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1 **Aliphatic polyketone-based thin film composite membrane with mussel-inspired polydopamine**
2 **intermediate layer for high performance osmotic power generation**

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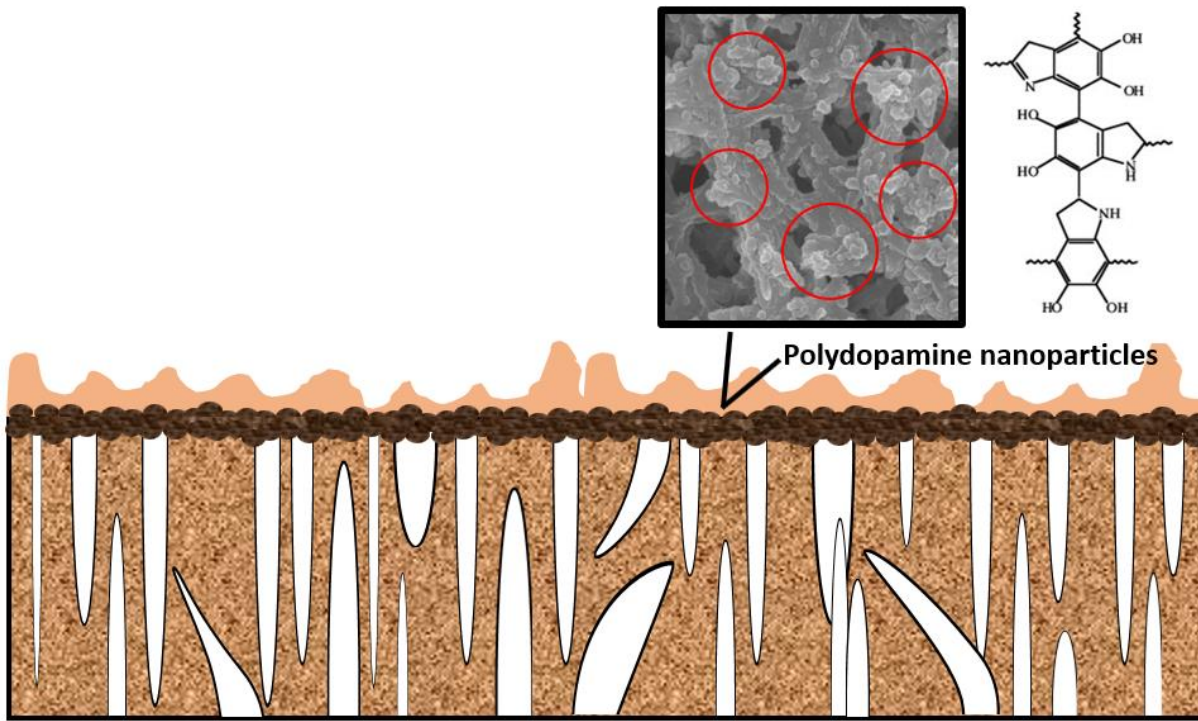
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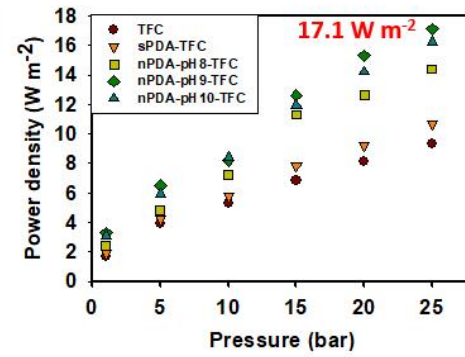
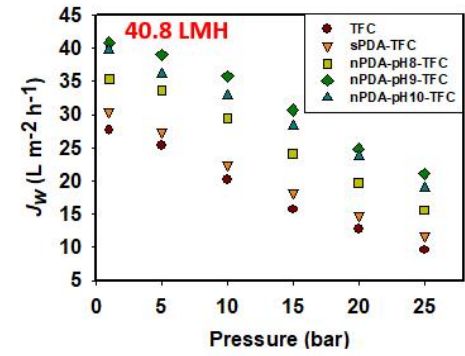
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Thin film composite PRO membrane with nano-sized polydopamine particulate interlayer



13 **Abstract**

14 Polydopamine (PDA), formed from self-polymerization of dopamine, was coated on aliphatic polyketone
15 membrane substrate prior to interfacial polymerization (IP), preparing a pressure retarded osmosis (PRO)
16 thin film composite (TFC) membrane with a PDA interlayer. The effect of the formation of two types of
17 PDA interlayers — smooth and particulate — on substrate morphology, polyamide formation, and PRO
18 osmotic performance were investigated. Also, the effect of pH on the particulate PDA interlayer was studied.
19 It was found that the introduction of both smooth and particulate PDA contributes to enhanced water flux
20 and power density of the PRO membranes. pH was found to have significantly affected the formation of
21 particulate PDA and the polyamide formation, as well. At higher pH, PDA self-polymerization led to the
22 formation of more nanoparticles, the subsequent increase in surface roughness and decline in the polyketone
23 substrate porosity. The particulate PDA interlayer formed looser polyamide, compared to the thinner and
24 denser polyamide formed on pristine and smooth PDA-interlayer-coated TFC membranes. The membrane
25 performance was evaluated using deionized water and 1.0 M NaCl as feed and draw solutions, respectively.
26 The TFC membrane with nanoparticulate PDA layer formed at pH 9.0 exhibited the best initial water flux
27 of $40.8 \text{ L m}^{-2} \text{ h}^{-1}$, and this membrane also showed the highest power density of 17.1 W m^{-2} at 25 bar. The
28 results of this study indicate that nanoparticulate PDA interlayer formation is a simple and scalable TFC
29 membrane development method for engineered osmosis.

30

31 **Keywords:** Pressure retarded osmosis; Thin film composite membrane; Polydopamine; Osmotic power
32 generation; Interlayer

33 1. Introduction

34 Currently, alternative sources of energy are needed to augment the world's continuously increasing
35 energy requirements due to increased population and advanced economic development [1]. Until present,
36 most of the earth's energy is obtained from vastly depleting fossil fuel reserves, which also causes
37 greenhouse gas emissions giving rise to global warming and climate change. As a result of this energy crisis,
38 scientific research from all over the world has looked into the following more sustainable energy sources:
39 solar, wind, geothermal, biomass, and osmotic power [1-3].

40 Generation of osmotic power by means of pressure retarded osmosis (PRO) is achieved due to the
41 natural occurrence of osmosis between two solutions of varying salinity levels and osmotic pressures with
42 a semi-permeable membrane separating each solution [4, 5]. The difference in osmotic pressure allows
43 water permeation from the feed solution (FS), whose concentration is lower, toward the pressurized higher
44 concentrated draw solution (DS). Osmotic power can then afterwards be harvested by delivering the
45 pressurized DS to a hydro turbine, which produces electrical energy [6].

46 Alongside the development of PRO as a commercially-viable and technologically-feasible process is
47 the manufacture of membranes specifically tailored for PRO application. Recently, research on PRO
48 membrane development has shown a tangible progress, especially for the manufacture of thin film
49 composite (TFC) membranes with high power density.

50 TFC membranes are consisted of a membrane substrate with high porosity and a dense thin film
51 selective layer, typically polyamide, formed atop the porous substrate through interfacial polymerization
52 (IP). In the previous studies, efforts were made to engineer TFC PRO membranes using a variety of methods
53 [7, 8], modifying agents [9-11], and nanomaterial fillers [12, 13]. These techniques were aimed at
54 improving the water permeability, selectivity, and mechanical strength of the membranes, in order to
55 improve overall osmotic power generation performances.

56 Among the chemical agents used in modifying PRO TFC membranes is polydopamine (PDA) [14].
57 This polymer is formed from the self-polymerization of dopamine (3,4-dihydroxyphenylalanine) in alkaline
58 conditions. The properties of PDA are known to be similar to the phenolic protein that are found in mussels,
59 causing PDA to be called a bio-inspired or mussel-inspired material. PDA is mainly used for coating a wide
60 range of materials, from nanomaterials to huge surfaces, and it is also widely used as an adhesive. PDA is
61 highly hydrophilic, owing to the presence of hydrophilic hydroxyl and amine groups. Therefore, this
62 polymer has been used for modification of membranes utilized in water-based technologies to enhance
63 separation performance and even fouling resistance [15, 16]. PDA has become a versatile and efficient
64 engineering agent in membrane development, and it can be applied on membranes using any of the
65 following techniques: (a) surface deposition [17], (b) addition in polymer dope solution prior to casting
66 [18], (c) covalent linkage with other molecules by exploitation of Michael or Schiff base reactions [19],
67 and (d) intermediate layer formation [20]. PDA has been widely used to facilitate the formation of
68 intermediate layer and reinforce the incorporation of amphoteric substances [21] and nano-sized inorganic
69 fillers, such as graphene oxide [22], silica [23], taurine-modified hydroxyapatite [24], halloysite nanotubes
70 [25], and covalent organic framework [26].

71 Recent studies have revealed the formation of nano-sized PDA particles when, instead of Tris-HCl
72 buffer, ammonia is used to initiate the dopamine self-polymerization [20]. This method is different from
73 the conventional PDA modification which uses an aqueous PDA and Tris-HCl buffer solution on the surface
74 to form a smooth film. In this study, both smooth and particulate PDA interlayers were formed between an
75 aliphatic polyketone substrate and the selective polyamide layer. Furthermore, it has been long established
76 that the catechol and amine groups present in PDA can react with the acyl chloride precursor during IP.
77 The introduction of a PDA interlayer between the membrane substrate and selective layer has been found
78 to facilitate excellent formation of mechanically-strong thin film, as caused by the adhesive properties of
79 PDA.

80 The introduction of nanomaterials in TFC membranes for PRO has led to a new class of membranes
81 known as thin film nanocomposite (TFN) membranes. PRO TFN membranes have been developed by
82 incorporating nano-sized materials, like graphene oxide [27], covalent organic framework [12], porous
83 polymer [13], carbon quantum dots [28, 29], and carbon nanotubes [30], either in the membrane substrate
84 or the polyamide selective layers. TFN membranes have exhibited enhanced water permeability, mainly
85 caused by the presence of the highly porous and hydrophilic nanomaterials. Nanomaterial incorporation in
86 the intermediate layer between the substrate and selective layers of PRO TFC membranes has not yet fully
87 been explored, and this study aims to exploit nano-sized particulate PDA as an intermediate media for PRO
88 TFC membranes.

89 This study aimed to discover whether the formation of particulate PDA interlayer could retain the
90 high permselectivity of the pristine TFC and TFC membrane with smooth PDA interlayer, and how the
91 interlayer structure could affect polyamide formation and the subsequent PRO operation performance.
92 Furthermore, ammonia-initiated polymerization of dopamine was also investigated in terms of varying
93 alkaline pH. While research on TFC membrane modification using PDA has been abundant, this manuscript
94 provides insight on how changes in polymerization agent and alkalinity could influence a technique as
95 simple as PDA modification and the PRO performance, as well.

96

97 **2. Experimental**

98 **2.1. Chemicals**

99 The membrane substrate was prepared using aliphatic polyketone (PK, $M_w = 200000 \text{ g mol}^{-1}$, Asahi
100 Kasei Co., Japan) and polyester (PET) nonwoven fabric (90HP, Awa Seishi Co., Japan). To prepare the PK
101 polymer dope solution, resorcinol (> 99.0%) and deionized (DI) water were used as solvents. Acetone (>
102 99.8%) and hexane (> 99.8%) were used during flat sheet membrane casting. Membrane modification was
103 conducted using dopamine hydrochloride (dopamine HCl), Tris(hydroxymethyl) aminomethane (Tris),

104 hydrochloric acid (HCl, 1 mol L⁻¹), ammonia (25.0~27.9%, in solution), and ethanol (99.5%). *m*-
105 phenylenediamine (MPD), trimesoyl chloride (TMC, > 98.0 %), (±)-10-camphorsulfonic acid (CSA, >
106 98.0 %), sodium dodecyl sulfate (SDS), triethylamine (TEA, > 99.0 %), and heptane (> 99.3 %) were used
107 for interfacial polymerization. Sodium chloride (NaCl) was used for membrane osmotic performance
108 testing. Resorcinol, TMC, and CSA were supplied by Tokyo Chemical Industry Co., Ltd., Japan. Dopamine
109 HCl and Tris were supplied by Sigma-Aldrich, Japan. Lastly, acetone, hexane, HCl, ammonia, ethanol,
110 MPD, TEA, SDS, heptane, and NaCl were supplied by Fujifilm Wako Pure Chemical Corp., Japan.

111 All the reagents used in this study were used without further treatment.

112

113 **2.2. Membrane preparation**

114 **2.2.1. Membrane casting and polydopamine interlayer formation**

115 Flat sheet PK membrane substrate was fabricated by conventional nonsolvent-induced phase
116 separation (NIPS). First, a 10 wt% PK solution was prepared by dissolution of the polymer into a
117 resorcinol/DI water (65:35) solvent mixture at 80°C with constant stirring. Ahead of casting, the polymer
118 solution was degassed at 50°C overnight. The solution was poured onto a glass plate and was spread with
119 a casting thickness of 400 μm using a stainless-steel casting knife. The PK film was afterwards immersed
120 in a 35% aqueous methanol coagulation bath for 20 min for phase separation to occur. The membrane
121 substrate was then placed in steel frames to avoid shrinkage before successive 20 min immersion cycles in
122 acetone and hexane baths. The membrane substrate was air-dried afterwards until further use.

123 A polydopamine (PDA) interlayer was formed *in situ* on the top side of the PK membrane substrate.
124 Two types of PDA interlayer were formed: the conventional smooth PDA and the nano-sized particulate
125 PDA, afterwards denoted as PK-sPDA and PK-nPDA, respectively.

126 The typical smooth PDA layer formation method required 2 mg mL^{-1} of dopamine HCl dissolved in
127 a 0.01 M Tris-HCl solution (pH 8.5). The membrane substrate surface was immersed in the solution for 8
128 h while agitating at 400 rpm using a shaker (Eyela, Japan) in room temperature [31].

129 For the formation of particulate PDA interlayer, a 2 mg mL^{-1} dopamine HCl was dissolved in an
130 aqueous 30% (v/v) ethanol solution [20]. The pH of the solution was adjusted to 8,9, and 10 by adding 25%
131 ammonia solution dropwise, corresponding to PK-nPDA-pH8, PK-nPDA-pH9, and PK-nPDA-pH10,
132 respectively. The membrane substrate surface was immersed in the solution for 8 h at light agitating
133 conditions.

134 All PDA-coated membrane substrates were washed with copious DI water prior to interfacial
135 polymerization.

136

137 ***2.2.2. In situ selective polyamide layer formation via interfacial polymerization***

138 The selective polyamide thin film layer was constructed *in situ* on top of the PDA-coated PK
139 substrate by interfacial polymerization (IP) reaction between MPD and TMC. The membrane substrate was
140 initially submerged for 5 min in an aqueous 2 wt% MPD solution, which also contains the following: 2.3
141 wt% CSA, 1.1 wt% TEA, and 0.15 wt% SDS. The excess solution was drained and completely eliminated
142 using an air knife, prior to 2 min immersion of the membrane substrate in 0.15 wt% TMC in hexane solution.
143 The membrane was then cured at 90°C for 10 min. The TFC membrane was afterwards washed with and
144 stored in DI water.

145

146 ***2.3. Determination of intrinsic transport parameters and evaluation of membrane osmotic performance***

147 The membrane intrinsic transport properties, pure water permeability constant ($A, \text{L m}^{-2} \text{ h}^{-1} \text{ bar}^{-1}$) and
148 solute permeability constant ($B, \text{L m}^{-2} \text{ h}^{-1}$), were determined using a cross-flow reverse osmosis (RO) system

149 [32] whose membrane cell has an effective membrane area (A_m) of 8.07 cm², under a pressure of 10 bar, a
150 flow rate of 150 mL min⁻¹, and temperature of 25°C. DI water and 1000 mg L⁻¹ NaCl were used as the feed
151 for determination of A and B , respectively.

152 The TFC membranes were evaluated for PRO performance using a bench-scale system [33]. DI water
153 and 1.0 M NaCl were respectively used as the FS and DS. The system consists of a stainless-steel membrane
154 cell (Nitto Denko C-70F), whose effective membrane area is 38.15 cm². The cell was equipped with a tricot
155 spacer to facilitate flow of the FS and a metal plate with hole openings was positioned to lessen the contact
156 of the membrane with the cell wall. The FS was delivered with a flow rate of 100 mL min⁻¹ using a plunger-
157 style pump system (NP-KX-840, Nihon Seimitsu Kagaku, Japan), while the DS was delivered and
158 pressurized using a high-pressure diaphragm pump (Hydra-Cell, Wanner Engineering Inc., Japan) with a
159 flow rate of 600 mL min⁻¹. The changes in mass and conductivity of the FS were monitored during the PRO
160 operation.

161 Water flux (J_w , L m⁻² h⁻¹) and the A value can be calculated using the following:

$$162 \quad J_w = \frac{\Delta V}{A_m \Delta t} \quad (1)$$

$$163 \quad A = \frac{J_w}{\Delta P} \quad (2)$$

164 where ΔV , A_m , Δt , and ΔP are the permeate volume (L), membrane area (m²), operation time (h), and
165 difference in transmembrane pressure (bar), respectively [34]. Using changes in volume and conductivity,
166 the solute rejection (R) and B value can be calculated using the following equations:

$$167 \quad R = \left(1 - \frac{C_p}{C_f}\right) \times 100\% \quad (3)$$

$$168 \quad B = \left(\frac{1-R}{R}\right)(\Delta P - \Delta\pi)A \quad (4)$$

169 where C_f , C_p , and $\Delta\pi$ are the salt concentration of the feed, salt concentration of the permeate, and
170 transmembrane osmotic pressure difference, respectively. Reverse salt flux (J_s , g m⁻² h⁻¹) is determined
171 using the concentration changes (C_f) and feed solution volume (V_f) at time t , according to the following
172 equation:

173
$$J_s = \frac{\Delta(C_f V_f)}{A_m \Delta t} \quad (5)$$

174 Using J_w and the hydraulic pressure applied ΔP , the power density W can be calculated using this equation:

175
$$W = J_w \Delta P \quad (6)$$

176 The membrane structure parameter, S , can be calculated from the following equation:

177
$$J_w = \frac{D}{S} \ln \frac{A \times \pi_D + B}{A \times \pi_F + J_w + B} \quad (7)$$

178 where π_D and π_F are the respective osmotic pressures of the draw and feed solutions, and D is the solute
179 coefficient [4, 12].

180

181 **2.4. Membrane characterization**

182 Membrane surface and cross-sectional structure was imaged and characterized using field-emission
183 scanning electron microscopy (FE-SEM, JSF-7500F, JEOL, Japan). Prior to imaging, the samples were
184 dried overnight using a freezer drier (FD-1000, EYELA, Japan), and were sputter-coated with osmium with
185 10 nm thickness. Measurements were done in triplicate. Surface pore size was measured from the FE-SEM
186 images using ImageJ image processing software (National Institute of Health, USA).

187 Surface chemistry was characterized using Fourier transform infrared spectroscope (FTIR, Nicolet
188 iS5, Thermo Fisher Scientific, Japan) with an attenuated total reflectance (ATR) detector (iD5, Thermo
189 Fisher Scientific, Japan), and X-ray photoelectron spectroscopy (XPS, JPS-9010 MC, JEOL, Japan) using
190 Al K α X-rays.

191 The bulk porosity of the membrane substrates was measured using gravimetric method; the support
192 was initially soaked in ethanol for 12 h [35]. A capillary flow porometer (CFP-1500AXLC, Porous
193 Materials Inc., USA) was used to determine the membrane pore size. The membrane was initially soaked
194 in a wetting liquid, Galwick[®] prior to porosity measurement.

195 The membrane hydrophilicity was evaluated by an optical contact angle instrument (Drop Master
196 300, Kyowa Interface Science Co., Japan) [36]. Water static contact angle measurements were obtained

197 after dispensing a 4 μL water droplet onto the membrane sample. Surface roughness was measured through
198 an atomic force microscope (AFM, SP13800N/SPA400, SII Co., Japan) in tapping mode.

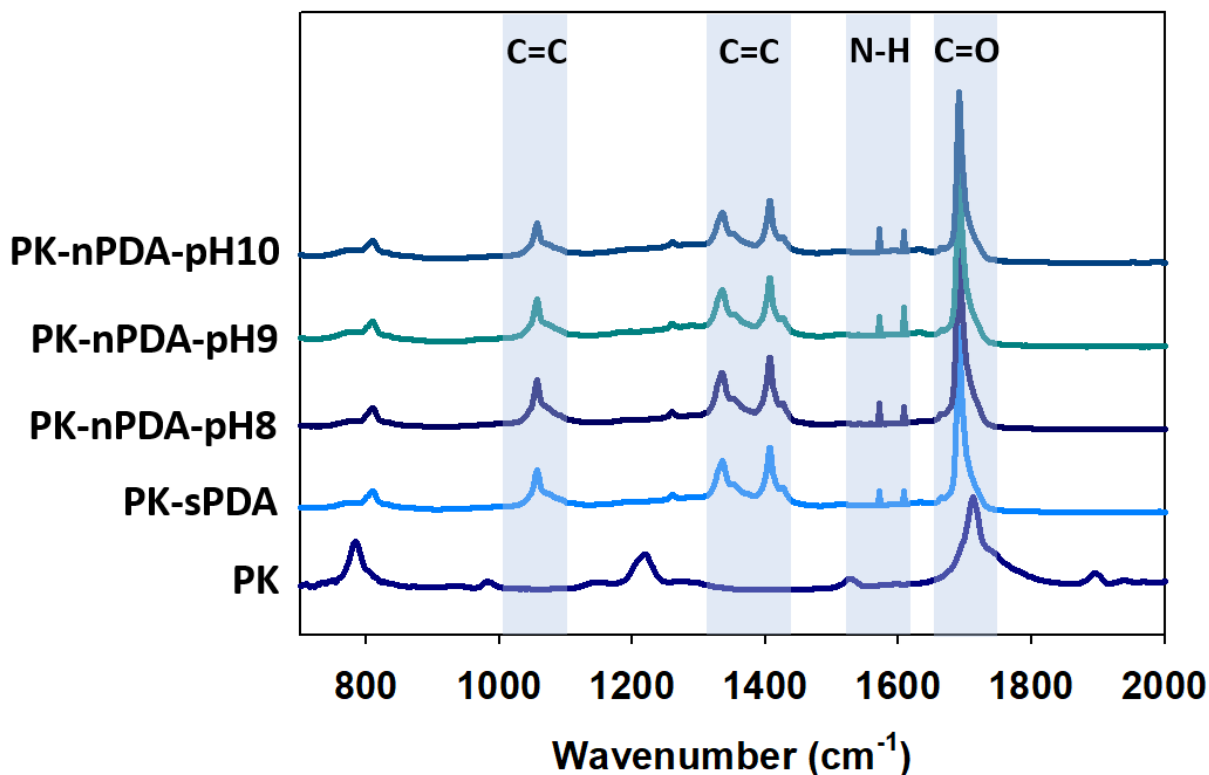
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200 **3. Results and discussion**

201 *3.1. Polydopamine modification and formation of smooth and nanoparticle polydopamine interlayer*

202 Figure 1 shows the FTIR spectra of the membrane substrates, comparing the plain polyketone with the
203 smooth and particulate PDA-coated ones. Peaks at 1705 cm^{-1} correspond to the PK substrate, as evidently
204 seen from all samples. All the PDA-coated membrane substrates exhibited peaks attributed to the presence
205 of PK and PDA groups, indicating the successful incorporation of the PDA interlayer. The peaks at around
206 1020 and 1350 cm^{-1} correspond to the C=C groups, while vibrations around 1600 cm^{-1} correspond to N-H
207 group, indicating that PDA was formed during the self-polymerization of dopamine [35]. There was no
208 substantial difference from the FTIR spectra of the membranes coated with PDA, whether smooth or
209 particulate.

210



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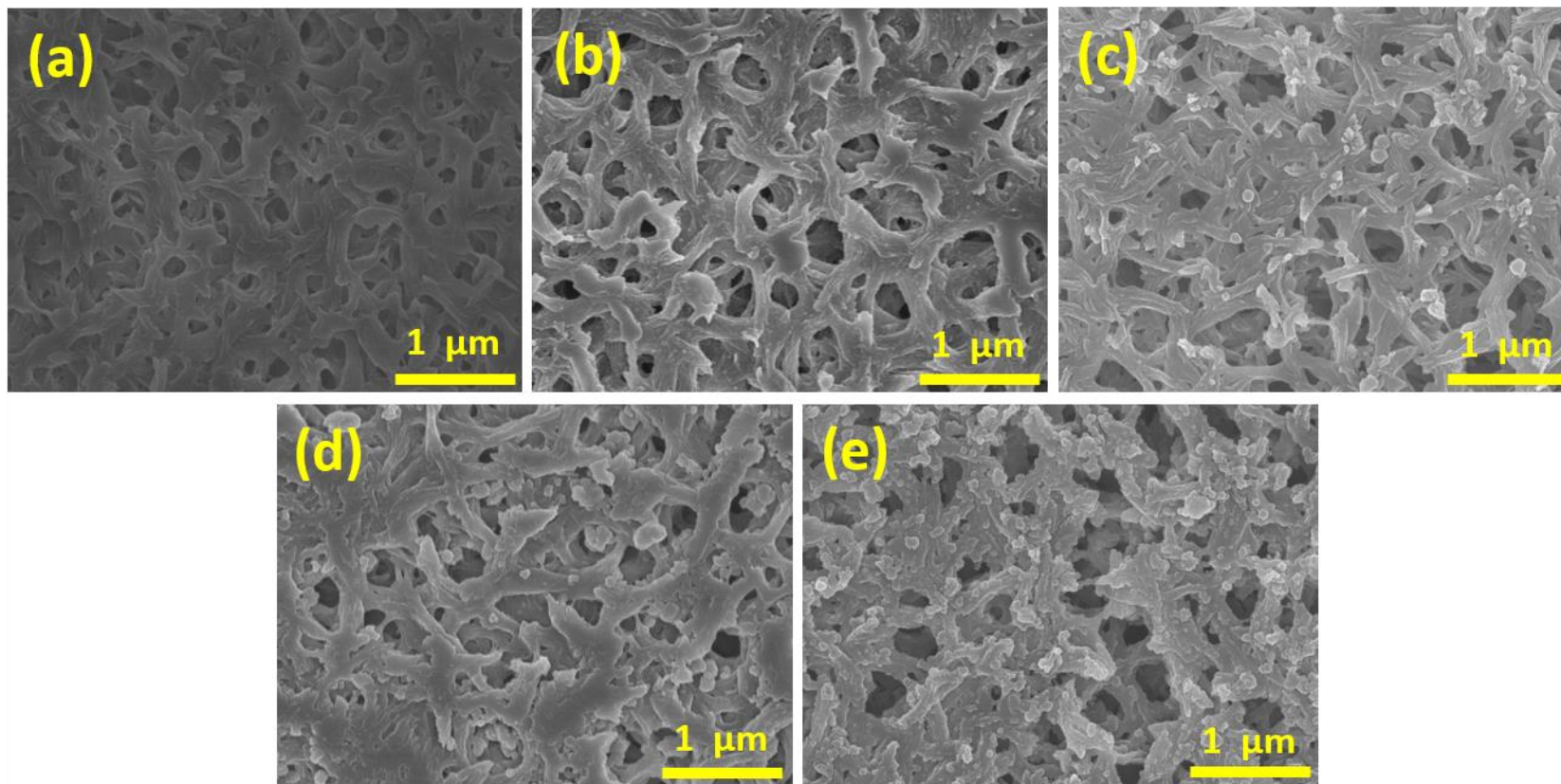
212 Figure 1. FTIR spectra of the pristine polyketone and PDA-modified membrane substrates.

213

214 To further analyze the surface chemistry of the PDA-coated membrane substrates, XPS analysis was
 215 performed, and the elemental XPS analysis results are shown in Table 1. The elemental composition of the
 216 membrane substrate surface confirms that there was indeed the presence of the PDA coating on top of the
 217 polyketone, as evidenced by the increase in N content, starting from zero in pristine polyketone to the
 218 5.28 % N content of the membrane substrate modified with PDA at pH 10. The self-polymerization of
 219 dopamine to polydopamine is highly controlled by the pH of the reaction medium, with OH⁻ ions acting as
 220 the catalyst. Typically, the pH of the self-polymerization reaction is controlled using a buffer to ensure the
 221 gradual polymerization and facilitate the formation of a smooth polydopamine coating onto a surface. In
 222 the case of this study, the self-polymerization was catalyzed by ammonia, and the pH of the solution

223 influenced the reaction. At lower pH (pH 8), the self-polymerization reaction was not fully catalyzed, as
224 the reaction mostly takes place between pH 8.5 to 9, thus the dopamine which has not polymerized during
225 the reaction period might have been washed off from the membrane surface, resulting in a lower N content,
226 as seen from the XPS characterization, indicating the presence of less polydopamine on the membrane
227 substrate surface. The higher reaction pH of the succeeding samples ensured that the self-polymerization
228 reaction took place and resulted in the formation of the PDA nanoparticles which were embedded within
229 the membrane substrate, and led to higher N content.

230 The morphology of the membrane substrates characterized using FE-SEM are shown in Figure 2. In
231 general, PDA modification leads to enhancement of hydrophilicity and smoothness of the surface. However,
232 in this study, it could be seen that while hydrophilicity was definitely enhanced upon introduction of the
233 PDA interlayer [14], surface smoothness was only achieved when the conventional PDA modification
234 technique was performed (Figure 2(b)). The smooth PDA interlayer appeared to be a smooth coating on the
235 fibril-like structures of polyketone, still revealing the intrinsic highly porous structure of the membrane
236 substrate. In the case of PDA nanoparticle formation as an interlayer, the roughness of the membrane
237 substrate was observed to increase as pH increased (Figure 2(c)-2(e)). The particulate PDA interlayer was
238 shown to exhibit granular surface with open pore structure. As the pH of dopamine self-polymerization
239 reaction increased, there was an observed increase in the presence of particulate PDA structure attached
240 onto the fibrous polyketone substrate. The nPDA-pH10 substrate has clearly shown aggregation of the PDA
241 nanoparticles, which was known to be more prevalent at elevated pH [20], thus self-polymerization of
242 dopamine is not performed at pH values higher than 10.



243

244 Figure 2. Surface morphology of the (a) pristine polyketone and PDA-coated membrane substrates: (b) sPDA, (c) nPDA-pH8, (d) nPDA-pH9, and
245 (e) nPDA-pH10.

246 Table 1. Surface elemental analysis of the PDA intermediate layer using X-ray photoelectron spectrometry.

Membrane substrate	C (%)	N (%)	O (%)	Others (%)
Polyketone	70.6	-	29.0	0.4
sPDA	72.3	3.1	24.3	0.3
nPDA-pH8	71.9	2.8	25.0	0.3
nPDA-pH9	73.9	4.6	21.2	0.3
nPDA-pH10	75.3	5.3	19.2	0.2

247

248 The membrane substrate roughness was also characterized using AFM. Table 2 shows the average root-

249 mean-square surface roughness (R_{ms}) of the prepared polyketone and PDA-coated membrane substrates.

250 The surface roughness of both pristine polyketone and sPDA samples showed statistically similar average

251 R_{ms} values, with sPDA having a lower average, indicating how the conventional PDA coating could result

252 in the formation of a smooth coating on top of the membrane substrate. The surface roughness, however,

253 increased as pH increased during the self-polymerization of dopamine. The surface roughness was heavily

254 influenced by the formation of the nano-sized PDA particles, which have been formed upon exposure of

255 dopamine monomer with NH_3 initiator.

256

257 Table 2. Average root-mean-square roughness (R_{ms}) of the membrane substrates characterized using

258 AFM.

Membrane substrate	Average Roughness, R_{ms} (nm)
Polyketone	59.3 ± 10.2
sPDA	48.2 ± 8.4
nPDA-pH8	96.6 ± 17.1
nPDA-pH9	129 ± 21.6
nPDA-pH10	160 ± 33.6

259

260 Figure 3(a) shows the contact angle measurements of the plain and PDA-modified polyketone

261 substrates. Due to the functional groups present in the structure of PDA, the membrane substrate surfaces

262 were also effectively functionalized upon coating and the hydrophilicity of the surfaces were effectively

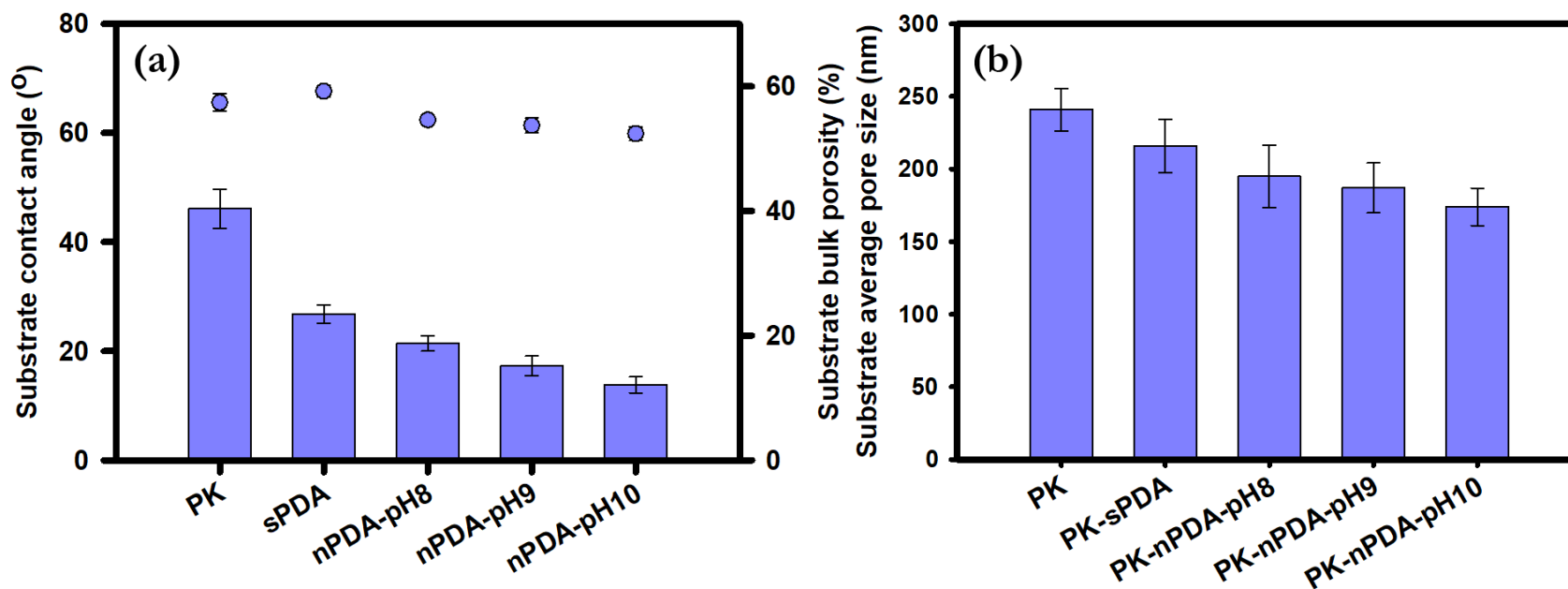
263 enhanced as well [37]. The effect of surface roughness also influenced the surface wettability of the

264 membrane substrates. The aggregation of nanoparticulate PDA on the membrane substrate surface
265 effectively increased the surface roughness, as shown in Table 2. Aside from hydrophilicity, surface
266 wettability is also influenced by the roughness, as given by the Wenzel equation [38]:

$$267 \cos \theta_{app} = r \cos \theta \quad (8)$$

268 where θ_{app} , θ , and r are the respectively apparent contact angle, actual contact angle on a perfectly smooth
269 solid surface, and the ratio of the rough surface area and projected area. Thus, it is expected that a lower
270 contact angle measurement would be obtained on a rougher surface.

271 It is also interesting to note that PDA modification of the membrane substrates resulted in smaller
272 surface pores and narrower pore size distribution as shown in Figure 3. It was expected that PDA
273 modification could cause partial blockage of the surface pores [31]. The number of surface pores decreased
274 due to the introduction of the intermediate layer following the PDA polymerization. The nano-sized PDA
275 particles were positioned on the support layer interface. Aggregated nano-sized particles were observed on
276 the surface, as shown in Figure 2, which blocked the surface pores [22]. The PDA coating, however, makes
277 up for the decrease in porosity by providing higher functionality and hydrophilicity to the membrane
278 substrate. The bulk porosity of the membrane substrates were determined using gravimetric method, to
279 account for the “wetted porosity” effect of PDA coating, as introduced by Arena et al. [39]. As shown in
280 Figure 3(a), the bulk porosity of PK-sPDA substrate increased from that of the pristine PK substrate, as an
281 effect of the PDA coating. The porous structure of the membrane substrates coated with nanoparticulate
282 PDA exhibited lower bulk porosity values; however, this is mainly attributed to the partial blockage of the
283 surface pores due to the growth of PDA nanoparticles. This was supported by the average surface pore size
284 values shown in Figure 3(b).



285

286 Figure 3. (a) Surface contact angle and bulk porosity and (b) average surface pore size of the pristine polyketone and polydopamine-coated
 287 polyketone membrane substrates (measured using ImageJ software).

288 3.2. TFC membrane characterization

289 The morphology of the TFC membranes observed using FE-SEM are shown in Figure 4. Upon
290 inspection of the polyamide surface morphology of the membrane samples (Figure 4(a)), it is interesting
291 that the polyamide of the plain TFC and that with smooth PDA interlayer were observed to be highly dense,
292 while the polyamide of the membranes with nanoparticle PDA interlayer were observed to be less dense,
293 which may indicate the formation of looser polyamide structure. This difference in the morphology of the
294 polyamide formed is highly attributed to the properties of the intermediate layer between the substrate and
295 the polyamide layer. The polyamide selective layer formation resulted from the uniform distribution of the
296 aqueous solution on the highly hydrophilic surface and the slower diffusion of MPD during IP as a result
297 of the introduction of the intermediate layer [40]. Furthermore, the cross-section images show the changes
298 in the morphology of the selective layers formed on top of the porous substrate. There was a noticeable
299 change in the polyamide layer thickness for the samples. Polyamide has a typical thickness of 100 to 150
300 nm, and the measured thickness of the polyamide selective layer of the plain TFC membrane was well
301 within this range. The same observation was true for the polyamide thickness of the TFC membrane with a
302 smooth PDA interlayer; however, the polyamide layer of sPDA TFC was observed to be even thinner than
303 that of the plain TFC membrane. This shows that the smooth PDA interlayer forms denser and thinner
304 selective layers. Aside from being caused by the smoothness of the membrane substrate, this phenomenon
305 is also most likely due to the ability of PDA to penetrate into the pores of the membrane substrate, enhancing
306 its hydrophilicity [41] and improving the retention of the MPD precursor during IP, leading to the formation
307 of denser, yet thinner polyamide [42]. On the other hand, there was an increase, albeit slightly, in the
308 apparent polyamide thickness observed for the membranes modified with a nanoparticulate PDA interlayer.

309 Surface roughness of the TFC membranes was measured and shown in Figure 5(a). The surface
310 roughness data support the membrane surface morphology shown from the FE-SEM images. The
311 morphology of the substrate influenced the formation of the polyamide selective layer. The roughness of
312 the membrane substrates coated with nanoparticulate PDA intermediate layer resulted in the formation of

313 rougher polyamide layer, and TFC membrane roughness increased as the pH of PDA polymerization
 314 increased. The R_{ms} values of the PK-nPDA samples were much greater than those of the pristine TFC and
 315 PK-sPDA TFC membranes. The polyamide selective layer surface roughness can be attributed to the
 316 presence of more ridge structures formed atop the particulate PDA intermediate layer. Membrane surface
 317 roughness is directly related to enhanced affinity to water, and tuning the membrane surface roughness
 318 could enhance the membrane permeability [43]. Membrane separation performance is highly dependent on
 319 the roughness of the polyamide selective layer, due to the increased surface filtration area [44]. The surface
 320 water contact angle of the TFC membranes was also measured and presented in Figure 5(b). Due to the
 321 relatively hydrophobic property of polyamide, the contact angle measurements of the TFC membranes were
 322 observed to increase from those of their substrate counterparts. The lowest contact angle measurement was
 323 observed for PK-nPDA-pH10 (47.2°), owing to the membrane surface roughness (Figure 5(a)).

324 To further characterize the polyamide selective layer of the TFC membranes and how its formation
 325 was influenced by the introduction of the intermediate layer, XPS analysis was performed to determine the
 326 atomic composition and degree of cross-linking of the polyamide layer in each membrane. Based on the
 327 atomic composition analysis, the ratio of O and N can provide the degree of cross-linkage of the polyamide
 328 layer. The O/N ratio is inversely proportional to the degree of cross-linkage, which can be calculated using:

$$329 \quad a + b = 1 \quad (7)$$

$$330 \quad \frac{O}{N} = \frac{3a+4b}{3a+2b} \quad (8)$$

331 where a and b are the fractions of the crosslinked structure and linear structure components of polyamide,
 332 respectively [45].

333

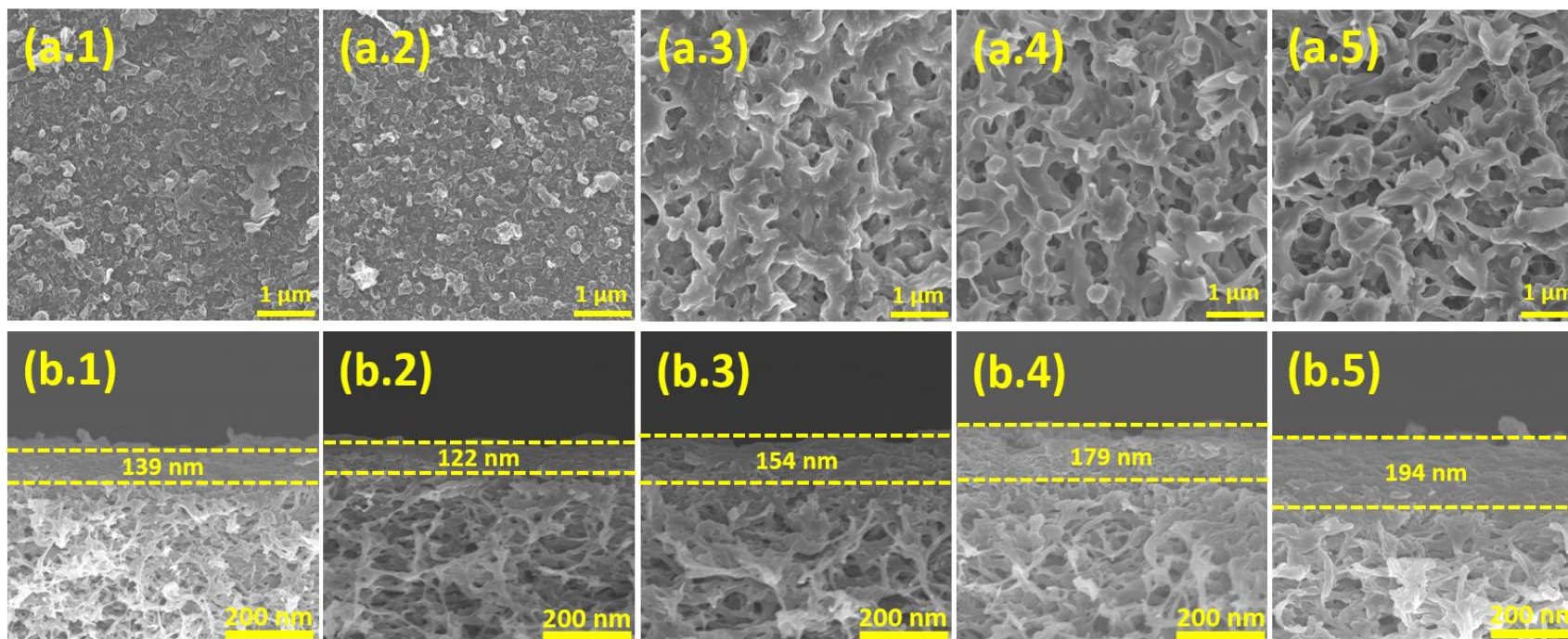
334 Table 3. Atomic composition and the degree of cross-linking of the polyamide selective layer of the TFC
 335 membranes.

Membrane	C (%)	N (%)	O (%)	O/N	a value
----------	-------	-------	-------	-----	---------

Polyketone TFC	72.3	13.1	14.6	1.12	0.83
sPDA TFC	72.1	13.3	14.6	1.09	0.87
nPDA-pH8 TFC	71.7	12.8	15.5	1.21	0.72
nPDA-pH9 TFC	72.6	12.1	15.3	1.26	0.66
nPDA-pH10 TFC	73.0	11.7	15.3	1.30	0.61

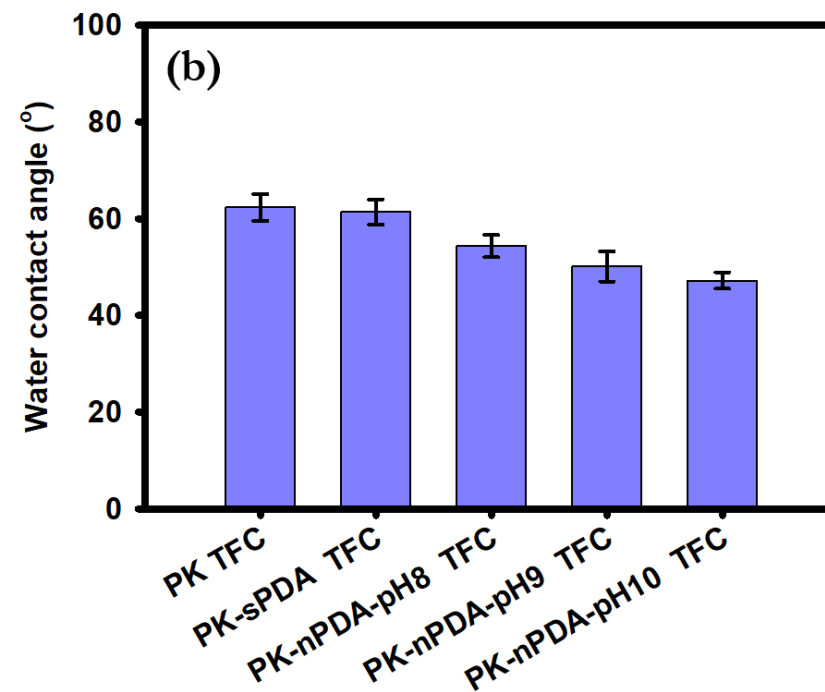
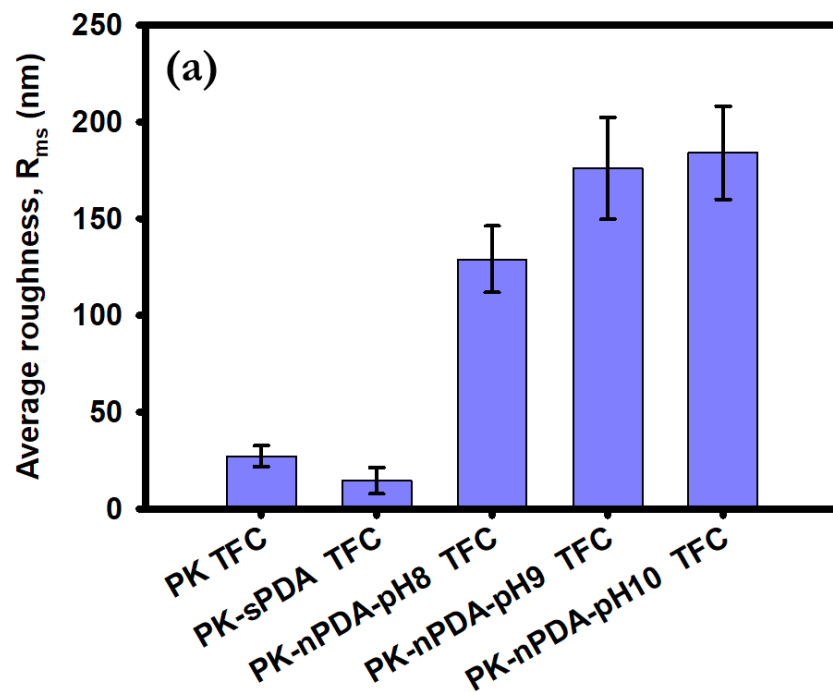
336

337 Table 3 shows the atomic composition and fraction of cross-linking component of the polyamide
338 layers of the TFC membranes prepared in this study. The XPS analysis shows and confirms that the
339 introduction of intermediate PDA layer could influence formation of the polyamide selective layer. Both
340 the pristine TFC and sPDA TFC membranes show a high degree of cross-linkage of 0.83 and 0.87,
341 respectively, while those with nanoparticulate PDA intermediate layer exhibited lower degree of cross-
342 linkage between 0.60 and 0.72. The presence of the embedded PDA nanoparticles increased roughness of
343 the membrane substrate, causing a less even distribution of the MPD and TMC monomers during IP and
344 resulting in lower degrees of cross-linkage. The lower degree of cross-linkage then explains the appearance
345 of looser polyamide layer of the nPDA TFC membranes, as shown by the FE-SEM images (Figure 5).



346

347 Figure 4. Polyamide selective layer (a) surface and (b) cross-section FE-SEM images of the TFC membranes: (1) plain TFC, (2) sPDA TFC, (3)
 348 nPDA-pH8 TFC, (4) nPDA-pH9 TFC, and (5) nPDA-pH10 TFC. Polyamide layer thickness of the TFC membranes are indicated in each figure.



349
 350 Figure 5. (a) Average root-mean-square surface roughness (R_{ms}) and (b) surface water contact angle of the TFC membranes.

351 3.3. Membrane intrinsic transport properties

352 The membrane intrinsic transport properties were evaluated and presented in Table 4.

353

354 Table 4. Intrinsic transport properties of the TFC membranes.

Membrane	A ($L m^{-2} h^{-1} bar^{-1}$)	B ($L m^{-2} h^{-1}$)	B/A (bar)	R (%)	S (μm)
PK TFC	1.36 ± 0.13	0.33 ± 0.05	0.24	97.5 ± 0.3	212
PK-sPDA TFC	2.12 ± 0.08	0.39 ± 0.12	0.18	98.3 ± 0.4	198
PK-nPDA-pH8 TFC	2.58 ± 0.15	0.87 ± 0.18	0.34	95.3 ± 0.4	225
PK-nPDA-pH9 TFC	2.76 ± 0.19	1.08 ± 0.11	0.39	94.7 ± 0.6	230
PK-nPDA-pH10 TFC	2.41 ± 0.23	1.44 ± 0.16	0.60	92.8 ± 0.8	247

355

356 The membranes exhibited higher water permeability (A) upon PDA modification, when compared to
357 the plain TFC membrane. The water permeability increased for membranes with dopamine self-
358 polymerization at higher pH (pH 8 to 9). However, the membrane modified with particulate PDA interlayer
359 at pH 10 showed slightly lower water permeability which can most likely be explained because of the
360 surface pore blockage due to the presence of the PDA nanoparticles [22]. The highest A value of $2.76 L m^{-2}$
361 $h^{-1} bar^{-1}$ was observed for the PK-nPDA-pH9 TFC sample. The best water permeability is likely caused
362 by the satisfactory coating on top of the membrane surface, without blocking the porous structure of the
363 polyketone, while enhancing the surface hydrophilicity, and the presence of higher free volume of the looser
364 polyamide thin film.

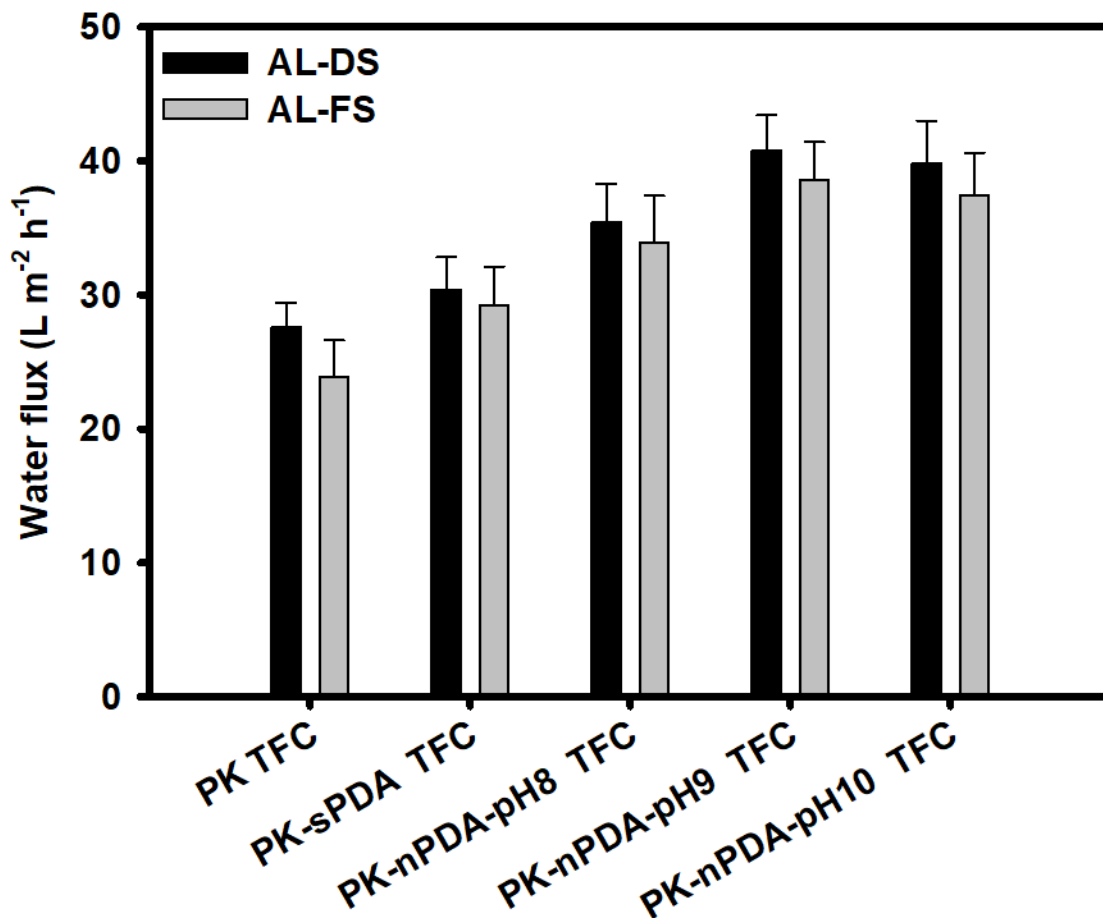
365 As expected, the increase in water permeability is accompanied with corresponding increase in solute
366 permeability as indicated by the solute permeability (B) values in Table 4. The membranes with thinner and
367 denser selective layers exhibited high solute rejection values, while those with looser, less cross-linked, and
368 quite thicker polyamide layers exhibited a decrease in rejection and increase in solute permeability. Overall
369 selectivity of the TFC membranes were given by the B/A values, which indicated that the TFC membranes
370 suffered lower selectivity after formation of PDA interlayer at higher pH.

371 The structure parameter (S value) of the membranes were also determined and shown in Table 4. The
372 introduction of the smooth PDA intermediate layer facilitated the formation of a thin, dense polyamide
373 selective layer and the resultant TFC membrane was found to have a lower S value (198 μm) than the
374 pristine TFC membrane (212 μm). The formation of nanoparticulate PDA intermediate layer, on the other
375 hand, increased the S values within the range of 225-250 μm range.

376

377 ***3.4. Membrane osmotic performance and energy generation capability***

378 Membrane performance for the PRO process was evaluated using DI water and 1.0 M NaCl as the FS
379 and DS, respectively. The TFC membranes were first evaluated for forward osmosis (FO) performance.
380 The water flux (J_w) was obtained after operation with two different membrane orientations: (a) active layer
381 facing the DS (AL-DS) and (b) active layer facing the FS (AL-FS), and the results are shown in Figure 6.
382 Without the application of hydraulic pressure, consistently with the pure water permeability data, PK-
383 nPDA-pH9 TFC showed the highest initial water flux of 40.8 L m⁻² h⁻¹, with a sharp flux decline at PK-
384 nPDA-pH10 TFC. Comparison of the water flux values at both AL-DS and AL-FS membrane orientations
385 could give light on the internal concentration polarization (ICP) occurring in the TFC membrane, which
386 could dramatically decrease water flux. While all the J_w values were observed to be lower during AL-FS
387 operation, it can be noted that the difference was smaller for the TFC membranes with PDA interlayer,
388 which could indicate that the effect of ICP did not affect the membrane water permeability performance
389 [46]. The water flux of the TFC membranes was observed to increase after the introduction of the PDA
390 intermediate layer. In the case of the PK-sPDA TFC membrane, the formation of thinner, dense polyamide
391 layer resulted in increased water permeability. On the other hand, the PK-nPDA TFC membranes had
392 increased water flux values compared to pristine PK-TFC and PK-sPDA, due to the increased surface
393 roughness and formation of looser polyamide selective layer. However, the observed flux decline for PK-
394 nPDA-pH10 TFC could be attributed to the pore blockage within the membranes resulting from the
395 aggregation of the nanoparticulate PDA.



396

397 Figure 6. Water flux of the TFC membranes during forward osmosis (FO) operation at active layer facing
 398 the draw solution (AL-DS) and active layer facing the feed solution (AL-FS) membrane orientations. (FS:
 399 DI water; DS: 1.0 M NaCl).

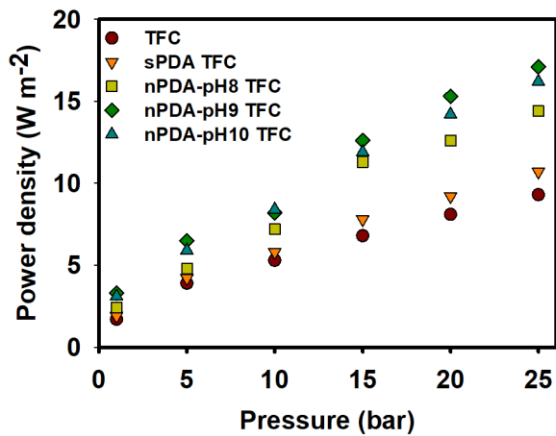
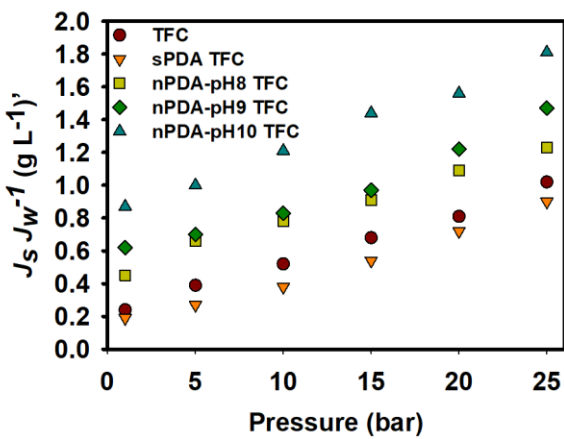
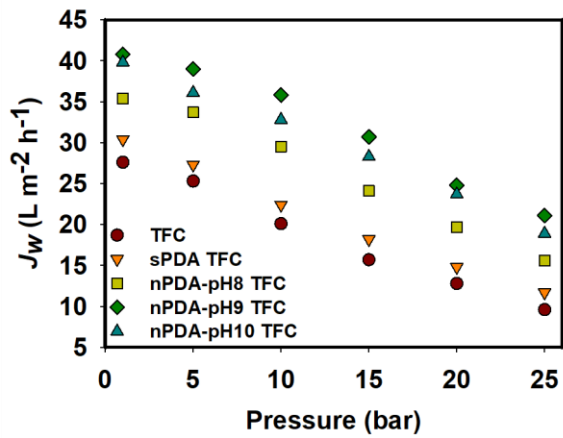
400

401 Figure 7 shows the water flux, specific reverse salt flux ($J_s J_w^{-1}$, g L⁻¹), and power density (W m⁻²) of
 402 the TFC membranes at different applied pressures. Water flux dramatically increased upon the introduction
 403 of the PDA interlayer, indicating the enhanced hydrophilicity of the membrane substrate, which facilitated
 404 water transport. The highly hydrophilic nature of the intermediate layer complemented the microporous

405 structure of the polyketone membrane substrate, thereby resulting in better water transport during osmotic
406 process.

407 The introduction of nanoparticulate PDA intermediate layer and the subsequent production of looser
408 polyamide selective layer caused decline of the membrane selectivity and an increase of reverse salt
409 diffusion during PRO process operation. These were not observed for the plain TFC membrane and the
410 smooth PDA interlayer-modified TFC membrane, which both exhibited specific reverse salt flux values
411 lower than 0.3 g L^{-1} for osmotic operation at minimal applied hydraulic pressure (Figure 7(b)). With the
412 formation of looser polyamide atop the nanoparticulate PDA intermediate layer, even at minimal applied
413 hydraulic pressure, the specific reverse salt flux increased dramatically to 0.45 g L^{-1} for nPDA-pH8 TFC
414 and 0.87 g L^{-1} for PK-nPDA-pH10 TFC. As the applied pressure increased during PRO operation, the
415 specific reverse salt flux values were observed to increase as well. The formation of nanoparticulate PDA
416 intermediate layer significantly affected the selectivity of the TFC membranes; thus, the PDA nanoparticle
417 formation should be optimized to enhance the water permeability, without sacrificing the membrane
418 selectivity which may cause considerable loss of osmotic driving force by severe concentration polarization
419 during operation [47].

420 In terms of power density, the highest power density of 17.1 W m^{-2} was obtained from PK-nPDA-
421 pH-9 TFC at an applied hydraulic pressure of 25 bar as presented in Figure 7(c). The increase in the power
422 density was dramatic compared to that of the plain TFC membrane, whose power density at 25 bar was
423 only 8.6 W m^{-2} . Based on the increasing trend in the power density of the membranes as presented in Figure
424 6(c), the maximum power density has yet to be achieved. The membranes were not tested at applied pressure
425 higher than 25 bar because of limitation with the mechanical strength of the membrane. This shows that
426 with further optimization of the membrane mechanical strength, these membranes have the potential to
427 achieve even higher power density at higher applied pressure.



428

429 Figure 7. Osmotic performance, in terms of (a) water flux (J_w), (b) specific reverse salt flux ($J_s J_w^{-1}$), and
 430 (c) power density, of the TFC membranes during PRO operation at different applied hydraulic pressures
 431 using DI water as FS and 1.0 M NaCl as DS.

432 4. Conclusion

433 In this study, polydopamine (PDA) intermediate layer was formed in between the aliphatic
434 polyketone membrane substrate and the polyamide selective layer of a pressure retarded osmosis (PRO)
435 thin film composite (TFC) membrane. Two types of PDA interlayer were formed and compared: the
436 conventional smooth PDA and particulate PDA. Furthermore, the effect of pH during the self-
437 polymerization of dopamine was investigated and evaluated in terms of the ease of PDA nanoparticle
438 formation and its subsequent effect to the polyamide formation during interfacial polymerization and the
439 membrane performance during PRO process operation. The following conclusions can be drawn from this
440 study:

- 441 1. The morphology of the PDA intermediate layer significantly influences the formation of polyamide
442 selective layer. Both the pristine TFC and sPDA TFC (modified with smooth PDA interlayer)
443 membranes exhibited highly dense and thinner polyamide layer, while those with the particulate
444 PDA interlayer were observed to have formed looser and quite thicker polyamide layers.
- 445 2. Not only does the structure of the polyamide selective layer was affected by the intermediate layer
446 morphology, the membrane performance was likewise affected. Introduction of the particulate PDA
447 interlayer created more free volume and transport channels, which compensated the pore-blocking
448 phenomenon of the nano-sized PDA particles. This resulted in higher water permeability for nPDA-
449 pH8 TFC and nPDA-pH9 TFC. However, for nPDA-pH10 TFC, the more alkaline environment
450 during dopamine self-polymerization resulted in better nucleation of the PDA nanoparticles, which
451 as evidenced by the membrane characterization techniques.
- 452 3. The looser polyamide selective layer formed on top of the particulate PDA intermediate layer also
453 led to slight decrease in the membrane selectivity, compared with the pristine and sPDA TFC
454 samples.
- 455 4. The membrane performance was evaluated using deionized water and 1.0 M NaCl as feed and draw
456 solutions, respectively. The TFC membrane with nanoparticulate PDA layer formed at pH 9.0

457 exhibited the best initial water flux of $40.8 \text{ L m}^{-2} \text{ h}^{-1}$, and this membrane also showed the highest
458 power density of 17.1 W m^{-2} at 25 bar.

459 This study was able to prove that a simple, facile, and versatile technique such as the ammonia-
460 initialized introduction of nanoparticulate PDA intermediate layer, at controlled conditions, could
461 significantly improve the water transport properties of a TFC membrane, while maintaining satisfactory
462 rejection ability.

463

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