
whey	alumina	20 °C	mL/g bacteria		
	membrane,		(R. aquatilis)		
	55 cm^2				

953 Table 2: Production of biohydrogen from AnMBR

Property	Methane	Hydrogen
Energy Density (MJ Kg ⁻¹)	143	55.6
Specific heat capacity (Jkg ⁻¹ K ⁻¹)	2.22	14.89
Flammable limit (% v/v in air)	5.3 - 15.0	4.0-75.0
Density (Kg/m ³)	0.65	0.08
Stoichiometric air/fuel ratio (kg/kg)	17.1	34.2
Temperature of flame (K)	2148	2318
Explosion limit (% v/v in air)	6.3–14.0	13.0–59.0

Table 3: Comparison of different fuel properties between methane and hydrogen [40, 41]

Bioreactor	Substrate	Operating conditions	Highest biohydrogen yield	Reference
Continuous Stirred-tank Reactor	Starch	37°С pH - 5.5	2.3 mol H ₂ / mol hexose	[42]
Stirred-tank Reactor	Food waste	35±1°C pH - 6.0	1.73 mol H ₂ / molhexose	[43]
Batch Fermenter	Distillery wastewater	37°С pH -5.5	1000 ml H ₂ /L medium	[44]
Continuous Stirred-tank Reactor	Anaerobic sludge	70°C pH – (7.0-8.0)	1.11 mol H ₂ /mol-hexose	[45]
Batch Fermenter	Activated sludge	55°C pH - 7.0	1.25 mol H ₂ /mol glucose	[46]
Upflow anaerobic sludge blanket	Glucose	37°С pH - 6.5	$1.44 \pm 0.1 \text{ mol } \text{H}_2/\text{mol-hexose}$	[47]
Fluidized bed reactor	Municipal sewage	40°C	4.26 mol H ₂ /mol sucrose	[48]
Upflow anaerobic sludge blanket digestion	Activated sludge	35±1°C	1.7 mol H ₂ /mol-hexose	[49]
Batch Fermenter	Corn stalk	60°C	89.3 mL H ₂ / g dry biomass	[50]
Continuous Stirred-tank Reactor	Glucose	37°С pH - 5	1.3 mol H ₂ /mol glucose	[51]
Continuous Stirred-tank Reactor	Tofu- processing waste	60°C pH - 5.5	2.3 mol H ₂ /mol glucose	[52]
Continuous Stirred-tank Reactor	Cow dung compost	33.5°C pH - 5	2.15 mol H ₂ /mol glucose	[53]
Trickle Bed Sequenced Batch Reactor	Municipal wastes	pH – (4.65– 5.87)	1.67 mol H ₂ /mol glucose	[54]

957Table 4: Different anaerobic process for production of biohydrogen

Bioreactor type, design	Feedstock	Operating conditions	Hydrogen/ VFA yield	Methane yield	Reference
Two-stage	Palm oil mill	55 °C	H ₂ :	342 L/Kg-	[71]
thermophilic	effluent	HRT: H ₂ - 2 days;	73 L/Kg-VS	VS	
anaerobic bioreactor		CH ₄ -10 days			
Two-stage	landfill	pH: VFA- 5.5 , 11	VFA: 48 g/L	6 g/L	[72]
anaerobic digester	leachate	$CH_4 - N/A$	leachate	leachate	
		T: 37 ± 2 °C			
Two-stage	Food waste	$H_2-55 \pm 2 \ ^{\circ}C; CH_4-37 \pm 2$	115.2 ± 5.3	334.7 ± 18.6	[73]
anaerobic CSTR		°C	L/Kg-VS	L/Kg-VS	
$(H_2) + AFBR (CH_4)$		HRT: H ₂ - 3.5 days; CH ₄ -			
		1.5 days			
Two-stage induced	Dairy	H ₂ - 60 °C;	160.7 ml/g-	168.8–	[55]
bed reactors (IBR)	processing	$CH_4 - 40$ °C	COD	178.1 ml	
	waste	HRT: $H_2 - 3$ days;		CH4 ml/g-	
		CH ₄ - 15 days		COD	
Two-stage	Ethanol	35 °C	48	344 L/Kg-	[74]
anaerobic CSTR	stillage, cake	HRT: H ₂ -3 days;	L/Kg-VS	VS	
	and glycerol	CH ₄ -12 days			
	waste				
Two-phase	Bio-waste	T - 55°C	51 L/Kg-VS	780 L/Kg-	[75]
thermophilic CSTR		HRT - 32 days		VS	
Two-stage	Wheat bran	$37 \pm 1^{\circ}C$	18.9 L/Kg-	243.5 L/Kg-	[76]
anaerobic batch		HRT: $H_2 - 4$ days;	VS	VS	
process		CH ₄ - 40 days			
Two-stage	Corn silage	37 °C	59.4 ± 4.1	328.8 ± 16.8	[77]
anaerobic leach bed	and cattail	HRT - 60 days	L/Kg-VS	L/Kg-VS	
+ UASB					
Two-stage	Grass silage	H ₂ -35 °C;	6.46 L/Kg-	467 L/Kg-	[78]
anaerobic batch		CH ₄ - 55 °C	VS	VS	
process		HRT: H ₂ - 6 h;			
		CH ₄ - 6 days			
Two-stage pilot	Sargassum sp.	55 °C	66.7 L/Kg-	720 L/Kg-	[79]
scale CSTR		HRT: H ₂ - 3.3 days; CH ₄ -	VS	VS	
		12.6 days			
Two-stage anaerobic	Vinasse	HRT:	14.8 l/kg VS	274 ml/g	[80]
digestion		H ₂ - 2 d H ₂ ;	substrate	VS substrate	
		CH ₄ - 15 days			

959 Table 5: Two-stage anaerobic bioreactors for multiple product recovery

Bioreactor type	Operating Temperature (°C	Scale	CH ₄ lost in effluent (%)	CH ₄ recovery process, (%) CH ₄ recovered	Reference	
EGSB	16	Pilot	45	Sweep gas desorption, 72%	[84]	
UASB	35	Bench	12	Degassing membrane 89% - 97%	[85]	
UASB	25	Pilot	41	-	[86]	
UASB	21–28	Pilot	70	Down-flow hanging sponge 76.8%	[87]	
UASB	25	Bench	45	Desorption by membrane contactor 98.9%	[88]	
AFBR- AFMBR	25	Bench	63	N/A	[89]	
AnMBR	15	Bench	40–50	N/A	[90]	
AnMBR	22	Pilot	19	N/A	[91]	
SAnMBR	33	Pilot	43	Biogas assisted Stripping 57.4%	[92]	
AnMBR	25	Bench	88	Desorption by membrane contactor 98.9%	[88]	
EGSB	25	Bench	N/A	polypropylene membrane contactor 98%	[93]	
Anaerobic digester	39.4	Bench	N/A	Adsorption by zeolite 56.82 ± 3.09	[94]	

962 Table 6. Estimation of dissolved methane from different anaerobic bioreactors treating municipal963 wastewater

964 EGSB - Expanded Granular Sludge Bed; AFBR - Anaerobic Fluidized Bed *Reactor*; AFMBR - Anaerobic Fluidized

965 Membrane Bioreactor