

Elsevier required licence: © 2021

This manuscript version is made available under the  
CC-BY-NC-ND 4.0 license

<http://creativecommons.org/licenses/by-nc-nd/4.0/>

The definitive publisher version is available online at

<https://doi.org/10.1016/j.scitotenv.2021.145793>

# Microplastic Particles in the Aquatic Environment: A Systematic Review

Mohammad Boshir Ahmed<sup>a,b,c†</sup>, Md. Saifur Rahman<sup>a,b†</sup>, Jahangir Alom<sup>b†</sup>, Md. Saif Hasan<sup>b†</sup>, M.  
A. H. Johir<sup>c</sup>, M Ibrahim H Mondal<sup>b</sup>, Da-Young Lee<sup>a</sup>, Jaeil Park<sup>a</sup>, John L. Zhou<sup>c\*</sup>, Myung-Han  
Yoon<sup>a\*</sup>

<sup>a</sup>School of Material Science and Engineering, Gwangju Institute of Science and Technology,  
Gwangju, 61005, Republic of Korea

<sup>b</sup>Department of Applied Chemistry and Chemical Engineering, University of Rajshahi, Rajshahi  
6205, Bangladesh

<sup>c</sup>Centre for Green Technology, School of Civil and Environmental Engineering, University of  
Technology Sydney, 15 Broadway, Sydney, NSW 2007, Australia

<sup>†</sup> equally contributed

## **Corresponding Authors:**

Prof. John L. Zhou (JunLiang.Zhou@uts.edu.au)

Prof. Myung-Han Yoon (mhyoon@gist.ac.kr)

23 **Abstract**

24 Microplastics (MPs) pollution has become one of the most severe environmental concerns today.  
25 MPs persist in the environment and cause adverse effects in organisms. This review aims to present  
26 a state-of-the-art overview of MPs in the aquatic environment. Personal care products, synthetic  
27 clothing, air-blasting facilities and drilling fluids from gas-oil industries, raw plastic powders from  
28 plastic manufacturing industries, waste plastic products and wastewater treatment plants act as the  
29 major sources of MPs. For MPs analysis, pyrolysis-gas chromatography-mass spectrometry (Py-  
30 GC-MS), Py-MS methods, Raman spectroscopy, and FT-IR spectroscopy are regarded as the most  
31 promising methods for MPs identification and quantification. Due to the large surface area to  
32 volume ratio, crystallinity, hydrophobicity and functional groups, MPs can interact with various  
33 contaminants such as heavy metals, antibiotics and persistent organic contaminants. Among  
34 different physical and biological treatment technologies, the MPs removal performance decreases  
35 as membrane bioreactor (> 99%) > activated sludge process (~98%) > rapid sand filtration  
36 (~97.1%) > dissolved air floatation (~95%) > electrocoagulation (> 90%) > constructed wetlands  
37 (88%). Chemical treatment methods such as coagulation, magnetic separations, Fenton, photo-  
38 Fenton and photocatalytic degradation also show moderate to high efficiency of MP removal.  
39 Hybrid treatment technologies show the highest removal efficacies of MPs. Finally, future research  
40 directions for MPs are elaborated.

41  
42 **Keywords:** Microplastics analysis; Contaminant interactions; Physical treatment; Chemical  
43 technology; Biological process

## 44 **1. Introduction**

45       Plastics have found their importance for improving human lives due to their long durability,  
46 excellent mechanical properties, resistance to weathering and long service life. Plastic production  
47 has started to increase worldwide since 1950, and continues to grow on a massive scale. Its global  
48 production was 361 million tonnes in 2019, 299 million tonnes in 2014, and only 1.7 million tonnes  
49 in the first year of its journey in 1950 (Auta et al., 2017). Microplastics (MPs) are plastic particles  
50 smaller than 5 mm in size (Thompson et al., 2009). MPs are either derived from large plastic  
51 particles in various mechanical and photo-oxidative processes (i.e. secondary MPs) or  
52 manufactured in such small sizes deliberately to use for particular purposes (i.e. primary MPs)  
53 (Hidalgo-Ruz et al., 2012). The use of primary MPs is increasing tremendously in various fields  
54 daily. They are widely used in personal care products (e.g., cosmetics, toothpaste, facial cleansers,  
55 deodorant, baby products, peelings, sunscreen, body washes, resin pellets, hair colors, nail polish,  
56 bath gels, eye shadows, insects' repellents), air-blasting facilities, synthetic clothing, gas-oil  
57 drilling fluids, plastic powders in molding and countless everyday tasks (Alomar et al., 2016). On  
58 the other hand, hills of secondary MPs are being formed in the environment every day through the  
59 wrecking of various sized plastic debris over time in terrestrial and marine environments  
60 (Evangelidou et al., 2020). Biological, physical and chemical processes break down the structural  
61 integrity of larger plastic particles which later turn into microscopic fragments. Different  
62 environmental factors (e.g. temperature, humidity, salinity), polymer properties (such as  
63 morphology, size, shape, density) and biogeographic conditions can influence the formation of  
64 secondary MPs (Auta et al., 2017; Min et al., 2020; Chamas et al., 2020; Anjana et al., 2020). Major  
65 sources of MPs are shown in the appendix (**Figure A1**).

66       MPs enter the environment in various ways. MPs from personal care products and other  
67 different sources are incorporated directly with municipal wastewater, and due to their microscopic  
68 sizes, often are incompletely removed during wastewater treatment plants (WWTPs) and  
69 discharged into the aquatic ecosystems. Moreover, atmospheric fallouts, wind advection, storm-  
70 water runoff, and improper waste management are the major causes of MP release to the  
71 environment (Mason et al., 2016).

72       To effectively assess the risks of MPs to the aquatic environment and human health requires  
73 rigorous analytical methods, which typically follow several steps including sampling, extraction,  
74 isolation (or separation), identification and quantification (or classification). The identification of  
75 MPs is carried out depending on the physical properties (size, shape and colour) and chemical

76 characteristics (polymer type) of isolated particles in mixtures of inorganic and organic residue  
77 particles. Due to the small size of MPs, their quantitative analysis is still a challenge to overcome.

78 MPs highly persist in the environment due to their slow rates of degradation (Eerkes-Medrano  
79 et al., 2015). To date, chronic toxicity of MPs has been reported in various studies due to their  
80 extended exposure in human and other organisms, although no evidence has been found of their  
81 acute effects (Li et al., 2018b; Sussarellu et al., 2016; Prata et al., 2020; Chen et al., 2020;). They  
82 can influence the physiological activities of living communities through leaching of contaminants  
83 from plastics (e.g. plasticizers, flame retardants), and by acting as a vector of persistent organic  
84 contaminants and heavy metals (Hu et al., 2020; Benson and Fred-Ahmadu, 2020; Luo et al., 2020;  
85 Lee et al., 2020; Kalcíková et al., 2020). Some of the abundant types of MPs in the environment,  
86 their typical forms of appearance, as well as common organic contaminants and heavy metals for  
87 which MPs act as a significant vector to biotic organisms, are listed in the appendix (**Table A1**).

88 Due to their small size, MPs can be mistakenly ingested by a wide range of organisms which  
89 cannot distinguish between their nourishments and MP particles (Kaposi et al., 2014; Lönnstedt  
90 and Eklöv, 2016; Dong et al., 2020; Fu et al., 2020). MPs directly impair many species of birds,  
91 reptiles, and fishes as the ingested MPs can stay in the gastrointestinal tract for a long time or may  
92 clog their stomachs and guts (Carlin et al., 2020; Solleiro-Villavicencio et al., 2020; Roch et al.,  
93 2020; Cole et al., 2013).

94 A number of physical, chemical and biological treatment technologies have been implemented  
95 recently to degrade or remove MPs from the environment. Physical methods are most widely used,  
96 including adsorption, screening, grit/primary sedimentation, membrane filtration, density  
97 separation, dissolve air flotation and magnetic separation. Till now, filtration technology such as  
98 ultrafiltration (UF), sand filtration and granular filtration has been found as the most effective,  
99 economic, energy-efficient process used in WWTPs of different sizes. In addition, reverse osmosis  
100 (RO) has become popular over the past few years for MPs removal. In many cases, UF is coupled  
101 with RO process to increase the efficiency of treatment process (Qin et al., 2004). With more than  
102 99% efficiency, membrane bioreactor (MBR) has become the most popular treatment technology  
103 among all of the biological treatment methods for MPs removal (Carr et al., 2016;). Other  
104 biological methods include activated sludge process, aerobic digestion, anaerobic digestion (AD),  
105 biological degradation and constructed wetlands (CWs). Chemical treatment methods include  
106 oxidation, photo-oxidation, photo-catalytic degradation, coagulation, Fenton, photo-Fenton and  
107 acid-alkali treatment. Electrochemical methods such as electrocoagulation (EC) and electro-Fenton

108 processes have added a new dimension in chemical methods to improve their efficiency. Currently,  
109 pyrolysis and co-pyrolysis are being touted as one of the most promising approaches for MPs  
110 removal with extra advantages of low-cost fuel production (Ahmed et al., 2011; Burra and Gupta,  
111 2018b; Pinto et al., 2002).

112 Several review articles are available based on MPs especially on the occurrence of MPs in the  
113 marine environment, MPs analysis, and MPs remediation technologies. However, none of these  
114 review articles provides a systematic overview of MPs especially from the sources to treatment  
115 technology. Hence, the main aim of this review is to provide the state-of-the-art overview of MPs  
116 in the aquatic environment by incorporating all relevant aspects and critically analyzing the  
117 available information from literature together with advantages and limitations of each technology.  
118 Future research directions for MPs are also proposed based on the review.

119

## 120 **2. Sources of MPs**

### 121 **2.1. Abundance of MPs**

122 MPs have been found in urban and rural areas (Hirai et al., 2011), seawater, sediment (Hidalgo-  
123 Ruz et al., 2012; Van Cauwenberghe et al., 2015; ). MPs are also found in the air (Gasperi et al.,  
124 2018), soil (Bläsing and Amelung, 2018) and organisms (Zhao et al., 2016). Several reviews have  
125 been published on the presence of MPs in rivers, ocean and lakes (Horton et al., 2017; Mendoza et  
126 al., 2018). Hidalgo-Ruz et al. (2012) studied the abundance of MPs in the marine environment, and  
127 suggested MPs density to be 0.001-1 items/m<sup>2</sup> on the sea surface and 1-100000 items/m<sup>2</sup> in the  
128 sediment. Light MPs have been found in the freshwater surface as floating debris ranging from  
129  $1 \times 10^3$  to  $68 \times 10^5$  particles/km<sup>2</sup> (Anderson et al., 2017; Free et al., 2014; Su et al., 2016).  
130 Furthermore, the denser MPs are usually found in the soil and sediment (Di and Wang, 2018; Su et  
131 al., 2016). The abundance of MPs in the aquatic environment is presented in **Table A2**.

132 The reported MPs concentration in the sediment of St. Lawrence River was 13832  
133 microbeads/m<sup>2</sup>, implying high MPs content in association with high population density (Castañeda  
134 et al., 2014). Similar suggestion was made in the analysis of MPs distribution in sediment from the  
135 Rhine and Main rivers (Klein et al., 2015). However, water column caught a notable quantity of  
136 suspended plastics particles (da Costa et al., 2016; Leslie et al., 2017); the measured concentration  
137 ranging from 1.6 to 12.6 pieces/L in the three Gorges Reservoir, China (Di and Wang, 2018).  
138 During wastewater treatment, MPs are simply removed but remain in sludge as MPs does not  
139 degrade. Therefore, wastewater sludge contains relatively high MPs e.g., as high as 56000

140 particles/kg (Li et al., 2019a). During the measurement of MP in an urbanized river in Chicago, it  
141 was found that the MPs concentration exceeded that in oceans, which demonstrated that the WWTP  
142 effluent was the primary source of MPs to the coast water (McCormick et al., 2014).

143 The sources of MPs in the aquatic system included domestic wastewater (Carr et al., 2016;  
144 Duis and Coors, 2016), sewage discharge (Eerkes-Medrano et al., 2015; Mintenig et al., 2017),  
145 plastic manufacturing industries (Sadri and Thompson, 2014), and decomposition of large plastics  
146 (Eerkes-Medrano et al., 2015). Road dust such as tyres, bitumen, and road marking paints are other  
147 MPs that transport through the freshwater system to the ocean. It is estimated that an average of  
148 63, 125 and 240 kilo tonnes MPs are discharged through wear and tear of tyres each year in the  
149 United Kingdom, Germany and Japan, respectively (Ngo et al., 2019).

150 Moreover, numerous studies have shown that the abundance of MPs is highly dependent on  
151 the population density and type of human activities (Yin et al., 2020). The abundance of MPs has  
152 been discussed in ocean (Cole et al., 2011), sediment (Alimi et al., 2021), WWTPs (Murphy et al.,  
153 2016), and freshwater including rivers and lakes (Castañeda et al., 2014). The concentrations of  
154 MPs from different sources and locations are summarized in **Table A2**.

## 155 **2.2. MPs in wastewater**

156 Studies show that WWTPs are one of the major sources of MPs in the environment (Carr et al.,  
157 2016; Murphy et al., 2016, Alvim et al., 2020). MPs from different personal care products (e.g.  
158 hand cleaners, toothpaste, skin cleansers, facial scrubs) have gained considerable public attention  
159 during the past few years (Duis and Coors, 2016; Sun et al., 2020;). MPs are incorporated as  
160 ‘exfoliation tools’ in some of these care products and may pass through WWTPs due to their micro  
161 size (Chang, 2015). Some plastic pre-products such as plastic resin pellets and plastic powder  
162 (flakes and fluffs) are also considered as another source of MP particles, which are used in the  
163 fabrication of various plastic products. They may reach the environment by improper handling and  
164 accidental losses during transportation. Residues and garbage from plastic recycling and plastic  
165 processing factories may also pollute waters as a significant contributor of MPs (Andrady, 2011;  
166 Derraik, 2002; Moore, 2008). Protective paints (e.g. ship paints, furniture paints) contain many  
167 synthetic polymers such as polyacrylate, polystyrene (PS), poly-urethane, alkyds and epoxy resins  
168 which may release MPs during application, removal or abrasion of the paints (Song et al., 2014,  
169 Hale et al., 2020, Kwon et al., 2020). Also, abrasion of MPs from some household plastic materials,  
170 car tires, and synthetic textiles (e.g. shirts, PS fleeces, blankets) are major sources of MP particles  
171 in wastewater (Duis and Coors, 2016). MPs are used in some industrial abrasives, drilling fluids

172 (in gas and oil exploration), cleansing agents of engines as well as some other industrial aids and  
173 equipment. Improper disposal and handling of them may cause significant input of MPs in  
174 wastewater (Derraik, 2002).

175 Although significant removal of MPs has been reported in several WWTPs, surprisingly very  
176 few of them can remove MPs with 100% efficacy. An observation by the Swedish Environmental  
177 Research Institute revealed that release of only 0.009 MPs/L from a WWTP (> 99% MPs removal  
178 efficacy) resulted in the total discharge of about 1770 MP particles every hour on average. Murphy  
179 et al., (2016) evaluated a significant number of MPs release from a WWTP despite having 98.41%  
180 removal efficacy. The release of 0.25 MPs/L of final effluent resulted in almost 65 million MPs  
181 release daily with the treated water. In comparison, two tertiary treatment plants in New South  
182 Wales, Australia were found to release about 1.0 MP/L of MP particles in the final effluent.  
183 Variations in the number of treatment steps, type and operating conditions of treatment technologies,  
184 and age of facilities involved in the WWTPs are responsible for the different removal and release  
185 of MPs. Some WWTPs from different geographic regions with varying capacities of MPs treatment  
186 are compared in **Table A3**, which clearly illustrates WWTPs as a significant secondary source of  
187 MPs.

188 Therefore, the WWTPs are considered as one of the most significant sources of MPs though  
189 being a secondary one. Either by implementing more efficient MPs treatment technologies, or by  
190 increasing the number of treatment steps, or by treating the contaminated water in two/three  
191 repeating cycles instead of the one-time flow system, the release of MPs from this major source  
192 can be mitigated significantly.

### 193 **3. Methods of MPs analysis**

194 To accurately assess the risks of MPs in the aquatic environment requires a standard analysis  
195 method for their identification (Koelmans et al., 2019). Analytical method of MPs in environmental  
196 samples follows several steps including sampling, extraction, isolation (or separation),  
197 identification and quantification (**Figure A2**). First of all, the water and sediment samples are  
198 collected from the field which are then filtrated or sieved via density separation techniques. Due to  
199 the presence of biological materials in the sample, the digestion step is required to reduce the  
200 organic matter without affecting the structural or chemical integrity of MPs (Felsing et al., 2018).  
201 In the analysis of MPs, a visual inspection can be used to sort and identify the large-sized MPs (1-  
202 5 mm), whereas dissection microscope can be used to sort smaller sized MPs particles (Doyle et  
203 al., 2011). The digestion step is mainly recommended before the application of visual inspection



204 for MPs identification, via acid digestion, alkali digestion, oxidizing agent, and enzymatic  
205 digestion. However, under high acid concentration and high temperatures, polymers such as nylon,  
206 polyethylene terephthalate (PET) and polyamide (PA) have a low resistance to acid and may also  
207 be degraded or melted at high temperature, and polymers such as polypropylene (PP) and polyvinyl  
208 chloride (PVC) can change their colour (Maes et al., 2017;). In alkali digestion, KOH and NaOH  
209 solutions are widely used to recover MPs and remove biological material (Dehaut et al., 2016). In  
210 comparison, oxidizing agent such as hydrogen peroxide (30-35% H<sub>2</sub>O<sub>2</sub>) is more effective to  
211 degrade organic matter than NaOH and HCl, with little effect on MPs integrity. On the other hand,  
212 enzymatic digestion is less hazardous and causes little impact on MPs integrity when it is used to  
213 degrade organic materials (Maes et al., 2017). Enzymatic digestion followed by an incubation of  
214 H<sub>2</sub>O<sub>2</sub> worked quite well and was able to efficiently destroyed all remaining organic material  
215 (Karlsson et al., 2017).

216 After the extraction and clean-up steps, the identification of MPs is carried out depending on  
217 the physical (size, shape and color) and chemical (polymer type) characteristics of isolated particles  
218 in mixtures of inorganic and organic residue particles. However, MPs analysis methods are still at  
219 development stage, hence there is not yet so-called standard method (Zhang et al., 2020b). The  
220 combination of two or more analytical techniques is required to identify MPs of various sizes,  
221 shapes and polymer types thoroughly and reliably from complex environmental matrices. Among  
222 the available optical analytical methods, Fourier-transform infrared (FTIR) spectroscopy, X-ray  
223 photoelectron spectroscopy (XPS) and Raman spectroscopy have been widely used to analyze MPs  
224 and nanoplastics (NPs) (Hernandez et al., 2019). XPS along with scanning electron microscopy  
225 (SEM) technique is limited to polymer identification but cannot provide elemental information  
226 about MPs (Hernandez et al., 2019; Hernandez et al., 2017), whereas Raman can provide elemental  
227 information about polymer through fingerprint spectra (Schwaferts et al., 2020). The thermal  
228 analysis has also been used to identify MPs.

### 229 **3.1. Microscopy methods**

230 A microscope is a fundamental instrument which helps to identify MPs particles by measuring  
231 their physical property. Stereo microscopy method is broadly-used for MPs identification with size  
232 > 100- $\mu$ m (Kang et al., 2015; Mathalon and Hill, 2014; Nel and Froneman, 2015; Song et al., 2015).  
233 Magnified images help to identify ambiguous, plastic-like particles through detailed surface texture  
234 and structural information of objects using microscopy. Biological materials from sediment  
235 samples make it difficult for microscopic observation to identify MPs because these materials

236 cannot be wholly eliminated by chemical digestion. Several investigations have indicated that false  
237 identification of plastic-like particles and transparent particles using microscopy was often over  
238 20% and 70% respectively, which was confirmed with subsequent spectroscopic analysis (Hidalgo-  
239 Ruz et al., 2012; Song et al., 2015). Under high magnification, it is difficult to identify transparent  
240 or whitish particles using fluorescence microscopy (Löder and Gerdts, 2015).

241 On the other hand, SEM produces apparent and high-resolution images of the surface texture  
242 of MPs particles which are better identified from organic particles (Cooper and Corcoran, 2010).  
243 SEM analysis with energy-dispersive X-ray spectroscopy (EDX) provides the fundamental  
244 component of these particles which is useful for identifying carbon-dominant plastics from  
245 inorganic particles. The combination of SEM and EDX micro-analyzer has also been used in low  
246 vacuum mode to determine the chemical and morphological characteristic of MPs particles (Fries  
247 et al., 2013). In addition, polarized optical microscopy is an advanced microscopy technique which  
248 has been used to identify specific MPs particles such as PE (Von Moos et al., 2012). The main  
249 obstacle of the microscopy method is to accurately distinguish between the synthetic and natural  
250 fibers (**Table 1**) (Song et al., 2015). Therefore, the microscopy method should be combined with  
251 chemical analysis (spectroscopic or thermal analysis) to identify small-sized MPs (<1 mm) (Shim  
252 et al., 2017).

### 253 **3.2. Spectroscopic methods**

254 The spectroscopic technique is more effective for MPs identification, specifically for particles  
255 <500  $\mu\text{m}$ . Micro-Raman or micro-FTIR spectroscopy is a fundamental technique which has been  
256 applied to evaluating marine samples through detailed information on polymer composition. FTIR  
257 and Raman spectroscopy are complementary vibrational techniques which are vital and effective  
258 in analysing MPs (Hidalgo-Ruz et al., 2012). These methods can scan the complete sample filters  
259 in a short time by combining spectroscopic techniques with fast area-resolved measurements such  
260 as FPA detectors, and chemical imaging (Harrison et al., 2012). Fibres of the bigger fraction (> 750  
261  $\mu\text{m}$ ) can be spontaneously measured through Raman or FTIR spectroscopy (Käppler et al., 2016).

#### 262 **3.2.1. FTIR spectroscopy**

263 FTIR spectroscopy is the most common method of analysing surface chemical composition or  
264 specific chemical bonds of plastic particles (Hidalgo-Ruz et al., 2012). FTIR technique provides  
265 unique infrared spectra from the change in the dipole moment of chemical bond and identify MPs  
266 by comparison with known reference spectra. The unique spectra of the different bond composition  
267 of plastic in FTIR can easily discriminate plastics from other organic or inorganic particles (Löder

268 and Gerdts, 2015), and determine the composition of MPs particle and specific polymer type  
269 (Doyle et al., 2011; Harrison et al., 2012). By analysing different band patterns using the FTIR  
270 spectroscopy, physicochemical weathering of sampled MPs particles could be identified (Corcoran  
271 et al., 2009) (**Table 1**).

272 Attenuated total reflectance (ATR)-FTIR spectroscopy is employed to detect large particles  
273 fast (< 1 min) with high accuracy (Löder and Gerdts, 2015). For example, MPs particles from <500  
274  $\mu\text{m}$  to 5 mm can be determined via ATR-FTIR and transmission measurements (Huppertsberg and  
275 Knepper, 2018). The reflectance and ATR mode does not require the sample preparation step for  
276 thick and opaque MPs like the transmission mode. Moreover, in the case of irregular MP surfaces,  
277 the ATR mode produces stable spectra, compared with unstable spectra in the reflectance mode.  
278 MPs as small as the IR beam aperture (e.g., 10  $\mu\text{m}$ ) are measurable by the ATR probe.

279 Micro-FTIR mapping has been successfully applied for MPs identification with sequential  
280 measurement of IR spectra at spatially separated and user-defined points on the sample surface  
281 (Levin and Bhargava, 2005). But this technique is still very lengthy when targeting the whole  
282 sample surface at a high spatial resolution as it uses only a single detector element (Harrison et al.,  
283 2012). Only 1.4% of the particles with synthetic polymer origin were detected in sediment with  $\mu$ -  
284 FTIR (Löder and Gerdts, 2015). Surface contact analysis forming ATR-FTR offer a useful method  
285 ( $\mu$ -ATR-FTIR), which has been applied to identify MPs in environmental samples (< 50  $\mu\text{m}$ ).

286 MPs (150–250  $\mu\text{m}$ ) with larger surface areas can be detected using single beam mapping focal  
287 plane array (FPA)-based reflectance imaging method without compromising spatial resolution  
288 (Tagg et al., 2015). The simultaneous recording of several thousand spectra within an area with a  
289 single measurement and the generation of chemical images was possible with this technique  
290 (Harrison et al., 2012; Levin and Bhargava, 2005). The disadvantages of the FPA method are high  
291 instrumental cost and requiring high processing power. The advantage of this method is that no  
292 visual pre-selection is needed to automatically analysing the whole filter. This method may  
293 facilitate the measurement of large sample areas of filter i.e. whole filter surfaces in one single  
294 measurement run. Overall, FPA-micro FTIR method is fast and facilitates fully automated analysis  
295 of MPs of small size. Therefore, the FTIR spectroscopy is the most frequently used technique in  
296 MPs identification and quantification at now. This technique demonstrates a promising tool for  
297 automated MPs analysis. However, more research should be needed to progress automated analysis  
298 methodology to increase accuracy and shorten the identification time.

### 299 **3.2.2. Raman spectroscopy**

300 Raman spectroscopy is a non-destructive scattering method used to identify MPs particles in  
301 environmental samples with detailed information about the chemical structure of particles (Cole et  
302 al., 2013; Imhof et al., 2012). Molecular vibration of a sample leads to characteristic spectral  
303 fingerprint in the Raman spectra which help to identification of MPs by comparison with known  
304 reference spectra (Käppler et al., 2016). Once the laser beam (500-800 nm) striking on an object  
305 results in different frequencies of back-scattered light based on the molecular structure and atoms  
306 present, it produces a unique spectrum for each polymer. This so-called Raman shift can be detected  
307 to form substance-specific Raman spectra. This technique can be applied to detect plastic polymers  
308 within minutes by comparison with reference spectra as the plastic polymers possess characteristic  
309 Raman spectra (Löder and Gerdts, 2015). The smaller diameter of the laser beam in Raman  
310 spectroscopy allows for the identification of a broad range of size classes down to tiny plastic  
311 particles < 1  $\mu\text{m}$  (Cole et al., 2013). However, chemical additives and pigments of MPs are  
312 interfered in this method to identify polymer types (Tagg et al., 2015). In a complex MP  
313 identification, the different responses and spectra between FTIR and Raman spectroscopy can  
314 compromise each other. Confocal microscopy can be coupled with Raman spectroscopy to detect  
315 MPs particles in zooplankton samples (Cole et al., 2013). Raman spectroscopy can also be  
316 combined with Raman spectral imaging to generate spatial chemical images of a sample.

317 Raman spectroscopy has some limitations (**Table 1**) e.g. the laser used to excite the fluorescent  
318 samples cannot be analysed since it precludes the formation of interpretable Raman spectra. More  
319 research is required to explore the optimum laser wavelength for a compromise between suppressed  
320 fluorescence and low signal intensity for assessments of MPs in environmental samples. Usually,  
321 before measurements of Raman spectroscopy, a purification step of samples is needed to prevent  
322 fluorescence for precise identification of the polymer type of MPs particles (Löder and Gerdts,  
323 2015). However, Raman spectroscopy is a better analytical method to identify small-sized MPs (<  
324 20  $\mu\text{m}$ ).

### 325 **3.2.3. Pyrolysis-gas chromatography-mass spectrometry (Py-GC-MS)**

326 Py-GC-MS can be applied to identify and quantify the types of potential MPs particles with  
327 high certainty through their characteristic decomposition products (Dümichen et al., 2015). This  
328 method determines the type of polymer through analysing thermally decomposed gases from  
329 polymers, by comparing with GC-MS profiles of virgin polymers standards. It also enables to  
330 identify the organic plastic additives at a lower temperature in one run (Fries et al., 2013). The Py-

331 GC-MS technique permits a comparatively good assignment of potential MPs to polymer types,  
332 but a drawback is the complete destruction of the sample hence actual chemical composition  
333 measurement is difficult to infer. Py-GC-MS analysis method identified PA, PE, PVC, PP, PS, PET  
334 and chlorinated or chloro-sulfonated PE MPs particles from sediment samples (Dekiff et al., 2014).  
335 Regarding the size limitations of plastic particles, Py-GC-MS method is often used to analyze  
336 plastic particles with the maximum size of 1.5 mm (Fries et al., 2013). Its other limitation is long  
337 processing time per sample run (**Table 1**).

338 Py-MS has been used to overcome the limitation of Py-GC-MS method (Zhang et al., 2020b).  
339 This process enables the rapid identification of MPs (<1.2  $\mu\text{m}$ ) compared with Py-GC-MS and  
340 TGA-GC-MS, and can reduce the interference of the environmental matrix in analysing MPs.  
341 Another advantage of this method is the ability to analyse mixed polymers in one run without  
342 preselection and separation which is difficult in Py-GC-MS method. Py-MS is often used to analyze  
343 a large amount of MPs samples. MPs are identified in this process by their chemical fingerprint  
344 including characteristic ions and the ratio of characteristic ions from the mass spectra (Zhang et al.,  
345 2020b).

#### 346 **3.2.4. Atomic force microscopy (AFM)**

347 The combination of AFM with either IR or Raman spectroscopy can provide an effective  
348 method for micro and nanoparticles detection and identification which is the major challenge in  
349 MPs research. This combined method can identify polymer types through their chemical  
350 composition. Automated FTIR/Raman mapping or particle tracking with simultaneous Raman  
351 spectroscopy can resolve the problems such as missing small and transparent MPs particles (Dazzi  
352 et al., 2015). AFM-IR can generate IR absorption spectra and absorption images within the spatial  
353 resolutions of 50–100 nm (Dazzi et al., 2012). For example, 100-nm PS beads were successfully  
354 analyzed by AFM-IR combination.

355 In addition, Nile Red or NR (9-diethylamino-5H-benzo( $\alpha$ )phenoxazine-5-one) is a strong  
356 fluorescent dye which is useful for staining to identify hydrophobic MPs (Andrady, 2011). NR  
357 staining can be used as a pre-step of spectroscopic analysis and identify hidden MPs. The NR  
358 staining method presents high efficiencies for identifying <100  $\mu\text{m}$  PE, PP and PS particles mixed  
359 with high numbers of inorganic particles (Shim et al., 2016). However, co-staining of other natural  
360 materials is the main limitations in applying the NR staining method.

### 361 **3.3. Thermal analysis**

362 Thermal analysis is a destructive method which has been recently used for MPs identification  
363 (Castañeda et al., 2014; Majewsky et al., 2016). This method can easily measure the changes in the  
364 physical and chemical properties of polymers depending on their thermal stability (Dümichen et  
365 al., 2015). Thermo-gravimetric analysis can completely pyrolyze the samples at a temperature up  
366 to 600 °C (Dümichen et al., 2017). The decomposition profile of the TGA is used to measure the  
367 homogeneity of the samples. Differential scanning calorimetry (DSC) is a useful method for  
368 identification of specific polymer types by comparison with reference polymeric materials.  
369 Different plastic has different characteristics in DSC (Kalčíková et al., 2017), which help to identify  
370 MPs accurately. Due to the availability of reference materials, this method can identify specific  
371 primary MPs such as PE microbeads (Castañeda et al., 2014). The combination of TGA and DSC  
372 has been used to identify PE and PP (Majewsky et al., 2016). Due to their overlapping phase  
373 transition signal, coupled TGA-DSC method failed to identify PVC, PET, PA, polyester and  
374 polyurethane (PU) (Majewsky et al., 2016). TGA can be combined with solid-phase extraction  
375 (SPE), coupled with thermal desorption gas chromatography-mass spectrometry (TDS-GC-MS).  
376 This combined technique has added advantages of measuring larger sized particles (**Table 1**). TGA-  
377 SPE/TDS-GC-MS method is used to identifying PE, PP and PS MPs in different samples  
378 (Dümichen et al., 2015). The combination of thermo-extraction and desorption with gas  
379 chromatography-mass spectroscopy (TED-GC-MS) has been used for relatively high sample  
380 masses and enabling measurement of complex sampling matrix (Dümichen et al., 2017). TED-GC-  
381 MS has been used as a fast tool for MPs analysis and the identification of polymers in different  
382 solid samples. MPs identification in this process is through pyrolysis and analysis of the  
383 decomposition gases (Dümichen et al., 2015).

384 In summary, spectroscopy analysis should be combined with chemical analysis such as  
385 spectroscopic or thermal for MPs <1 mm. Currently, the  $\mu$ -ATR-FTIR method is recommended for  
386 the routine analysis of environmental samples. FPA-FTIR method should be used to analyze  
387 samples of known polymer types. TGA-DSC method needs further research to develop the  
388 performance for MPs identification. In comparison, spectroscopic (Raman and FTIR) and thermal  
389 analysis (Py-GC-MS, TED-GC-MS and Py-MS) methods are the most promising technologies for  
390 MPs identification and quantification. However, there is a need for significant improvement in  
391 these analytical techniques, especially in the quantification of the MPs. There is also a need to  
392 develop tools which can identify and quantify a broad range of MPs.

393  
394  
395  
396  
397  
398  
399  
400  
401  
402  
403  
404  
405

**[Table 1]**

**4. MPs interactions with other contaminants**

Many studies found that MPs interact with contaminants by the adsorption of harmful chemicals, antibiotics and metals on their surface (Li et al., 2019b; Liu et al., 2019a), hence acting as a carrier of other hazardous chemicals. These contaminants can pose a significant impact on microorganisms. Many studies observed the adsorption of organic contaminants or heavy metals on different types of MPs (Bakir et al., 2014; Hüffer and Hofmann, 2016; Wang et al., 2015). Plastics type and environmental factors (e.g. ionic strength and pH) can affect the adsorption of contaminants on MPs (Wang et al., 2015). MPs can easily sorb and concentrate toxic chemicals from the surrounding for their large specific surface area and hydrophobicity which can make them more dangerous for the natural ecosystems (Teuten et al., 2009).

#### 406 **4.1. MPs interaction with toxic organic chemicals**

407 Through adsorption, toxic contaminants can easily interact with MPs, especially those with  
408 irregular shapes (Lambert et al., 2017). The shape of MPs could influence the interaction between  
409 MPs with other contaminants or microorganisms in water (McCormick et al., 2014; Wang et al.,  
410 2018a). Wang et al. (2018a) reviewed the interaction of toxic chemicals with MPs. The large  
411 surface area to volume ratio, crystallinity, hydrophobic, physical (e.g. particle size) and chemical  
412 (e.g. functional group) properties of MPs promote their adsorption of a wide range of persistent  
413 organic contaminants (POPs), such as polychlorinated biphenyls (PCBs) or polycyclic aromatic  
414 hydrocarbons (PAHs) (**Figure A3**) (Bakir et al., 2014; Browne et al., 2013; Herbolt and Schuhen,  
415 2017). Many additives such as polybrominated diphenyl ethers (PBDE) and phthalate esters are  
416 added in the production of plastics (**Table A4**) (Cole et al., 2011). MPs may sorb much more  
417 organics than larger plastics particles because of their broad surface area (Lee et al., 2014). For  
418 example, PAHs and PCBs were detected in extracts from the plastic fragments in North Pacific  
419 Central Gyre, at concentrations of 4-249 and 1-223 ng/g, respectively (Mendoza and Jones, 2015).  
420 Similar findings from two beaches of the Portuguese coast showed that the MP pellets contained  
421 PAHs, PCBs and dichloro-diphenyl-trichloroethanes (DDTs) at 319.2 ng/g, 15.56 ng/g and 4.05  
422 ng/g, respectively (Frias et al., 2010). MPs can adsorb persistent, bio-accumulative and toxic  
423 chemicals from surrounding water and act as a unique medium to transport these chemicals into  
424 the food chain, which can lead to deadly effect in ecological habitat (Koelmans et al., 2016). On  
425 the other hand, some researcher reported that MPs are not major carriers of toxic chemicals (Gouin  
426 et al., 2011; Koelmans et al., 2013). Alimi et al. (2018) reported that PE generally exhibits a higher  
427 sorption capacity for contaminants than other MPs types, and discolored MPs adsorb more PCBs  
428 than coloured ones. Another study reported that PE particles show higher affinity with organic  
429 chemicals than other plastics like PP and PVC pellets.

#### 430 **4.2. MPs interaction with antibiotics**

431 Different antibiotics such as tetracyclines, macrolides, fluoroquinolones and sulfonamides are  
432 widely found in the aquatic environment (**Table A5**) (Jiang et al., 2011; Li et al., 2012). Antibiotics  
433 can impact the microbial community and generate resistance genes (Yang et al., 2017). In addition,  
434 antibiotics are hazardous chemicals which may affect the survival and growth of aquatic organisms  
435 (Ahmed et al., 2015). Li et al. (2018c) studied the adsorption of five antibiotics (sulfadiazine,  
436 amoxicillin, tetracycline, ciprofloxacin, trimethoprim) on five types of MPs (PE, PS, PP, PA, PVC)



437 in the freshwater and seawater systems. They reported the adsorption capacity decreased as  
438 ciprofloxacin > amoxicillin > trimethoprim > sulfadiazine > tetracycline. Due to the pore structure,  
439 PS, PP and especially PA showed higher sorption capacity than PE and PVC. Because of the  
440 formation of hydrogen bond, amoxicillin, ciprofloxacin and tetracycline are highly adsorbed on  
441 PA (Li et al., 2018c). Antibiotics are adsorbed on the MPs because of their large surface area and  
442 lipophilic nature (Guo et al., 2018; Guo and Wang, 2019). Further, the adsorption capacities of  
443 MPs varied based on their physicochemical properties such as specific surface area, the abundance  
444 of rubbery, polarity, degree of crystallinity and chemical properties such as Van der Waals  
445 interaction,  $\pi$ - $\pi$  interaction and hydrogen bonding between antibiotics and organic matters  
446 (**Figure-A4**) (Li et al., 2018c; Wang et al., 2015). Solution pH, ionic strength and salinity have a  
447 significant effect on antibiotics sorption in MPs (Guo et al., 2018). For this reason, MPs more  
448 highly adsorb antibiotic in freshwater than in seawater (Li et al., 2018c). Due to the combined  
449 pollution of antibiotics and MPs, a more toxic effect was shown in the freshwater and marine  
450 ecosystems (Li et al., 2018c). However, more research is needed on the interactions between MPs  
451 and antibiotics, in order to further evaluate their environmental risks.

#### 452 **4.3. MPs interactions with heavy metals**

453 Many studies reported that heavy metals were associated with MPs in the aquatic environment  
454 (Holmes et al., 2012; Wang et al., 2019a; Yu et al., 2019). Heavy metals (Ag, Cd, Co, Cr, Cu, Hg,  
455 Ni, Pb and Zn) can be adsorbed into MPs in freshwater at pH 6.5 (Turner and Holmes, 2015).  
456 Metals highly accumulated on a few specific MPs types such as PET, PVC, PP, low-density PE  
457 and high-density PE (Rochman et al., 2013). Cr and Cu were strongly adsorbed with an adsorption  
458 capacity of 0.297 and 0.261  $\mu\text{g/g}$ , respectively, while Ni, Co, Pb and Cd showed no obvious  
459 adsorption. It was found that adsorption capability of Ag, Cd, Co, Cu, Hg, Ni, and Pb was 0.0128  
460  $\mu\text{g/g}$ , 0.0101  $\mu\text{g/g}$ , 0.0692  $\mu\text{g/g}$ , 0.100  $\mu\text{g/g}$ , 0.170  $\mu\text{g/g}$ , 0.0166  $\mu\text{g/g}$ , and 0.191  $\mu\text{g/g}$ , respectively  
461 in freshwater (**Table A6**) (Turner and Holmes, 2015).

462 The large surface area, polarity, and organic polymer composition are the major causes of  
463 heavy metal adsorption on MPs (Ashton et al., 2010). Cations or complexed metals were directly  
464 adsorbed on the charged or neutral regions of MPs surfaces (Ashton et al., 2010). The adsorption  
465 of Ag, Cd, Co, Ni, Pb and Zn increased with increasing pH of river water (Godoy et al., 2019;  
466 Turner and Holmes, 2015). Dissolved organic matter may play a major role in the trace metal  
467 adsorption on MPs through interacting with metals or polymers (Godoy et al., 2019). Other factors

468 such as specific surface area, porosity and morphology also play an important role in metal  
469 adsorption on MPs (Godoy et al., 2019), who revealed that the adsorption mechanism of metals in  
470 MPs might be chemical adsorption due to the adsorption isotherm following the Langmuir model.  
471 Intermolecular interaction such as van der Waals interactions, hydrogen bonding interactions, and  
472 cavity formation between contaminants and polymers may influence the sorption capacity (Godoy  
473 et al., 2019). Many studies indicated that the metal adsorption kinetic was relatively fast (Godoy  
474 et al., 2019; Holmes et al., 2012). Metals are specific to adsorption on MPs, due to their different  
475 chemical structure, as demonstrated by that Pb, Cr and Zn are significantly adsorbed on the surface  
476 of PE and PVC MPs with little adsorption on PET (Godoy et al., 2019). Similarly, Pb, Zn and Co  
477 are adsorbed significantly on the surface of PE, PP and PET polymers (Rochman et al., 2013). Zn,  
478 Cu, Cr and Pb seem to have a higher affinity for plastics than Ni and Co (Godoy et al., 2019;  
479 Holmes, 2013).

480

## 481 **5. Toxicity of MP contaminants**

### 482 **5.1. Biological toxicity**

483 Studies have been conducted to evaluate the toxicity of MPs on biotic communities, and  
484 revealed higher toxic effects of MPs on the non-selective filter feeders, which mistakenly ingest  
485 MPs instead of other nourishing substances. MPs may cause various internal injuries, blockage in  
486 alimentary canals, reduction of dietary intake, or translocating into the internal circulatory system  
487 (Murray and Cowie, 2011; Shen et al., 2020). MPs ingestion may cause chronic effects, e.g. MPs  
488 from high-density polyethylene (HDPE) (particle size < 80  $\mu\text{m}$ ) were tracked in the digestive  
489 system and epithelial cells of *Mytilus edulis* with adverse impacts on the tissues and intestinal tract  
490 (Von Moos et al., 2012). Adverse effects on the fecundity of *Calanus helgolandicus* (pelagic  
491 copepods) were reported while exposed to 20- $\mu\text{m}$  MPs beads (Cole et al., 2015). The laboratory-  
492 based observations for the toxicity of MPs on biotic communities and organisms are described in  
493 **Table A7.**

494 Different types of chemical additives are incorporated with plastic compounds to obtain and  
495 improve the desired properties of the products according to consumer demands. For example,  
496 PBDEs are used to increase heat resistance, and phthalates to increase malleability, colorants,  
497 plasticizers, and flame retardants. Leachates from the waste plastic debris and their fragments may  
498 impose toxicity to a wide range of species (Lithner et al., 2009; Lithner et al., 2011). Batel et al.

499 (2016) reported that benzo[ $\alpha$ ]pyrene-loaded MPs would release contaminants retaining in gut and  
500 liver of zebrafish. Browne et al., (2013) evaluated the desorption of nonylphenol and phenanthrene  
501 from PVC. A mathematical model using equilibrium partitioning and experimental data  
502 demonstrated the transfer of contaminants from plastic to organisms (Teuten et al., 2009).

503 A positive relation was observed between MP concentration in the sediment and both uptake  
504 of plastic particles and weight loss by *A. marina* (Besseling et al., 2013). Lungworms exposed to  
505 micro-PS and PCBs showed reduced feeding capability and lost weight. In terms of immune  
506 function and oxidative status, PVC alone made lungworms >30% more susceptible to oxidative  
507 stress (Browne et al., 2013). Adverse effects of phenanthrene loaded MPs from low-density  
508 polyethylene (LDPE) and PS beads (incorporated with fluoranthene) were observed in African  
509 catfish and *Mytilus* spp., respectively (Paul-Pont et al., 2016). Plastic-sorbed chemicals were found  
510 to induce liver toxicity in Japanese medaka (*Oryzias latipes*) (Rochman et al., 2013). Luís et al.,  
511 (2015) investigated the potential influence of MPs on the short-term toxicity of chromium to  
512 juveniles of *Pomatoschistus microps*. They discovered a significant decrease of the predatory  
513 performance ( $\leq 67\%$ ) and an inhibition of acetylcholinesterase (AChE) activity ( $\leq 31\%$ ).

514 Furthermore, MPs possess high surface area that supports the particles to act as a significant  
515 vector of various contaminants such as POPs and heavy metals during the transports in the  
516 environment (Browne et al., 2013). Some of the contaminants can be retained in the tissues of the  
517 organisms, e.g. PBDE-47 and triclosan were found in the tissues of lugworm at 330 ng/g and 1250  
518  $\mu\text{g/g}$ , respectively (Browne et al., 2013). Some of the sorbed contaminants such as DDT,  
519 phenylalanine and di(2-ethylhexyl) phthalate (DEHP) can transfer from microscopic particles of  
520 PVC and PE to the benthic invertebrates (Bakir et al., 2016; Chua et al., 2014). **Table A8** listed the  
521 concentrations of contaminants in the tissues of the organisms due to the transfer of chemicals  
522 from MPs. Here it is worth mentioning that, the concentration gradient of such adhering pollutants  
523 on the MPs surfaces are often too much low to induce potential toxicities alone to the organisms.  
524 However, there are several lines of evidences that disclose the joint adverse impacts of MPs and  
525 their adhering toxic pollutants on various living organisms. In **Table A8**, some of them have been  
526 listed in detail. Overall, MPs can pose a significant risk of contaminating aquatic food chains, and  
527 potentially increase human exposure via such sources. Rather than being a vector of various POPs,  
528 MP itself pose a huge threat as a persistent pollutant to the environment, to the health of individual  
529 organisms and potentially the overall ecosystems as well.

## 530 **5.2. Impact on the aquatic ecosystems**

531 MPs in the open ocean support various microbial biofilms which are different in taxonomic  
532 composition from the microbial assemblages of the surrounding water, in heterotrophs, autotrophs,  
533 predators and symbionts. Sediments with *O. edulis* showed significantly different assemblage  
534 structures when dosed with MPs, at any density or type of plastic than controls (Green et al., 2017).  
535 Negative impacts on the producers of aquatic ecosystems (e.g. algae) have been observed, e.g.  
536 reduction of chlorophyll, growth rate and photosynthesis, with implications for all of the producer-  
537 dependent primary, secondary and tertiary components of the total ecosystem. Furthermore,  
538 individual organisms (e.g. fish) may also ingest MPs mistakenly and undergo various disorders  
539 through the direct action of MPs. The influence pathways of MPs in an ecosystem have been  
540 illustrated in **Figure 1**.

541 MPs represent a wide range of stressors to interfere in the ecosystem jointly. For instance, a  
542 mixture of additives, sorbed contaminants and copolymers may cause additive effects (Jang et al.,  
543 2020; Sait et al., 2020). In the aquatic ecosystem, MPs affect the water translucency (Sjollema et  
544 al., 2016), sedimentation (Cole et al., 2016), and thermal conductivity (Carson et al., 2011) as well  
545 as the ‘aquatic biocoenosis’. Moreover, they may act as a vector for a large number of pathogens  
546 and some other invasive species. MPs may also influence the ‘predator-prey interactions’ and the  
547 community structure by interfering with the inter- and intra-species signaling in the ecosystem  
548 (Besseling et al., 2014).

549 **[Figure 1]**

550

## 551 **6. MPs treatment technologies**

### 552 **6.1. Physical treatment**

#### 553 **6.1.1. Adsorption through biochar and activated carbon**

554 Biochar (BC) and activated carbon (AC) are extensively used as an adsorbent to treat  
555 stormwater containing MPs and NPs (Ahmed et al., 2016; Mohan et al., 2014; Mohanty et al.,  
556 2018; Sommer et al., 2018). Adsorbent surface area and porosity are two major properties for  
557 effective removal of MPs (Siipola et al., 2020). Siipola et al. (2020) reported that steam activated  
558 BC (at 800 °C) was most suitable adsorbent for MPs removal, even with relatively low surface  
559 area (200–600 m<sup>2</sup>/g). Despite the small surface area (187 m<sup>2</sup>/g) with macro-scale porosity, spruce  
560 bark AC resulted in better performance for MPs retention than pine bark ACs with surface area

561 556-603 m<sup>2</sup>/g. Activated BC effectively retained large size MPs particles, whereas 10 µm spherical  
562 microbeads did not adsorb as efficiently. Hence, meso- and macro-porosity can be very beneficial  
563 for the removal of MPs. The BC surface roughness may influence the retention of large MPs  
564 particles most likely through physical attachment. They also found that PE particles and fleece  
565 fibers were 100% retained, although the mechanism of MPs adsorption is yet to be identified  
566 (Siipola et al., 2020). Both BC and AC may act as a filter when packed in a column for MPs  
567 removal (Zhang et al., 2020a). Therefore, adsorption with AC or BC via a filtration setup is an  
568 economical process to remove MPs.

### 569 **6.1.2. Membrane processes**

570 Membrane technologies such as UF, microfiltration (MF), RO and MBR have increased the  
571 removal efficiency of MPs and NPs. In the last five years (2015-2020), the application of  
572 membrane technology in MPs removal was only reported in 13% studies, with other treatment  
573 processes accounting for 87% of studies. The combination of a porous membrane with a biological  
574 process (i.e. MBR) could enhance the rate of MPs removal from primary effluent up to 99.9%  
575 (Talvitie et al., 2017a). Membrane material, pore size, thickness and surface characteristics affect  
576 the performance of the membrane process. In membrane filtration, the major drawback is the  
577 fouling phenomena which occur by adsorption of particles on the membrane surface. Following  
578 fouling, membrane filtration performance will be decreased which resulted in higher energy cost,  
579 operation time and maintenance (Malankowska et al., 2021). Enfrin et al., (2019) briefly reviewed  
580 and revealed that MPs could interact with the membrane surface because of their intrinsic  
581 physicochemical properties such as hydrophobicity, surface charge and roughness. Nevertheless,  
582 membrane technology is highly efficient in the removal of low-molecular weight contaminants  
583 such as small MPs (<100 µm) and NPs.

#### 584 **6.1.2.1. Ultrafiltration**

585 UF is represented as the most effective and economical process. UF is a low functioning  
586 pressure (1-10 bar) driven membrane process, which can separate dissolved macromolecules and  
587 tiny suspended particles in the colloid size range whose molecular weight is higher than a few  
588 thousand Dalton from feed fluid. It permits to move solvent and low molecular weight solutes  
589 (microsolutes) through the membrane in the range of pore size 0.001-0.1 µm (Basri et al., 2011).  
590 Sun et al., (2019) reported that polyethylene (PE, ~4%-51%) is the most abundant types of polymer.  
591 Owing to the large particle size, PE particles can be fully removed through the UF membrane (Ma

592 et al., 2019). UF with a nominal size of 0.2  $\mu\text{m}$  in MBR offered 100% removal of MPs. A detailed  
593 schematic diagram of MPs behavior during the coagulation and UF processes is shown in **Figure**  
594 **A6**. Overall, UF membrane treatment can be applied to eradicate PE particles, especially with UF  
595 and coagulation combination. Pilot-scale UF-MBR hybrid treatment also showed an adequate  
596 performance to remove MPs. The pore size of the UF membrane is similar to MPs (Enfrin et al.,  
597 2019), but MPs also interact with UF membrane surface and may cause abrasion due to their  
598 irregular shape which reduces the filtration performance.

#### 599 **6.1.2.2. Reverse osmosis (RO)**

600 RO is the most common and promising process to treat wastewater (Ibrar et al., 2020). The  
601 high pressure applied in the RO membrane process enhances treatment efficiency, but may produce  
602 nano plastic particles from MPs by fragmentation (Enfrin et al., 2019). Ziajahromi et al. (2017)  
603 studied the performance of RO process to removal MPs from a WWTP in Sydney, Australia. They  
604 reported that screening and sedimentation, biological treatment, flocculation, de-chlorination  
605 processes and tertiary UF treated effluent reduced plastic particles from 2.2 MPs/L to 0.28 MPs/L,  
606 which was finally treated by RO to reach 0.21 MPs/L. Overall, the RO process is not particularly  
607 efficient for MPs removal but the application of combined UF-RO/MBR-RO processes may  
608 enhance the removal efficacy.

#### 609 **6.1.2.3. Dynamic membrane (DM) technology**

610 The development of DM technology has attracted interest. DMs can be classified into self-formed  
611 and pre-deposited (Hu et al., 2018). DM acts as a secondary membrane when particles and other  
612 foulants in wastewater are filtered by forming a cake layer in supporting membrane surface  
613 (Ersahin et al., 2012). Instead of the traditional membrane, larger pore-sized mesh or other low-  
614 cost porous materials such as non-woven fabric or woven filter cloth and stainless steel are used  
615 as a supporting layer. DM technology is more effective to removed low density (poor settling) and  
616 non-degradable MPs because of the swiftly formed secondary membrane (DM layer) with micro-  
617 particles (Li et al., 2018e). In recent years, DM technology is an effective option to filtrate MPs  
618 from synthetic wastewater under a gravity-driven operation. Low TMP, low filtrate resistance and  
619 easy cleaning are the main benefits of DM (Li et al., 2018e). Li et al., (2018e) designed and  
620 operated a lab-scale DM filtration using 90  $\mu\text{m}$  supporting mesh, and reported about 90% of MPs  
621 removal from synthetic wastewater. The DM formation is essential in DM technology to resist the  
622 higher number of microparticles in wastewater filtration. After 20 min of MPs filtration, the

623 effluent turbidity has been reduced to <1 NTU (Nephelometric Turbidity Unit) which verified the  
624 rapid formation of DM inducing better MPs removal performance.

625 In contrast to MF and UF, DM filtration process has shown lower TMP value which indicates  
626 the reduction of energy consumption (Li et al., 2018d). The most substantial TMP value observed  
627 is 16 times lesser than conventional MF at 30 kPa (Huang et al., 2017). Overall, DM technology  
628 showed excellent performance to remove micro-contaminants including MPs during wastewater  
629 treatment, and mitigated the disadvantages of membrane fouling in MBR. The combination of DM  
630 technology with coagulation or activated sludge process can be highly effective to remove micro-  
631 contaminants and MPs in wastewater treatment (Li et al., 2018d). Further research is needed to  
632 unravel the mechanism of DM layer formation.

633 Overall, membrane treatment technology is not specially designed to remove MPs efficiently,  
634 due to common issues of membrane fouling and decreasing water flux. More research should be  
635 devoted to minimize the membrane abrasion and fouling in membrane-based treatment technology.  
636 However, membrane treatment technology can be attractive if it is combined with biological  
637 process such as MBR or chemical process such as coagulation.

### 638 **6.1.3. MPs removal in primary treatment**

639 Many studies showed that conventional WWTP with primary and secondary treatment could  
640 remove MPs up to 99.9% from wastewater (**Table A9**). Most of the MPs were removed during  
641 pre-treatment stages i.e. preliminary (35-59%) and primary treatment (50-98%) (Sun et al., 2019).  
642 Sun et al., (2019) reported that higher MPs concentration ( $9 \times 10^{-4}$  - 447 particles/L) of effluent  
643 present after primary and secondary treatment process than from a tertiary treatment process (0-  
644 51 particles/L). They also reported that large particles (1-5 mm) were clearly reduced from 45%  
645 to 7% after the primary treatment process.

#### 646 **6.1.3.1. Screen**

647 Zhang et al. (2020a) used fine screen (mesh size 2.5-10 mm) to remove larger size MPs (> 2.5  
648 mm) during primary wastewater treatment, as coarse screen (mesh size 50-100 mm) and medium  
649 screen (mesh size 10-40 mm) were not capable of removing MPs. Ultrafine screen (mesh size 0.2  
650 -2 mm) was used before MBR as an alternative for primary sedimentation to prevent membrane  
651 fouling, which is the main obstacle of MBR. Therefore, the larger size of MPs (> 0.2 mm) could  
652 be removed from the wastewater, when the ultrafine screen was employed. Due to MPs' irregular  
653 shape, some larger MPs could still pass through the ultrafine screen.

### 654 6.1.3.2. Density separation

655 Density separation is the most reliable and commonly method for the separation of MPs/NPs  
656 from sediment or water samples by the density difference between materials of interest and other  
657 unwanted materials (Schwaferts et al., 2019; Wang and Wang, 2018). Mainly it is used to isolate  
658 the MPs from the environmental samples, particularly sediment samples (Hidalgo-Ruz et al., 2012).  
659 Most plastics have a density of 0.8-1.70 g/cm<sup>3</sup>, compared with 2.65 g/cm<sup>3</sup> for sand and sediment,  
660 which sink to the bottom (Wang and Wang, 2018). It is difficult to remove MPs from water samples  
661 because of their density being close to water (**Table A10**). Flotation may be used after primary  
662 sedimentation in wastewater treatment plants to improve MPs separation. The low-density MPs  
663 (e.g. PP, PE) and moderate-density MPs (e.g. PS) will float at the surface of the wastewater. In  
664 density flotation method, saturated NaCl and NaI solutions was widely used to increase the density  
665 of water from 1 g/cm<sup>3</sup> to 1.2 g/cm<sup>3</sup> and 1.8 g/cm<sup>3</sup>, respectively (Enfrin et al., 2019), so that low  
666 densities MPs float more easily even the denser PET. Few studies reported that the majority of  
667 MPs was removed in the primary treatment stage of WWTP, mainly in skimming and settling  
668 processes (Carr et al., 2016; Lares et al., 2018). Murphy et al., (2016) observed that the majority  
669 of microbeads (PE) were effectively removed via skimming. A great amount of MPs was picked  
670 up through skimming of floating debris in primary treatment since lower density MPs can float on  
671 wastewater (Carr et al., 2016; Murphy et al., 2016). Saturated NaCl solution was widely used due  
672 to low cost, widely available and environmentally benign (Li et al., 2018b; Van Cauwenberghe et  
673 al., 2015) (**Table A10**). It can be seen that fragments (PP, PE, PET) and fibers (PES, PET, PA) are  
674 the most abundant MPs in wastewater. Small amount of granular (PE), film (PP, PE, PA), and  
675 foams (PU, PVC, PA) are also found in wastewater (Enfrin et al., 2019; Lares et al., 2018; Sun et  
676 al., 2019; Talvitie et al., 2017a). PE, PP and PS were dominated plastic types in some wastewater  
677 samples (Murphy et al., 2016; Ziajahromi et al., 2017). Other studies found that polyamide (PA)  
678 was the abundant (54.8%) type of MPs in wastewater followed by PE (9.0%), PP (9.6%) and PVC  
679 (2.5%) (Liu et al., 2019c). PE is the most abundant MPs in effluents of different WWTPs  
680 (Ziajahromi et al., 2017). PES was highly detected in the final effluent of a Scottish (28%) (Murphy  
681 et al., 2016) and an Australian WWTP (67%). However, the saturated NaCl solution is less  
682 effective to separate denser MPs including PVC (1.3-1.7 g/cm<sup>3</sup>) and PET (1.4-1.6 g/cm<sup>3</sup>) (Wang  
683 and Wang, 2018). To overcome this problem, some high-density salt solution including sodium  
684 iodide (1.8 g cm<sup>-3</sup>) (Wang and Wang, 2018; Ziajahromi et al., 2017), calcium chloride (1.30-1.35



685 g/cm<sup>3</sup>) (Stolte et al., 2015), sodium polytungstate (1.4 g/cm<sup>3</sup>) (Corcoran et al., 2009; Zhao et al.,  
686 2015) and zinc chloride (1.5-1.7 g/cm<sup>3</sup>) (Maes et al., 2017; Mintenig et al., 2017) were used to  
687 increase the extraction efficiency of denser MPs. The density of brine solution has been  
688 recommended to be >1.45 g/cm<sup>3</sup> for the separation of all plastic polymers from sediments (Van  
689 Cauwenberghe et al., 2015). Majewsky et al., (2016) reported 85% and 91% recovery rates of PE  
690 and PVC respectively, when ZnCl solution was used for primary density flotation separation.  
691 Quinn et al. (2017) recommended 25% ZnBr<sub>2</sub> (1.71 g/cm<sup>3</sup>) solution for density separation which  
692 will allow the flotation of PP, LDPE, HDPE, PE, PS, PVC, PET and PA. By using saturated NaCl,  
693 NaI and ZnBr<sub>2</sub>, Quinn et al. (2017) observed higher recovery rates of 200–400 µm MPs. Moreover,  
694 ZnCl<sub>2</sub> (1.7 g/cm<sup>3</sup>) brine solution recovered 100% of large size (1-5 mm) and 95% of smaller size  
695 (< 1mm) MPs from marine sediments. They also recovered 91-99% large size (1 mm) and 40%  
696 smaller size (40-309 µm) MPs particles using density separation method. However, heavy salts or  
697 higher density solutions are often used which are costly and even hazardous (NaI) for the  
698 environment.

#### 699 **6.1.3.3. Grit chamber/primary sedimentation**

700 High-density contaminants (> 1.5 g/cm<sup>3</sup>) such as sand and cinder (density around 2.65 g/mL) are  
701 separated by grit chamber. Some MPs may be attached on the sand and then settle together in grit,  
702 and can be easily removed from the stream. The specific densities range of most of the MPs are  
703 0.8-1.7 g/mL, while PET, PVC and polytetrafluoroethylene (PTFE) are exceptional. PET and PVC  
704 have density varied from 1.38 to 1.41 or 2 g/mL, and PTFE has a density 2.10-2.30 g/mL. Density,  
705 travelling distance (D) and water flow (V) are the main parameters of grit performance. The MPs  
706 will be removed when the MPs settling time is smaller than the D/V. Blair et al. (2019) reported  
707 the lower removal efficiency (~6%) of MPs during grit. Generally, two types of grit are applied in  
708 wastewater treatment, horizontal and aerated grit chamber. In aerated grit, the lighter MPs were  
709 also eliminated along with sediment. For this reason, the aerated grit chamber showed better  
710 performance than horizontal grit chamber. Around 60% of 50 µm sized MPs in the wastewater can  
711 be removed in aerated grit (Yang et al., 2019). Liu et al. (2019c) demonstrated that large size  
712 particles could be efficiently retained from wastewater by the grit and aerated grit chamber (with  
713 6 mm sized screen). In contrast, the smaller MPs are easily absorbed by sludge and settled during  
714 the secondary treatment process. They found that after the primary treatment, the reduction rate of  
715 MPs was 40.7% when treated with the coarse and fine grit, aerated grit chamber, and primary

716 settlement tank. The combination of preliminary (screening, Grit and grease process) and primary  
717 settle treatment could improve the efficiency of removing MPs. Few studies reported that Grit  
718 chamber and primary settling tank more efficiently decreased the concentration of MPs from  
719 wastewater influent through skimming and settling process (Carr et al., 2016; Lares et al., 2018).  
720 This process is effectively used as a first stage of the WWTPs. Murphy et al., (2016) observed that  
721 preliminary (grit and grease with screen) and primary settle treatment combinedly removed 78.3%  
722 MPs from the wastewater. A very recent study showed a higher MPs removal efficiency (69–79%)  
723 by screening and grit removal process (Ziajahromi et al., 2021). Moreover, Sun et al., (2019)  
724 reported that preliminary treatment including coarse screening, fine screening and grit removal  
725 phases also shown better performance with 35-59% removal efficiency of MPs from wastewater.  
726 Consequently, preliminary treatment may play an important role in MPs removal. The grit removal  
727 and primary sedimentation process can be used as an effective pre-process in wastewater treatment,  
728 but not sufficient for small sized large scale MPs removal. In summary, primary treatment is able  
729 to remove large sized MPs in WWTP by 60-98%.

#### 730 **6.1.4. Filtration using granular media**

731 Filtration is an effective process applied after density separation process to retain the MPs  
732 particles from bulk water samples or supernatant solutions. Various types of filter media are used  
733 in filtration process, among which glass fibers are the most frequently used, followed by  
734 nitrocellulose and polycarbonate (Wang and Wang, 2018). The most frequent used pore size of  
735 filters is 0.45-1  $\mu\text{m}$ . Because of the microscopic particulates or debris in liquid, performing filter  
736 media was lowered by clog formation. To overcome this issue, ferrous salt is added to flocculate  
737 solid fraction and set a pre-filtration step with large pore size filter (Crawford and Quinn, 2017).

##### 738 **6.1.4.1. Granular activated carbon (GAC) filtration**

739 GAC is widely used to remove various micro-contaminants from wastewater through a  
740 combination of biodegradation and physical adsorption. However, GAC is mostly combined with  
741 coagulation and sedimentation processes in drinking water treatment plant to remove MPs (Wang  
742 et al., 2020b). In an advanced drinking water treatment plant in China, the overall removal  
743 efficiency of MPs reached up to 82.1-88.6% (from  $6614 \pm 1132$  to  $930 \pm 71$  MPs/L) with sand  
744 filtration, sedimentation, ozonation and GAC filtration. GAC filtration only showed 56.8–60.9%  
745 of MPs removal efficiency mainly for small size MPs (Wang et al., 2020b). Granular filtration  
746 showed high ability range to filtering out MPs from  $86.9\% \pm 4.9\%$  (removal for middle sized

747 particles, 10-20  $\mu\text{m}$ ) to 99.9%  $\pm$  0.1% (removal for large sized particles, 106-125  $\mu\text{m}$ ) (Zhang et  
748 al., 2020c). By additional GAC filtration during drinking water treatment, Pivokonsky et al., (2018)  
749 reported the average 81% and 83% removal efficiency of MPs ( $\leq$  1  $\mu\text{m}$ ) in two plants (**Table A11**).  
750 Wang et al., (2020b) showed that the particles size between 1-5  $\mu\text{m}$  was efficiently removed (73.7–  
751 98.5%) by GAC filtration process. They also revealed that GAC filtration efficiently removed  
752 different types of MPs, including fibers (38-52.1%), spheres (76.8–86.3%) and fragments (60.3–  
753 69.1%). Therefore, GAC filtration process would be an effective technology for MPs removal at  
754 low MPs concentration ranges. However, the mechanism to remove MPs in GAC is yet unclear.

#### 755 **6.1.4.2. Rapid sand filtration (RSF)**

756 Due to the low operational and maintenance cost, RSF is the most widely applied water  
757 treatment worldwide. In the presence of three layers (anthracite grains, silica sand, and gravel),  
758 RSF is accelerating to grab suspended solids either by physical adsorption or mechanical straining  
759 (**Figure A7**). RSF very efficiently removed solid particles such as MPs from wastewater in the  
760 WWTPs (Simon et al., 2018). By the hydrophilic interaction, MPs are adhered to the surface of  
761 the sand grains or adsorbed with silica grains and clogged the second layer, which reduces the  
762 performance (Enfrin et al., 2019). Previously used coagulation process can increase the production  
763 of RSF to reduce the concentration of MPs from wastewater effectively. The addition of coagulant  
764 could further improve the adherence (Talvitie et al., 2017a). Enfrin et al. (2019) reported that  
765 NPs/MPs adsorption was hard to reverse due to the presence of functional groups such as hydroxyl  
766 groups on the surface of NPs/MPs resulting in the stronger interaction (Cai et al., 2018).

767 In a pilot-scale RSF tertiary treatment in Kakolanmaki WWTP, Turku City, Finland (Talvitie  
768 et al., 2017a), RSF process removed 97% MPs (0.7-0.02 MPs/L). of all shapes and size, even the  
769 smallest size fractions of 20-100  $\mu\text{m}$ . Hidayaturrahman and Lee (2019) revealed that the removal  
770 efficiency of MPs in RSF stage was 73.8%, and the overall removal rate was 98.9% when using  
771 the RSF in tertiary stage with coagulation process. Therefore, RSF was regarded as a suitable  
772 technology for MPs removal at low MPs concentration range. These studies showed that MPs  
773 removal efficiency in RSF process is smaller than the MBR process. Thus, RSF process can be  
774 very attractive if this process is combined with other treatment process such as coagulation.  
775 Moreover, the efficacy of RSF process is more significant than dissolved air flotation, GAC and  
776 membrane filtration, demonstrating its potentials as an effective process for MPs removal in  
777 WWTP.

#### 778 **6.1.4.3. Disc filter**

779 Disc filter (DF) is a promising technology to decrease the concentration of MPs in wastewater  
780 treatment effluent. Many studies reported that micro screen filtration with DF was applied in the  
781 final polishing stage of different WWTPs in many countries (Hidayaturrehman and Lee, 2019;  
782 Talvitie et al., 2017a). DF as a full-scale tertiary treatment was used in Viikinmaki WWTP at  
783 Helsinki (Talvitie et al., 2017a). Particles were removed with sludge cake formation inside the  
784 stacks of round filter based on the physical retention in filters. To remove the sludge cake, high-  
785 pressure backwash was executed. The pore size of filter mesh which is a woven material, typically  
786 PP, polyester or polyamide, is generally 10-40  $\mu\text{m}$ . Coagulation step before this process enhanced  
787 the removal efficiency. Hydraulic retention time, water flow, membrane fouling and pore size are  
788 the main factors of DF process. Membrane fouling also occurred in disc-filters and led to more  
789 frequent high-pressure backwash (Talvitie et al., 2017a). Due to this high-pressure backwash to  
790 clean DF, the secondary membrane layer (biofilm layer on DF) can be lost and MPs easily pass  
791 through membrane. Hidayaturrehman and Lee (2019) reported an overall MPs removal efficiency  
792 of 99.1% in a WWTP with membrane disc-filter (10  $\mu\text{m}$  pore size), while the MPs removal  
793 efficiency of 79.4% at DF stage. In another study, Talvaite et al. (2017a) observed the removal  
794 performance of various types of MPs by DF in a WWTP effluent was 40-98.5%. With DF, the MPs  
795 concentration was reduced from 0.5 to 0.3 MPs/L with 10  $\mu\text{m}$  filter and from 2.0 to 0.03 MPs/L  
796 with 20  $\mu\text{m}$  filter. Generally smaller sized filters shown higher removal efficiency for MPs.  
797 However, the opposite result in this investigation was owing to the interruption of preceding  
798 treatment stages. After that, DF can effectively remove all shapes and size fractions, however, a  
799 portion of smallest size fraction (20–100  $\mu\text{m}$ ) passes through the filter entering the final effluent  
800 (Talvitie et al., 2017a). From the literature review the DF offered a comparatively low efficiency  
801 in MPs removal.

#### 802 **6.1.5. Flotation processes**

803 Wang et al. (2019b) reported that selective flotation could easily remove fine plastics. Flotation  
804 with elutriation, combined with a most commonly used hypersaline solution (density flotation) or  
805 a surfactant such as sodium dodecyl sulfate, is widely applied for the isolation of MPs from  
806 seawater samples. Carr et al. (2016) separated MPs from WWTP influent by using the elutriation  
807 method, which was developed by Claessens et al. (2013) to isolate MPs from sediments. The MPs  
808 were isolated by exploiting their inherent buoyancies based on a combination of water flow and

809 aeration. Thanks to many MPs' low density and buoyant properties, flotation is a suitable process  
810 for those MPs removal from wastewater (da Costa et al., 2016; Di and Wang, 2018). One of the  
811 popular flotation processes is dissolved air flotation (DAF), where water is aerated to forming  
812 dispersed water at high pressure after processing to promote flocs formation by coagulation, to  
813 remove the flocculent materials and remaining suspended compounds including total suspended  
814 solids, oil and grease based on their buoyancy. To augment flocculation, flocculation chemical or  
815 coagulant is added to wastewater before the flotation. DAF was examined as a full-scale tertiary  
816 treatment plant at Paroinen WWTP in the city of Hameenlinna, Southern Finland (Talvitie et al.,  
817 2017a). During flotation, low-density MPs were efficiently removed by transported to the surface  
818 of the flocs due to their buoyancy and floated to the water which were removed by skimming  
819 (Talvitie et al., 2017a). Enfrin et al., (2019) reported that this process led to 20-70  $\mu\text{m}$  sized air  
820 bubbles dispersed in water, which adhered to the suspended matter (flocs) and resulted in an air-  
821 solid complex to float to the water surface, from where it is removed by skimming. Talvitie et al.,  
822 (2017a) reported 95% removal performance of various types of MPs by DAF process from the  
823 WWTP effluent. They also indicated that dissolved air floating removed any sized of MPs with  
824 the smallest size of 20–100  $\mu\text{m}$ . However, there have been insufficient investigations to assess the  
825 performance of DAF in removing MPs under various conditions such as density, size, shape, and  
826 composition of MPs. Thus, there is a striking research gap for future study.

#### 827 **6.1.6. Magnetic separation process**

828 Among all other extraction methods such as flotation (density separation), chemical digestion  
829 and sieving or filtration, magnetic extraction is used to isolate the small-sized ( $< 20 \mu\text{m}$ ) MPs  
830 effectively. Magnetic separation is the most reliable for the separation of MPs/NPs from sediment  
831 or water samples under magnetic force, although is not suitable for MPs removal in WWTP. This  
832 method is particularly effective for small-sized MPs, because of their large surface area to volume  
833 ratio which enhances the binding affinity of MPs with Fe nanoparticles. Grbic et al. (2019) recently  
834 observed the performance of magnetic extraction for MPs isolation from seawater, freshwater and  
835 sediment. Fe nanoparticles were coated with hexadecyltrimethoxysilane (HDTMS) to create the  
836 hydrophobic characteristics for allowing binding with MPs, which helps to isolate the MPs from  
837 water under a magnetic field (**Figure A8**). The hydrophobization mechanism of Fe nanoparticles  
838 is that the methoxysilane group ( $-\text{Si}-\text{OCH}_3$ ) of HDTMS reacts with hydroxyl groups in the native  
839 oxide layer of Fe and form siloxane bonds ( $-\text{Si}-\text{O}-\text{Si}-$ ) which are covalently bonded. As a result,

840 the hydrophobic hydrocarbon tail (alkyl chain) of HDTMS in Fe nanoparticles' surface makes  
841 hydrophobic, simplifying to sorption MPs. They reported 92% and 93% removal of small-sized (<  
842 20 µm) PE and PS and large-sized (> 1 mm) MPs from seawater. The recovery rate of medium-  
843 sized (200 µm-1 mm) MPs was 84% and 78% from freshwater and sediment. Additionally,  
844 lipophilic substances (e.g. fat in fish tissues) in the sediment samples can negatively affect the  
845 removal efficiency. Therefore, MPs recovery by magnetic extraction process is particularly  
846 suitable for drinking water treatment (Grbic et al., 2019).

#### 847 **6.1.7. Comparison among physical processes**

848 Physical treatment methods can be applied to remove a wide range of MPs from water, with their  
849 average removal efficiencies represented in **Figure 2a** and their comparison summarized in **Table**  
850 **2**. A wide range of MPs can be removed through some filtration process such as GAC filtration,  
851 rapid sand filtration and disc filter. Dissolved air flotation process also attractive to remove MPs  
852 efficiently with the flocculation process. Membrane treatment such as UF, RO and DM technology  
853 can be more effective if this process is combined with MBR. UF process is effective only for PE  
854 MPs removal. Among all other membrane treatment technology, DM technology is a desirable,  
855 cost-effective and highly efficient technology to remove MPs from synthetic wastewater but still  
856 insufficient to remove large scale MPs from wastewater. On the other hand, density separation and  
857 magnetic separation are more efficient to remove MPs from sample water. The adsorption process  
858 is suitable to adsorb MPs from water but this process was not studied sufficiently. Other physical  
859 process which applied in WWTP has not been studied extensively for MPs removal. Moreover,  
860 among the physical treatment technologies, the quantitative analysis revealed that filter-based  
861 methods showed the better MPs removal efficiency than others. Therefore, filter-based treatment  
862 technologies (UF, RSF, DF, GAC filtration) achieved the best performance in eliminating MPs.  
863 Among them, RSF treatment process provides rapid and efficient removal of MPs. MPs removal  
864 through physical methods followed the order: filtration process > flotation process > adsorption  
865 process > membrane process > magnetic and density separation process. Furthermore, a more  
866 detailed characterization of MPs in different treatment technologies is needed to select the most  
867 suitable methodologies for the efficient removal of MPs from the WWTP effluents.

868 **[Figure 2]**

869 **[Table 2]**

870

## 871 **6.2. Biological treatment technologies**

872 Secondary treatment in a WWTP assembles the (i) biological treatment processes and (ii)  
873 clarification process, following the preliminary treatment (35-59% MPs removal) and the primary  
874 treatment, removes 50-98% MPs (Sun et al., 2019). Different results have been obtained while  
875 evaluating the efficacy of MPs removal from biological wastewater treatment. Murphy et al., (2016)  
876 reported < 20% MPs removal efficacy while Sun et al., (2019) reported 0.2-14% decrease in MPs  
877 from wastewater in the secondary treatment. Other researchers reported 2-55% removal of MPs in  
878 biological treatment processes (Lv et al., 2019; Yang et al., 2019). These variations in MPs removal  
879 are due to (i) the change of microbes, (ii) nature of the MPs in the wastewater (size, shape, taste,  
880 surface structure), and (iii) abiotic factors (e.g. temperature, pH). Of biological treatment processes,  
881 microbial treatments (activated sludge method, biofilm-related process), MBR technology, aerobic  
882 digestion, AD and CWs have been identified as the most widely used and efficient method for MPs  
883 removal.

### 884 **6.2.1. Microbial treatment**

#### 885 **6.2.1.1. Activated sludge process**

886 In the activated sludge process, the sludge is first completely mixed with oxygen in a reactor,  
887 which incites the microorganisms to use the sludge as their food. MPs removal in this process  
888 occurs via adsorption, degradation or aggregation. Microorganisms secrete EPS to absorb the  
889 accessible contaminants as well as MPs and then degrade them to produce desirable products.  
890 Sometimes microbes take MPs mistakenly because of the visual similarity with their nourishments  
891 and then egest them after agglomerating into flocs due to their inability to degrade or transform  
892 the MPs into harmless substances. Activated sludge process along with modified processes such  
893 as anaerobic-anoxic-oxic process, sequencing batch reactor process and oxidation ditch, achieved  
894 3.6-42.9% removal of MPs from wastewater (Carr et al., 2016; Lares et al., 2018; Mason et al.,  
895 2016). Generally, MPs are not degraded or mineralized in the activated sludge processes, but  
896 mainly removed from wastewater by the aggregation with sludge flocs. This process exhibited  
897 discrimination for different sizes and shapes of MPs. For example, Liu et al., (2019a) found that  
898 most of the MPs removed in the activated sludge process were < 300 µm in size, whereas other  
899 researchers obtained the most removal efficacy for 1-5mm sized particles (Lares et al., 2018).  
900 Zhang et al., (2020a) suggested that MPs removal variations were due to the difference in the MPs

901 shapes, with high MPs removal efficacy for fibre shaped MPs. Since the MP particles are not  
902 completely degraded or mineralized in this process and often end up in sludge flocs, this process  
903 has been followed in only some rare studies to date. Due to the incomplete disposal of the  
904 pollutants, the non-degraded MPs portion present in the sludge further easily incorporate with  
905 terrestrial ecosystems and spread again throughout the entire environment. The fate and treatment  
906 of these non-degraded MPs in sludge phase have been rarely discussed in literature. Hence, more  
907 research is urgently required on the topic.

#### 908 **6.2.1.2. Biofilm process**

909 A tire of micro-organisms growing on the surface of MPs or other carriers is defined as  
910 biofilms. In this process, biofilms undergo periodically three repeating steps of (i) growing phase,  
911 (ii) stationary phase, and (iii) peeling period. After peeling one tire of biofilm, and another new  
912 film starts to form and acts to remove more MPs and other contaminants in wastewater (Zhang et  
913 al., 2020a). Biofilm related processes remove contaminants via adsorption and fixation. Like the  
914 activated sludge process, firstly MPs are adsorbed by the microorganisms with the help of EPS,  
915 secreted by the microorganisms. MPs become an attachable carrier and support the microbes in  
916 their augmentation. After a stationary phase, biofilms start to collapse from the surface of the  
917 carrier, accumulate the contaminants and emerge with the treated water (**Figure 2d**). Biofilms also  
918 play an important role in the biodegradation of MPs.

#### 919 **6.2.1.3. Biodegradation**

920 Biodegradation is an eco-friendly process compared to other treatment methods, by converting  
921 organic substances into their fragments and eventually CO<sub>2</sub> (Ahmed et al., 2018; Zheng et al., 2005;  
922 Gu et al., 2000). Biodegradation involves depolymerisation and mineralization. The process in  
923 which complex polymers impair into their monomers, dimers, or short chains of their oligomers,  
924 which can transgress through 'bacterial membranes' and act as a source of energy and carbon, is  
925 referred to as depolymerisation. Mineralization refers to the process in which the final products  
926 are water, carbon dioxide and methane. MPs undergo microbial breakdown using the activity of  
927 exoenzymes which promote depolymerisation or assimilation by the microbial species and result  
928 in mineralization (Shah et al., 2008; Yoshida et al., 2016). MPs are usually non-biodegradable but  
929 their biodegradability can be increased in two ways, *i.e.* by reducing their polymeric chain-length  
930 to such an extent that is accessible for microbial growth, and by increasing their hydrophilicity.



931 Biodegradation initiates with the formation of biofilms, which are shoals of microorganisms  
932 attached on various biotic or abiotic surfaces and deflect the surface characteristics and entity of  
933 MPs as well as to improve the hydrophilicity of the surface (O'Toole et al., 2000). Biofilms can be  
934 made of a single microbial species but often by multiple species. The formation and development  
935 process of biofilms involves four stages (i) initial attachment, (ii) irreversible attachment, (iii)  
936 maturation, and (iv) dispersion. The formation, mode of action and MPs removal by biofilms are  
937 presented in **Figure 3**.

938

### 939 **[Figure 3]**

940

941 Such type of bio-coatings contributes to MPs removal by acting as (i) a wetting agent for MPs  
942 which modifies MPs surface properties (e.g. density) to help the particles for skimming up or  
943 settling down in wastewater (Rummel et al., 2017), and (ii) the initial step for the postero  
944 biodegradation process.

945 Various mechanisms for biodegradation of plastics and MPs have been proposed. Lucas et al.  
946 (2008) suggested that the biodegradation of plastics and their fragments occurred first by cleaving  
947 enzymatically the polymers into their oligomers and monomers which were then assimilating by  
948 microbes. Similarly, Gu and Gu (2005) proposed that the biodegradation started with the cleavage  
949 of polymer backbone or its side chains by the act of extracellular enzymes, which resulted in the  
950 formation of smaller polymer units (i.e. monomers, oligomers). In most cases, it involves the  
951 hydrolysis of amides (in polyamides), esters (in polyesters), or urethane (in PU) bonds where the  
952 extracellular enzymes act as catalysts. In addition, abiotic hydrolysis can facilitate the polymers  
953 for cleavage (Müller et al., 2001). The cleaved and simpler molecules are then absorbed and  
954 metabolized by the microorganisms. These alterations further promote biodegradation  
955 (depolymerisation and mineralization). Plastics and MPs can undergo biodegradation both in the  
956 aerobic and anaerobic environments (Klein et al., 2018), but three main conditions should be  
957 maintained:

958 i. Presence of appropriate microbes capable of depolymerizing polymer substrate and  
959 mineralizing monomeric fractions by enzymes through metabolic pathways;

960 ii. Proper environmental conditions for the biodegradation (e.g. temperature, pH, nutrients);

961       iii. Morphology of the polymer substrate should be favourable for the microbial attachments  
962 and formation of biofilms.

963       In addition, a number of biotic and abiotic factors (**Table A15**) can affect the uptake and  
964 biodegradation of MPs even though the appropriate microbes are present in the environment. For  
965 the uptake of MPs (as a food of the microbes), physiological characteristics of MPs, their size and  
966 feeding type act as the major biotic factors; while temperature and pH are abiotic factors. The  
967 polymeric substrate's molecular weight, chemical composition, hydrophobicity, size of the invaded  
968 molecules, and other environmental conditions (e.g. temperature) were reported as essential factors.  
969 Obtaining an optimum environmental condition helps to improve the effectiveness of microbial  
970 treatments for MPs removal. A combination of field-based assessments and laboratory-based trials  
971 is required to facilitate the application of microbes for the degradation of different types of plastics  
972 and MPs under different environmental conditions.

#### 973 **6.2.1.4. Microorganisms for microbial treatment**

974       Several studies have reported the ability of some microbes for the degradation and ingestion  
975 of MPs from the environment. These microorganisms were isolated from different ecosystems  
976 (e.g., terrestrial to marine environment) to evaluate their capability in plastics and MPs removal  
977 processes. A large number of marine organisms including amphipods, decapod Crustaceans  
978 (Murray and Cowie, 2011), lungworms and barnacles (Thompson et al., 2004) were reported to  
979 have the capability of ingesting MPs in several studies. Some studies demonstrated the uptake of  
980 MPs by planktons as well as the lungworm *Arenicola marina* (Besseling et al., 2013) and the blue  
981 mussel *Mytilus edulis* (Wegner et al., 2012), while others showed the MPs ingestion by gastropods  
982 and zooplanktons (e.g. *Daphnia magna*) (Eerkes-Medrano et al., 2015). Planktonic organisms like  
983 chaetognatha, copepods, larval fish, salps, zooplanktons have been detected to uptake MPs (Moore  
984 et al., 2001). Cole et al., (2013) conducted experiments on 13 zooplankton taxa which included  
985 micro-zooplanktons, holoplankton, and meroplankton. However, some negative impacts on the  
986 zooplanktons, such as reduced health function of the individuals, transfer of contaminants in the  
987 food web, were also observed. Along with zooplanktons, primary consumers such as herbivorous,  
988 bacterivorous, detritivorous, and some deposits feeders can also ingest MPs (Scherer et al., 2018).  
989 Even some proofs of MP ingestion were found for protozoa or metazoans (Scherer et al., 2018).  
990 Most importantly, a diversity of invertebrates such as crustaceans, bryozoans, polychaetes,  
991 echinoderms, and bivalves have shown ability to ingest MPs (Murray and Cowie, 2011; ).

992 Suspension feeding organisms including cladocerans, rotifers, mussels, protozoans are enthusiastic  
993 to MPs because they swallow suspended particulate matter (SPM) indiscriminately.

994 Herbivores and bacterivores ciliates (e.g. *Halteria* sp.), rotifers (e.g. *Anuraeopsis fissa*),  
995 cladocerans (e.g. *Daphnia* sp.) and flagellates (e.g. *Vorticella* sp.) were reported to feed plastic  
996 beads readily. Some aquatic larvae (*Chironomus riparius*), as well as blackworms (e.g.  
997 *Lumbriculus variegatus*) showed non-selective feeding on a wide range of sediment MPs. Bivalves  
998 swallow SPM effectively including MPs, where bivalves such as marine mussels (*Mytilus edulis*),  
999 and freshwater (e.g. *Sphaerium corneum*, *Anodonta cygnea*) exhibit size selectivity on particles  
1000 (Scherer et al., 2018). Surface-grazing gastropods (e.g. *Potamopyrgus antipodarum*, *Physella*  
1001 *acuta*) also demonstrated the ingestion of MPs through both food-associated route (e.g. *P.*  
1002 *antipodarum*) and water sediment-borne route (e.g. *P. acuta*). *Mytilus edulis* showed ingestion of  
1003 MPs in laboratory experiments. Von Moos et al., (2012) observed the intake of high-density  
1004 polyethylene, i.e. HDPE in laboratory experiments by the cells and tissues of *Mytilus edulis*  
1005 although they only ingested particles larger than 80 µm. Microbes of different genera were found  
1006 to contain such species that have the potential for plastic degradation, including *Aeromonas*,  
1007 *Arcobacter*, *Aquabacterium*, *Zymophilus* and *Pseudomonas* (McCormick et al., 2014). Various  
1008 copepods (e.g. *Calanus pacificus*, *Acartia tonsa*, copepodites, crustaceans (e.g. nauplii),  
1009 echinoderm larvae, ciliates, *Oxyrrhis marina* (dinoflagellates), and salps showed their ability to  
1010 uptake MPs, although the size of the MPs was limited to <100 µm range (Cole et al., 2013).

1011 Overall, a large number of microbes have demonstrated their capabilities for MPs uptake,  
1012 either as their comestibles or mistakenly for various factors of similarity with their nourishments.  
1013 Therefore, the uptake of MPs does not necessarily mean biodegradation of those particles.  
1014 Sometimes MPs may have been preserved in the microbial body, affecting the microbes by  
1015 interrupting their normal biological activities, or may use the microbes as media to diffuse in their  
1016 ecosystem. Another significant limitation of microbial treatment is that no such microorganism has  
1017 been found that can interact with all types of MPs under any environmental conditions. Microbes  
1018 show their selectivity according to the size of MPs, shape of the feeds, taste discrimination, mode  
1019 of feeding, variability in locus, and individual selectivity of feeds for MPs uptake. For example,  
1020 *Tenebrio molitor* Linnaeus can chew and swallow styrofoam with the help of their gut bacteria  
1021 (Yang et al., 2015) while no evidence has been found for their ability or interest to interact with  
1022 other types of synthetic polymers such as PU and PP. Some bacterial strains (e.g. *Bacillus cereus*,

1023 *Pseudomonas otitidis*) were found to degrade only PVC plastic beads (Ali et al., 2014).  
1024 *Enterobacter* sp., *Alcaligenes* sp., *Citrobacter sedlakii*, *Brevundimonas diminuta* exhibited their  
1025 ability for degradation of high impact PS beads only (Braun, 2004). *Ideonella sakaiensis* 201-F6,  
1026 a bacterial strain has been found to degrade PET into terephthalic acid and ethylene glycol, and  
1027 eventually CO<sub>2</sub> and water in aerobic condition with the help of an extracellular and an intracellular  
1028 hydrolase enzyme (Avérous and Pollet, 2012; Joo et al., 2018; Yoshida et al., 2016). *Bacillus*  
1029 *cereus*, *Pseudomonas otitidis* and *Acanthopleurobacter pedis* showed their capability of degrading  
1030 PVC while some fungi species (e.g. *Lentinus tigrinus*, *As. fumigatus*, *As. niger*, *Phanerochaete*  
1031 *chrysosporium*, *Aspergillus sydowii*) were found to degrade only the plasticized ones (Ali et al.,  
1032 2014). A significant extent of degradation in PE shopping bags was observed during the simple  
1033 contact of about 100 *Galleria mellonella* within 12 hours (Yang et al., 2015). *Comamonas*  
1034 *acidovorans* was found to degrade only PU as a sole nitrogen and carbon source (Zheng et al.,  
1035 2005). All of these microorganisms exhibit individual selectivity in MPs uptake. Moreover, the  
1036 effect of size and shape of MPs have also been noticed in some investigations. For instance, in  
1037 filter-feeding taxa, the maximum particle size for ingestion is determined by the morphology of  
1038 mouthparts of the microbes, while in cladocerans, it is determined by the opening width of the  
1039 carapace. In laboratory experiments with different sized MPs, *Daphnia cucullata* exhibited higher  
1040 filtering rates for 3 and 6 µm MPs in comparison with the 0.5 µm sized beads where *Conochilus*  
1041 *unicornis* (rotifer) filtered 3 µm MPs more frequently than 0.5 µm MPs (Agasild and Nöges, 2005).  
1042 Therefore, microbial treatment process of MPs requires the presence of suitable microorganisms,  
1043 appropriate environmental conditions, and specific types of MPs in order to obtain higher efficacy  
1044 for MPs removal.

### 1045 **6.2.2. Membrane bioreactor**

1046 MBR has gained recognition in the last few decades due to its high effluent quality, small  
1047 footprint, diversity and scalability. Sun et al. (2019) reported that MPs concentration in wastewater  
1048 was reduced from 10044 particles/L to < 450 particles/L by conventional wastewater treatment.  
1049 Some researchers showed that WWTP could remove up to 99% MPs (Carr et al., 2016;). On the  
1050 other hand, some researchers showed that wastewater treatment was not efficient for the removal  
1051 of MPs (Leslie et al., 2017). Despite the high efficiency to removal MPs by conventional  
1052 wastewater treatment processes, advanced treatment is required to decrease the number of MPs  
1053 from the final effluents (Bayo et al., 2020; Carr et al., 2016; Mintenig et al., 2017; Talvitie et al.,

1054 2017b; Yang et al., 2019). Lares et al. (2018) observed that MBR treatment technology reduced  
1055 the concentration of MPs in influent from 57600 MP/m<sup>3</sup> to 400 MP/m<sup>3</sup>, and showed a better  
1056 removal (99.4%) of MPs compared to the activated sludge treatment (98.3%). Mintenig et al. (2017)  
1057 reported 95% removal of the 20-500 µm MPs. Further, Carr et al. (2016) reported 99.9% removal  
1058 of MP from the secondary treatment processes. Michielssen et al. (2016) suggested that MBR is  
1059 more efficient than conventional activated sludge process, and removed 99.4% of MPs from  
1060 wastewater. Talvaite et al. (2017a) reported 99.9% MPs removal from wastewater by MBR. Blair  
1061 et al. (2019) demonstrated that advanced WWTP could efficiently remove MPs in the size range  
1062 60-2800 µm. The MP particles in the range of 100-200 µm in various WWTPs are widely reported  
1063 (Carr et al., 2016; Kalčíková et al., 2017; Ziajahromi et al., 2017). Talvitie et al. (2017b) revealed  
1064 70% and > 95% removal for MP in size range 20-100 µm and 20-300 µm in wastewater,  
1065 respectively. Among all shapes of MPs, fibers have been seen to dominate the effluent (average  
1066 75%) of WWTPs (Dris et al., 2016; Michielssen et al., 2016). Ruan et al. (2019) observed that the  
1067 most abundant shape of MPs was fibers in influent (55-71%) which is consistent with previous  
1068 studies (Lares et al., 2018). Murphy et al., (2016) detected no PE microbeads in the effluent of  
1069 secondary WWTP in Scotland. A few studies have shown that fibers were more effectively  
1070 removed than fragments in pre-treatment (Talvitie et al., 2017b), at the same time fragments were  
1071 more effectively removed than fibers in the secondary treatment process (Sun et al., 2019; Wang  
1072 et al., 2018a). MBR process showed a high removal capacity for all size fractions (especially the  
1073 smallest size, 20-100 µm) and all shapes of MPs from wastewater compared to other advanced  
1074 treatment (Talvitie et al., 2017a). Lares et al., (2018) reported that MBR permeate contained an  
1075 average of 0.2 fiber MP/L. However, compared with other treatment technologies, the performance  
1076 of MBR seems to be not influenced by the shape, size and composition of MPs. It can be concluded  
1077 that MBR is the most promising technology to address MP problem. Further, the effect of MPs on  
1078 membrane fouling and the degradation and/or transformation of MPs in MBR should be studied  
1079 in the future research.

### 1080 **6.2.3. Anaerobic and aerobic digestion**

1081 Aerobic digestion is the natural attenuation process of heterogeneous organic matters or  
1082 contaminants by the acts of mixed microbial communities in the presence of oxygen, and occurs  
1083 in a warm and moist environment (Mohee et al., 2008). It is also designated as an aerobic  
1084 composting process where the organic matter is decomposed via microorganisms that can sustain

1085 in the presence of oxygen. The common end products are water vapour, CO<sub>2</sub> and dark brown to  
1086 black coloured organic effluents, defined as compost. On the contrary, anaerobic digestion (AD)  
1087 process is referred to the attenuation process through which the complex organic materials are  
1088 disintegrated in the absence of oxygen as a consequence of metabolic interactions in  
1089 microorganisms to yield CO<sub>2</sub>, methane, ammonia, hydrogen sulphide, water, hydrogen, and other  
1090 compost products (Mohee et al., 2008). AD can also be defined as biogasification process that  
1091 mainly converts the biodegradable organic wastes to biogases, which can be further combusted to  
1092 produce electricity and heat or further processed to become a source of renewable energies (Van  
1093 Doren et al., 2017). AD process has drawn attention in recent years because of its capability of  
1094 destroying pathogenic microbes, reducing sludge amount, recovering sustainable energy as well  
1095 as the production of biogases at the same time. Mahon et al., (2017) analysed the effluents of seven  
1096 WWTPs after treatment by AD, lime stabilization and thermal drying, and detected comparatively  
1097 lower MPs abundances, hence in favour of accepting AD as an effective strategy for MPs  
1098 treatment. Moreover, while treating sludge without and with plastic wastes in AD process,  
1099 comparatively high production of biogases in the plastic containing batch proved the feasibility of  
1100 plastic waste removal (Mahon et al., 2017).

#### 1101 **6.2.3.1. Comparison between aerobic and anaerobic digestion for MPs treatment**

1102 The rate of digestion for some biodegradable plastic polymers has been found to be higher in  
1103 aerobic conditions than in anaerobic conditions. In experimenting with polycaprolactone,  
1104 polylactic acid, polybutadiene adipate-co-terephthalate, and starch/polycaprolactone blend in both  
1105 aerobic and anaerobic conditions, Massardier-Nageotte et al. (2006) demonstrated a higher  
1106 biodegradation in aerobic conditions than under anaerobic condition. Moreover, polycaprolactone  
1107 exhibited a significant biodegradability (35%) under aerobic condition while no biodegradation  
1108 was observed in anaerobic condition. The biodegradation of polybutylene adipate terephthalate  
1109 (Kijchavengkul et al., 2010) and ecoflex® (Witt et al., 2001) has been reported at aerobic  
1110 conditions. Therefore, aerobic digestion may be a significant method for the destruction of  
1111 biodegradable MPs. Some abiotic and biotic factors (e.g. temperature, pH, enzymes, biosurfactants)  
1112 as well some internal and external characteristics of the substrate (e.g. crystallinity, functional  
1113 groups, chain flexibility, molecular weight) can influence the rate and extent of biodegradation in  
1114 aerobic or composting conditions (Dřimal et al., 2007; Kale et al., 2007; Kijchavengkul et al.,  
1115 2010).

1116 Biodegradation of MPs has gained attention due to its performance of degrading petroleum-  
1117 based polymers (e.g. PE, PP) (Gómez and Michel, 2013). In addition, methane production from  
1118 AD has made the process more attractive as a contributor to renewable energies, and therefore  
1119 offering both environmental and economic rewards. Some of the most commonly used  
1120 biodegradable plastics reported are poly(lactic acid), poly( $\epsilon$ -caprolactone), poly( $\beta$ -  
1121 hydroxybutyrate), poly( $\beta$ -hydroxybutyrate-co-11.6%- $\beta$ -hydroxyvalerate) (Gómez and Michel,  
1122 2013; Kolstad et al., 2012).

1123 Abou-Zeid et al. (2001) tested the biodegradability of natural and synthetic polyesters, based  
1124 on the percentage weight loss. Natural polyesters such as poly- $\beta$ -hydroxybutyrate (PHB) and poly  
1125  $\beta$ -hydroxybutyrate-co-11.6%- $\beta$ -hydroxyvalerate (PHBV) exhibited complete and 60% conversion  
1126 in aerobic condition respectively, in just 9 days. Poly( $\epsilon$ -caprolactone) or PCL, a synthetic polyester  
1127 showed 30% weight loss at the same time. PHBV polymers were found to be biodegraded at about  
1128 85% extent within 45 days in the dry aerobic condition in another research (Mohee et al., 2008).  
1129 *Clostridium*, an anaerobic species was found to be capable of hydrolysing PCL and PHB polyesters  
1130 in anaerobic conditions (Abou-Zeid et al., 2001; Perz et al., 2016). Other than virgin biodegradable  
1131 polymers, some materials have come into the markets with various applications as “biological  
1132 products”, “degradable”, “green”, “compostable”, “oxo-biodegradable”, although very few studies  
1133 have been performed to determine their biodegradation extent in AD and/or composting process  
1134 (Gómez and Michel, 2013).

1135 Gomez and Michel (2013) observed 20-25% conversion of bio-based materials within 50 days  
1136 in anaerobic conditions while the additive containing conventional plastics was converted into  
1137 biogases only at 2%. They tested with (i) additives containing conventional plastics (e.g. PET  
1138 incorporated with 1% additives, PP with 2% additives), (ii) conventional plastics incorporated with  
1139 plastarch and other additives, (iii) co-polyesters with corn-based plastics, (iv) paper pulp and soy  
1140 wax mixture, and (v) plastarch, to compare their relative biodegradability during composting and  
1141 anaerobic condition. Plastarch (without additives) were found as the most biodegradable material  
1142 while conventional plastics with additives exhibited the smallest extent of biodegradation.

1143 Moreover, some synthetic non-biodegradable MPs interrupted the normal digestion of  
1144 biodegradable materials. Wang et al., (2013) investigated the influence of PE-based MPs on the  
1145 AD process, and reported a comparatively low rate of methane production from the toxicity of  
1146 polyethylene MPs (100-200 particles/g) on the normal activity of the relevant microorganisms.

1147 The abundances of hydrolytic microbes (e.g. *Rhodobacter* sp.), certain bacterial genera (relevant  
1148 to acidification) and protein utilizing *Proteiniclasticum* sp. (relevant to acetogenesis) significantly  
1149 decreased due to the exposure to PE MPs during AD (Luo et al., 2016; Wang et al., 2018b). MPs  
1150 of PET polymers inhibited hydrolysis, acetogenesis and acidogenesis in anaerobic fermentation of  
1151 biodegradable polymers via (i) shifting the microbes towards the opposite direction of hydrolysis  
1152 and acidification, (ii) raising the ROS (reactive oxygen species) level that causes the death of more  
1153 microbial cells (Wei et al., 2019b). The negative impact of PVC MPs also has been reported in AD  
1154 system (Wei et al., 2019a). Therefore, aerobic or AD processes have limited performance in  
1155 degrading non-biodegradable polymers or MPs. Biodegradable plastics that have predetermined  
1156 durability, are strongly recommended to be used as alternatives to non-biodegradable plastics.  
1157 Aerobic and AD with appropriate conditions can be applied as an effective way for their  
1158 degradation, although the maintenance cost would be higher than that of CWs.

#### 1159 **6.2.4. Constructed wetlands**

1160 CWs are familiar and natural technology for wastewater treatment with a comparatively lower  
1161 cost than other biological treatment methods. Studies have been conducted recently to investigate  
1162 the feasibility of MPs removal from wastewater using CWs (Liu et al., 2019b; Ziajahromi et al.,  
1163 2020). Vegetated wetlands are the prime locus for detaching, storing, transforming, and finally  
1164 releasing MP particles (Helcoski et al., 2020). A few studies have been conducted on the  
1165 contribution or performance of vegetated wetlands, including natural and CWs in MPs removal  
1166 from polluted water (Helcoski et al., 2020; Wang et al., 2020a). Plenty of macro-invertebrates (e.g.  
1167 snails, bristle worms, beetle) have been found in the wetlands capable of playing a significant role  
1168 in controlling the accumulation of sludges (Ouattara et al., 2009). Wang et al. (2020a) showed the  
1169 effective role of macro-invertebrates in MPs distribution throughout the wetlands. They claimed  
1170 that macro-invertebrates of the wetlands ingest a non-negligible amount of MPs. Over 90%  
1171 removal efficacy was achieved in both horizontal and vertical flow type CWs. Wang et al. (2020a)  
1172 obtained 88% average efficacy of MPs removal in CWs which is comparable with other  
1173 conventional tertiary treatment methods of WWTPs, such as biological filtration (84%), dissolved  
1174 air floatation (95%), DF (40-98.5%), MBR (99.9%), and sand filters (97.1%). 98% MPs removal  
1175 efficacy was obtained through the whole WWTP when CWs were used in its tertiary treatment  
1176 steps. Therefore, CWs can be an efficient, environmentally friendly and cost-effective tertiary  
1177 treatment process to reduce MPs significantly from wastewater. Moreover, the efficacy can be



1178 enhanced through integrating different features of different types of CWs (e.g. surface flow CWs,  
1179 subsurface flow vertical type, subsurface flow horizontal type CWs). Therefore, further combined  
1180 applications of such different CWs are strongly recommended for MPs removal from wastewater.

### 1181 **6.2.5. Comparison among different biological processes**

1182 Biological treatment methods can be applied to remove MPs at a significant extent from  
1183 different environmental conditions. A comparison between the efficiencies of different biological  
1184 treatments is represented in **Figure 2b**. MBR process and CWs showed the best efficacy among  
1185 all of them. Conventional activated sludge process also reached similar removal percentage, but  
1186 only in limited works. It is difficult to declare any exact removal percentage for microbial treatment  
1187 processes because they always fluctuate according to the microorganisms involved. On the other  
1188 hand, aerobic digestion and AD can be applied efficiently only for biodegradable MP particles. A  
1189 comparative overview of the biological methods, their advantages and drawbacks are summarized  
1190 in **Table 3**. The removal of MPs via biological methods decreased in the order: MBR > CWs >  
1191 activated sludge > microbe processes. The MBR process and CWs have potential in leading  
1192 biological methods of MPs removal.

1193

## 1194 **[Table 3]**

### 1195 **6.3. Efficacy of chemical treatment technologies of MPs**

1196 In addition to physical and biological methods, chemical methods are used in MPs treatment  
1197 purposes, either on their own e.g. chemical oxidation, advanced oxidation process, coagulation,  
1198 and EC, or used as an aid to improve the efficiencies of physical processes (e.g. density separation  
1199 process, elutriation). Several approaches e.g. catalytic, thermal, photo-oxidative and chemical  
1200 (ozonation, advanced oxidation) degradation of waste plastic materials have been discussed in  
1201 multiple studies.

#### 1202 **6.3.1. Oxidation**

1203 Several studies have reported MPs removal by oxidizing agents (e.g. ozone, hydrogen  
1204 peroxides, oxidizing acids) as well as some advanced oxidation methods (e.g. electro-Fenton  
1205 process, photo-Fenton process). Eventually, chemical oxidation aims to mineralize the polymeric  
1206 substances and convert them into CO<sub>2</sub>, water and other minerals. In some cases, radiation from  
1207 different sources (e.g. UV-vis radiation, solar energy), electric current, and ultrasound are used to  
1208 improve the efficiencies of these oxidation process (Miao et al., 2020). Advanced oxidation

1209 processes (AOPs) have gained much attention in recent years, by mineralizing the targeted  
1210 substances by producing highly reactive oxidizing species (e.g. ·OH radicals) under moderate  
1211 conditions (Klavarioti et al., 2009). Although many AOPs have been developed and implemented  
1212 for wastewater treatment purposes (Feng et al., 2011), very few of them are used for MPs treatment.  
1213 Ozonation, photo-Fenton, electro-Fenton method, photocatalytic oxidations are the most widely  
1214 used and efficient methods.

#### 1215 **6.3.1.1. Ozonation**

1216 Ozone is known as one of the most potent ancient oxidants that can react with various polymeric  
1217 substances, with the unsaturated bonds as well as the aromatic rings of the polymers (Ahmed et  
1218 al., 2017). Although very few studies have been reported on the influence of ozone in MPs  
1219 treatments, several studies have corroborated its effective influences on polymer degradations  
1220 (Chen et al., 2018), via highly reactive secondary oxidant species (e.g. hydroxyl radicals). This  
1221 process is applied either as a direct treatment method for MPs removal or used to improve the  
1222 efficacy of some conventional biological methods by facilitating biodegradation of the suitable  
1223 polymers. There is some evidence of significant changes in PE, PP, PET polymers exposed to  
1224 ozone. For instance, Chen et al., (2018) reported a high polymer degradation rate (> 90%) at 35-  
1225 45 °C by exposure to ozone. Ozonation may facilitate polymer degradation by increasing polymer  
1226 surface tension, boosting the polymer surface's adhesion properties, reducing hydrophobicity and  
1227 increasing solubility, reducing intrinsic viscosity, and decreasing melting points of the polymers  
1228 and modifying mechanical properties. Hidayaturrehman and Lee (2019) obtained the highest MPs  
1229 (particle size 1-5 µm) removal efficiency in ozonation method (89.9%) compared to other  
1230 advanced treatment methods such as membrane disc-filter (79.4%) and rapid sand filtration  
1231 (73.8%). These methods followed the other primary and secondary treatment methods and  
1232 coagulation (**Figure A9**). Moreover, enhancement of 17.2-22.2% removal efficacy was obtained  
1233 by integrating the GAC-filtration method with ozonation. There is evidence of enhancing  
1234 microbial mineralization and removal efficacy of MPs with ozonation. In a laboratory-based ozone  
1235 investigation, mineralization of  $\beta$ -<sup>14</sup>C PS films by *Penicillium variable* was found to be increased  
1236 significantly from 0.01±0.003% to 0.15±0.03% (Tian et al., 2017). These cases proved that  
1237 ozonation could be used as a useful tertiary treatment step in wastewater treatment. The main  
1238 challenge with ozonation is the high production cost of ozone and environmental issues (Ahmed  
1239 et al., 2017).

#### 1240 **6.3.1.2. Fenton process**

1241 Fenton process has been one of the most widely used AOPs for wastewater treatment. In this  
1242 process, highly reactive hydroxyl radicals are generated from the reaction of hydrogen peroxides  
1243 ( $\text{H}_2\text{O}_2$ ) and  $\text{Fe}^{2+}$ -containing heterogeneous catalysts, which further drastically oxidize the targeted  
1244 organic impurities and other contaminants to  $\text{CO}_2$ , water and mineral products. This process  
1245 enhances the oxidation power of hydrogen peroxides with the help of an iron-catalyst. As iron is a  
1246 non-toxic and abundant element of the environment, this method has become popular. MPs are  
1247 rarely affected by Fenton process. Tagg et al. (2017) examined the influence of Fenton's reagent  
1248 on PE, PP and PVC MPs, and observed no significant changes in any of the MPs even at three  
1249 different doses of  $\text{H}_2\text{O}_2$  and  $\text{Fe}^{2+}$ -containing catalysts (*i.e.*  $\text{FeSO}_4 \cdot \text{H}_2\text{O}$ ). An average of 25.49%  
1250 removal efficacy was obtained within 24 hours through this modified Fenton process. A significant  
1251 effect of pH was observed, as an enhancement of 1.69%-3.89% removal efficacy was attained by  
1252 adding sodium pyrophosphate as a chelating agent (at pH 7.95). Overall, more investigation is  
1253 required for the effective implementation of the Fenton process in MPs treatment.

#### 1254 **6.3.1.3. Electro-Fenton (EF) process**

1255 To overcome the limitations of classical Fenton process, EF method has been newly developed  
1256 in which the primary oxidant ( $\text{H}_2\text{O}_2$ ) is generated electrochemically under moderate conditions (de  
1257 Luna et al., 2012), with the rest of the process being the same as the classical Fenton process  
1258 (Ganiyu et al., 2018). Despite being a popular AOP for wastewater treatment, very few studies  
1259 have been reported on MPs treatment with this process. Miao et al. (2020) applied EF method to  
1260 degrade PVC MPs and obtained 75% dechlorination and 56% weight loss efficacy within 6 hours  
1261 of the experiment at 100 °C.  $\text{TiO}_2/\text{C}$  cathode was used to gather such higher efficiency, compared  
1262 with only 29% dechlorination efficacy using graphite cathode. As this method is effective for PVC  
1263 MPs, this can be a potential method for other chlorinated species such as 2,4-dichlorophenol, PS,  
1264 PP and PE also (Miao et al., 2020). More modification in cathodic materials and other  
1265 environmental conditions may make this method more efficient in future.

#### 1266 **6.3.1.4. Photo-Fenton process**

1267 The photo-Fenton process is another efficient AOP approach for MPs treatment, which utilizes  
1268 UV radiations to produce  $\cdot\text{OH}$  radicals from hydrogen peroxide molecules in the presence of iron  
1269 catalysts and to destruct the targeted contaminants effectively (Ahmed et al., 2017). Enhanced  
1270 production of the reactive hydroxyl radicals and a higher rate of propagation is induced in acidic

1271 or near-neutral pH conditions. In such pH range,  $\text{Fe}^{3+}$  ions from the catalyst substances form  
1272 different light absorptive hydroxyl complexes (e.g.  $[\text{Fe}(\text{OH})]^{2+}$ ,  $[\text{Fe}(\text{OH})_2]^{4+}$ ), which further  
1273 generate  $\text{Fe}^{2+}$  ions and  $\cdot\text{OH}$  radicals by utilizing the absorbed UV/visible light energy. The  
1274 produced oxidized reactive ligands then transform or mineralize the target microcontaminants  
1275 through different reactions (De la Cruz et al., 2012). This process is more rapid than the traditional  
1276 Fenton process, and recycling of  $\text{Fe}^{2+}$  ions can occur at a higher rate. Instead of hydroxyl radicals,  
1277 sometimes alkyl radicals can also be generated in the same way.  $\text{Fe}^{3+}$  ions precipitate easily by  
1278 forming amorphous ferric oxyhydroxides at higher pH range. Consequently, it becomes hard to  
1279 recycle the  $\text{Fe}^{2+}$  ions (Ahmed et al., 2017). Therefore, the entire process should be conducted at an  
1280 optimum low pH condition. While traditional Fenton process failed to induce significant changes  
1281 on MPs with the single effort of its produced  $\cdot\text{OH}$  radicals, UV irradiation was reported to enhance  
1282 the rate of this oxidative degradation. Feng et al., (2011) reported over 99% mineralization of  
1283 cross-linked sulfonated PS foams within only 250 minutes in photoassisted Fenton process.  
1284 Despite being one of the widely used AOPs in wastewater treatment, research on this method for  
1285 MPs treatment purpose is limited and should be strengthened.

### 1286 **6.3.2. Photolysis and photocatalytic degradation**

1287 Photolysis has been used in wastewater treatment processes to remove various contaminants  
1288 (Ahmed et al., 2017). There is evidence that MPs can also be decomposed into the ultimate end  
1289 products (water and  $\text{CO}_2$ ) if they are irradiated to UV-radiations for a particular length of time (Tao  
1290 et al., 2019). The major problem is the lower rate of degradation in the natural weathering process.  
1291 Brandon et al. (2016) experimented with two types of MPs (PE and PP) under natural weathering  
1292 conditions, and observed only a slight change with the help of FTIR analysis after 3 years. The  
1293 prolonged degradation of MP particles resulted in hydroxyl, carbonyl groups, C=O bonds, and  
1294 other minor products. Among the very few studies on the degradation mechanism, luminance and  
1295 salinity of the corresponding media have been found to affect the degradation of MPs in aquatic  
1296 environments (Padervand et al., 2020). Enhanced degradation of MPs was observed in the artificial  
1297 seawater system where no significant deterioration was sighted at the same periods when the MPs  
1298 were exposed to UV-radiations alone. Both SEM images and FTIR analysis of the initial and end  
1299 materials confirmed that the salinity of media could facilitate the photodegradation of MPs  
1300 (Karlsson et al., 2018). Additional chemicals are being used to enhance the photo-degradation rate  
1301 of MPs, as photocatalysis has gained popularity during the past few years for MPs treatment.

1302 Photocatalysis process initiates with the excitation of the corresponding photocatalyst through  
1303 the absorption of an appropriate amount of energy from a definite light source. This photo-  
1304 excitation results in the generation of ‘exciton pair’s which further propagate other reactive species  
1305 (e.g. hydroxyl radicals, superoxides) by interacting with moisture or water molecules from the  
1306 surrounding environment (figure 4b). The highly reactive radicals then oxidize various organic  
1307 contaminants, including polymers effectively (Ali et al., 2016; Padervand et al., 2020; Qi et al.,  
1308 2017). It has been proposed to be an energy-efficient, durable, and cost-effective process for  
1309 polymer degradation (Tofa et al., 2019).

1310 Different mechanisms of MPs photocatalysis have been proposed, among which hydroxyl  
1311 radicals promoting degradation process (Liang et al., 2013) have been mostly granted, and  
1312 confirmed via FTIR analysis of the generated intermediated compounds during the degradation  
1313 process. Various nanostructured semiconductors are used as photocatalysts to generate the desired  
1314 reactive species, of which metal oxide nano-materials having semiconducting properties with a  
1315 particular bandgap (ZnO, TiO<sub>2</sub>) are most appropriate. As photocatalysis is a type of surface  
1316 phenomenon, nano-sized materials are especially used because of their high surface to volume  
1317 ratio. ZnO nanoparticles are considered as one of the most promising photocatalysts due to their  
1318 appropriate bandgap for catalysis (3.37 eV), high redox potential, non-toxicity, excellent electron  
1319 mobility and optical properties, ease of synthesis, and flexibility in sizes and shapes to be formed  
1320 (Qi et al., 2017). Photocatalytic degradation of LDPE based MPs was investigated through  
1321 heterogeneous rod-like zinc oxide nano-catalysts (Tofa et al., 2019).

1322 Photo-active micromotors have gained massive attention during recent years due to their  
1323 extensive capability for environmental contaminants remediation and water purification  
1324 (Eskandarloo et al., 2017; Zhang et al., 2018). Recently, several studies have been conducted on  
1325 the degradation capability and mechanism of TiO<sub>2</sub>-based nano-devices and micromotors in  
1326 photocatalysis of MPs. MPs have been treated in the photocatalytic process by using Au-decorated  
1327 TiO<sub>2</sub>-micromotors to make this process more efficient (Wang et al., 2019c). Protein-based N-TiO<sub>2</sub>  
1328 photocatalysts were also reported to hold the potential of degrading MPs in both aqueous and solid  
1329 phases. Ariza-Tarazona et al. (2019) obtained 6.40% mass loss of high-density polyethylene MPs  
1330 within 18 hours, while irradiated it with visible light radiations in the presence of N-TiO<sub>2</sub>  
1331 photocatalysts. The catalyst surface area, as well as the extent and nature of interactions between  
1332 the MPs and catalyst surface, influenced the removal efficacy significantly (Ariza-Tarazona et al.,

1333 2019). As this method is very new in MPs treatment compared to the other conventional and  
1334 advanced treatment methods, future research is strongly required to obtain more effective and  
1335 efficient advanced photocatalysts, so that the method can be successfully applied for MPs  
1336 treatment in real wastewater..

### 1337 **6.3.3. Coagulation**

1338 As MP particles are tiny in size (diameter < 5 mm), it is highly challenging to separate them  
1339 through filtration processes continuously. Difficulties like membrane fouling make them often  
1340 inefficient and discontinuous, hence, pre-treatment by coagulation or flocculation before filtration  
1341 process will improve filtration efficiency. Coagulation and agglomeration processes are widely  
1342 used in WWTPs worldwide to improve MPs removal efficacy through the formation of enlarged  
1343 contaminant particles that are more easily separated than their discrete tiny particles (Lee et al.,  
1344 2012). Of various coagulating agents used to coagulate and agglomerate the MP particles, iron-  
1345 based and aluminium-based salts are used widely. A variety of flocculating agents (e.g.  $\text{Fe}_2(\text{SO}_4)_3$ )  
1346 have been found to be capable of aggregating suspended particulate matters and forming flocs that  
1347 can impact positively on MPs removal in WWTPs during secondary treatments (Murphy et al.,  
1348 2016). Ariza-Tarazona et al., (2019) investigated the role of Fe and Al-based salts as coagulating  
1349 agents in removal of polyethylene-based MPs, and found that  $\text{Al}^{3+}$  ions showed better performances  
1350 in comparison with  $\text{Fe}^{3+}$  ions. Ma et al., (2019) also noticed a comparatively better performance  
1351 of Al-based salts than the Fe-based coagulants, with  $36.89 \pm 1.06\%$  PE MPs being removed  
1352 effectively by using  $\text{AlCl}_3 \cdot 6\text{H}_2\text{O}$  salts as coagulating agents. A higher dosage of  $\text{Al}^{3+}$  was  
1353 implemented (15 mmol/L at pH 7.0) to attain such higher removal efficacy than 8.0% removal  
1354 with normal doses. They also tested another flocculating agent, polyacrylamide (PAM) for the  
1355 same polyethylene MPs removal, and interestingly, the removal efficacy reached 85-90% and 50-  
1356 60%, respectively when PAM (3-15 mg/L dosage) was combined with Fe-based (2 mmol/L dosage)  
1357 and Al-based coagulants (5 mmol/L dosage). Ariza-Tarazona et al., (2019) also noticed the effect  
1358 of PAM in MPs treatment, obtaining enhanced MPs removal efficacy (MPs diameter < 0.5 mm)  
1359 from 25.83% to 61.19% by using 15 mg/L PAM.

1360 The size of the MPs is an important factor in the formation and growth of flocs. PAM showed  
1361 better performance in the removal of small MPs in comparison with that of the larger particles.  
1362 Ariza-Tarazona et al., (2019) reported the removal of MPs was raised as the concentration of the  
1363 coagulant ( $\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$ ) was increased, especially for the smaller MPs (diameter < 0.5 mm). Ma

1364 et al., (2019) also determined the effect of coagulant concentration on the removal of MPs, and  
1365 found that  $13.27 \pm 2.19\%$  PE MPs was removed using 2 mmol/L  $\text{FeCl}_3 \cdot \text{H}_2\text{O}$  coagulant which was  
1366 decreased to  $6.71 \pm 1.26\%$  with 0.02 mmol/L of the same coagulant. Therefore, selecting  
1367 appropriate coagulants and dosage is vital to facilitate the MPs treatment more effectively.

1368 Another new sustainable approach has been proposed recently for the agglomeration-based  
1369 removal of MPs (Herbort et al., 2018). This process initiates with the synthesis of active molecular  
1370 precursors at an inert atmosphere which are further applied to form 'bio-inspired' alkoxy-silyl  
1371 bonds through sol-gel reactions. MPs can become adhered to one another and form comparatively  
1372 large-sized 3D agglomerates (666% larger particles) through the sol-gel process, which are quickly  
1373 removed via cost-efficient filtration methods (Herbort et al., 2018) (Figure A10).

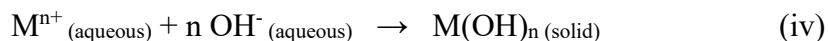
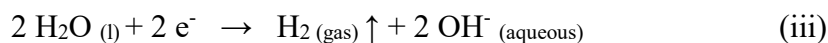
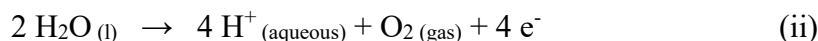
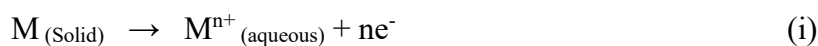
1374 Bio-inspired materials are deliberately prepared having a good extraction ability for a diverse  
1375 range of macro- and micro-sized substances. Their extraction capability may vary according to  
1376 their molecular structure, the morphology of the precursors and the corresponding hybrid silica-  
1377 gels. Herbort et al. (2018) used four different types of hybrid silica-gel from four separately  
1378 synthesized precursors to remove a commercially available polyethylene in laboratory experiments,  
1379 and obtained  $> 70\%$  removal efficacy. Among them,  $\text{N}^1, \text{N}^4$ -bis((3-(trimethoxysilyl) propyl  
1380 carbamoyl) terephthalamide was found as the most efficient precursor ( $> 95\%$  extraction capacity)  
1381 to bind the MPs.

1382 Overall, coagulation or agglomeration of MPs can be an effective step in MPs treatment  
1383 process to enhance the efficacy of WWTPs. Moreover, it can play a significant role to overcome  
1384 the fouling problems of the membrane-based treatments. The efficacy of MPs removal is  
1385 accelerated at high concentration of the coagulant, high pH, and the small size of MPs. Anionic  
1386 PAM exhibited better performance compared to that of the cationic PAM in some studies due to  
1387 the denser and facile formation of metal-based MPs trapping flocs (*e.g.* Fe-based flocs). Al-based  
1388 salts showed better performance in comparison with the Fe-based coagulants Efficacy of MPs  
1389 removal varies with pH, chemical compositions and concentration of the media, and types of the  
1390 coagulation agents. In addition, alkoxy-silyl induced agglomeration method can be potentially  
1391 effective for MPs treatment.

#### 1392 **6.3.4. Electrocoagulation (EC)**

1393 EC, electroflotation, and electro-decantation are cost-effective methods compared to the  
1394 microbial related methods to remove contaminants from wastewater (Garcia-Segura et al., 2017).

1395 EC is the advanced technology of chemical coagulation process, which is comparatively cost-  
 1396 effective, energy-efficient, and amenable to automation, with the help of electrodes. EC process  
 1397 initiates with the liberation of metal ions from the sacrificial electrodes via the electrolysis process.  
 1398 Primary electrodes are connected to the external electric source and are monopolar (anode and  
 1399 cathode). For convenience, the sacrificial electrodes are made bipolar which break down to form  
 1400 metal ions during the electrolysis. Often aluminium and iron is used as sacrificial electrodes due  
 1401 to the capability of  $\text{Al}^{3+}$  and  $\text{Fe}^{3+}$  ions to act as a coagulating agent effectively. The liberated metals  
 1402 ions in the water stream react with hydroxyl ions of the media to form metal hydroxide coagulants.  
 1403 These coagulants break up the emulsion or colloids and make changes in stabilization of the  
 1404 surface charges of suspended MPs which permits them to become close enough to each other and  
 1405 thus attached via Van der Waals forces (Akbal and Camcı, 2011). The coagulation agents trap the  
 1406 suspended solid particles (e.g. MPs, particulates) by forming a sludge blanket. The liberated  
 1407 hydrogen gas (produced from the electrolysis process) helps to elevate the resultant sludge flocs  
 1408 above the water surface, which is referred to as stable flocs (Moussa et al., 2017) (**Figure 4a**). The  
 1409 following anodic and cathodic reactions co-occur throughout the process:



1414 EC has been reported to be effective over pH 3-10 (Padervand et al., 2020), which makes it  
 1415 more attractive to be an effective pathway for MPs removal from many types of wastewater and  
 1416 their effluents without adding other chemicals. Also, alteration in current density rarely affects the  
 1417 efficacy of MPs removal, which is above 90% (Perren et al., 2018). In the presence of 0.2 g/L of  
 1418 NaCl and pH 7.5, 99.24% microbeads were removed using this method (Padervand et al., 2020).  
 1419 Recently, Akarsu and Deniz (2020) obtained up to 98% MPs removal from laundry wastewater  
 1420 using Fe-Al electrode in this process within only 60 minutes. Despite having such cost-effective  
 1421 and efficient performance in MPs treatments, EC process is constrained with some operational  
 1422 drawbacks such as requirements for continuous replacement of sacrificial anodes, cathodic  
 1423 passivation and high cost of power supply. The development of more viable anodes and future  
 1424 research on operational modifications to avoid the cathodic passivation are required to overcome  
 1425 these limitations.



1426  
1427 **[Figure 4]**  
1428

1429 **6.3.5. Comparison of chemical treatment technologies**

1430 Overall, chemical treatment methods are applied to enhance the MPs removal efficacy of  
1431 WWTPs significantly. An overview of each of the methods, their advantages, obtained efficiencies  
1432 and drawbacks is represented in **Table 4**. MPs were better removed via EC and ozonation process.  
1433 Efficacy of sol-gel agglomeration also reached similar performance, but which varied with the  
1434 implemented bio-inspired materials. Comparatively very limited research has been conducted on  
1435 electro-Fenton and photo-Fenton processes for MPs removal. Photocatalytic degradation is a  
1436 potential strategy but very few WWTPs have implemented this method so far due to their  
1437 miscellaneous drawbacks (**Table 4**). Acid-alkali pre-treatment method is still required to be  
1438 developed to be more cost-efficient. The average efficiencies of MPs removal obtained with  
1439 chemical methods followed the order: photo-Fenton process > electro-coagulation > ozonation >  
1440 electro-Fenton process > sol-gel agglomeration > coagulation > modified Fenton process.

1441 To date, the highest removal efficiencies obtained from different chemical methods are  
1442 presented in **Figure 2c** for a comparative overview (**Table A16**). Unfortunately, none of these  
1443 treatment strategies can remove MPs from contaminated sludge and wastewater when  
1444 implemented alone without any other physical or biological treatment strategies. Moreover, by-  
1445 products, as well as some secondary sludge produced in some methods such as coagulation, EC,  
1446 sol-gel agglomeration require further treatment.

1447  
1448 **[Table 4]**  
1449

1450 **6.4. Pyrolysis and co-pyrolysis of MPs**

1451 Pyrolysis and gasification processes are designated as thermo-chemical treatment technologies  
1452 which allow the thermal decomposition of plastic wastes and other biomass particles as well as  
1453 fuel production from them. In this process, the solid carbonaceous materials (e.g. plastic trash,  
1454 solid organic contaminants, biomass) are converted into different gaseous products (having  
1455 different heating values), which are also referred to as syngas. Composition of the produced gases  
1456 may vary due to the applied temperature range and characteristics of original materials. Some

1457 common fractions of the end gaseous products are H<sub>2</sub>, CO<sub>2</sub>, CO, CH<sub>4</sub>, and some hydrocarbons  
1458 (Burra and Gupta, 2018a). Plastic wastes with long polymeric chains can be degraded into  
1459 oligomers during various types of pyrolysis techniques such as catalytic pyrolysis, thermal  
1460 pyrolysis, and microwave aided pyrolysis (Sun et al., 2019). A requirement of high temperature to  
1461 break down the polymeric chains was one of the most significant challenges for this process. This  
1462 limitation of higher temperature requirements can be solved through co-pyrolysis of plastic wastes  
1463 along with other biomass or biogenic materials. Also, it may provide a feasible pathway to treat  
1464 the MPs of sewage-sludge where they are mixed up with other contaminants (Burra and Gupta,  
1465 2018a; Jin et al., 2019). Different mass fractions of different type plastic wastes (e.g. polyethene  
1466 terephthalate, polypropylene, polycarbonate) were treated in co-pyrolysis process with pinewood  
1467 and enhanced production of syngas and hydrogen fuel were observed in several studies (Ahmed et  
1468 al., 2011; Burra and Gupta, 2018b; Pinto et al., 2002). Furthermore, co-pyrolysis of polypropylene  
1469 with cellulose exhibited a reduction of the pyrolysis activation energy (Burra and Gupta, 2018a).

1470 Although some studies have been conducted on thermal treatment of plastic wastes, there has  
1471 been a negligible amount of work on MPs. One of the most probable reasons is the requirements  
1472 of isolation of MPs from the environment first with excess pre-treatment steps before the main  
1473 thermal treatments. As there is evidence that co-pyrolysis of plastic wastes with biogenic materials  
1474 requires even comparatively lower heat extent, therefore, co-pyrolysis may become a cost-  
1475 effective process for MPs removal in future along with another beneficial side of fuel production  
1476 at the same time without requirements for excessive accessories and costs. More investigations on  
1477 thermal treatment are still required to establish this process entirely for efficient MPs treatments.

## 1478 **6.5. Efficacy of hybrid treatment technologies of MPs**

1479 MPs-targeted wastewater treatment technology is not fully developed, and no specific  
1480 treatment process aimed at MPs removal has been applied in full-scale WWTP yet. In wastewater  
1481 treatment, significant improvements have been achieved during the last few years in the  
1482 application of a variety of hybrid treatment technologies, which consist of the combination of  
1483 different treatment technologies to obtain the maximum MPs removal efficacies (Ahmed et al.,  
1484 2017). Recently, hybrid systems have been widely used for the removal of MPs from water and  
1485 wastewater. Talvitie et al., (2017a) observed the removal efficiency of various types of MPs from  
1486 WWTP effluents by advanced treatment technologies including DF, RSF, DAF and MBR. They  
1487 concluded that MBR-UF hybrid process treating primary effluent could remove 99.9% of MPs of

1488 almost all sized ( $> 20 \mu\text{m}$ ) and all shapes, and showed that DF-coagulation and flocculation-DAF  
1489 hybrid processes removed 98.5% and 95% MPs respectively.

1490 Porous membranes and biological process combination could enhance the MPs removal  
1491 efficiency up to 99.9% (Talvitie et al., 2017a). Lares et al., (2018) observed the performance of a  
1492 municipal WWTP operating based on a combination of primary treatment and pilot MBR  
1493 technology for the removal of MPs. This hybrid system was found to achieve high retention  
1494 capacity of MPs over 98.3%. Similarly, the combination of MBR-RO is an effective advanced  
1495 technology for the wastewater treatment to produce high-quality water (Dolar et al., 2012; Qin et  
1496 al., 2006). RO influences the performances of MBR in wastewater treatment, and RO performance  
1497 is commonly impacted by membrane fouling from inorganic, organic and biological fouling  
1498 (Farias et al., 2014;). MBR-based anaerobic/anoxic/aerobic (A/A/O-MBR) systems effectively  
1499 removed MPs from influent by trapped in sludge and block into permeate (effluent) through micro-  
1500 membrane (pore size  $< 0.1 \mu\text{m}$ ) filtration (Lv et al., 2019). This structure could eliminate  
1501 necessarily all MPs from wastewater. A combination of sorption and filtration methodologies with  
1502 biological and sedimentation processed showed an excellent efficiency for the treatment of MPs  
1503 containing wastewater. MBR with other advanced physical and chemical treatment showed higher  
1504 MPs removal efficiency in the WWTP. While MBR coupled with sorption and filtration process  
1505 exhibited high removal percentage of MPs from the influent of water treatment plant (Padervand  
1506 et al., 2020). MBR based hybrid systems are more effective for the removal of high MPs  
1507 concentrate influent.

1508 EC and agglomeration, coupled with additional filtration stage, showed effective separation of  
1509 MPs from water. Ma et al., (2019) demonstrated that the coagulation process could remove more  
1510 than 36.89% of MPs from water. Coagulation with sedimentation could enhance the removal  
1511 efficiency up to 81.6% MPs of the secondary sediment effluent (Hidayaturrehman and Lee, 2019).  
1512 Chemical coagulation treatment is the most widely used process to combine with the physical  
1513 process including UF, DF, RSF, GAC and RO process to reduce the fouling problem or enhance  
1514 the removal performance. Coagulation is often coupled with rapid sand filtration, membrane  
1515 filtration and ozone oxidation in tertiary treatment of wastewater treatment plant  
1516 (Hidayaturrehman and Lee, 2019). This hybrid treatment, including coagulation-RSF, coagulation-  
1517 ozonation and coagulation-DF achieved the removal efficiency of MPs by 84.8%, 95.2% and  
1518 96.2%, respectively. Compared with coagulation (47.1-81.6% removal) and rapid sand filtration

1519 (73% MPs removal), membrane filtration (79.4% MPs removal) and ozone oxidation (89.9% MPs  
1520 removal) showed better performance to remove MPs. Small MPs are trapped with flocs which  
1521 formed in coagulation and stopped during filtration. Primary and secondary treated three different  
1522 WWTP in Daegu, South Korea was finally treated through different tertiary treatment process  
1523 combined with coagulation (Hidayaturrehman and Lee, 2019). They showed that MPs' overall  
1524 removal rate in different WWTPs is 99.2%, 99.1% and 98.9%, when using the ozonation,  
1525 membrane disc filter, and RSF in the tertiary stage. UF process coupled with coagulation as a pre-  
1526 treatment which is one of the main water treatment technology to remove organic contaminants  
1527 from wastewater and produce high-quality effluent in current water treatment plant (Park et al.,  
1528 2017; ). UF-Coagulation hybrid system is not perfectly designed for MPs removal from wastewater  
1529 (Mason et al., 2016). Recently Ma et al., (2019) observed the performance of UF-coagulation to  
1530 remove PE MPs for potential application in drinking water treatment. After coagulation slight  
1531 membrane fouling was induced due to the formation of loose cake layer by flocs, although PE  
1532 particles were completely eliminated during drinking water treatment. However, after coagulation  
1533 with PE particles (especially small size), membrane fouling was gently eased to increase the action  
1534 of UF membrane (Ma et al., 2019).

1535 Coagulation/flocculation combined with sedimentation (CFS) and granular filtration is applied  
1536 to MPs (180 nm – 125 µm) removal in drinking water treatment (Zhang et al., 2020c), but with  
1537 unsatisfactory removal. MBRs combined the biological activated sludge process with membrane  
1538 separation provided MPs free effluent. Many studies have shown MBR hybrid systems to be more  
1539 effective in the removal MPs from water up to 99.9% (Lares et al., 2018; Talvitie et al., 2017a).  
1540 GAC filtration could effectively remove contaminants such as MPs through a synergistic  
1541 combination of physical adsorption and biodegradation from the effluent of ozonation (Wang et  
1542 al., 2020b). It is commonly coupled with ozonation process to remove some emerging  
1543 contaminants (Fu et al., 2019; Li et al., 2018a) and larger molecular weight matter is converted  
1544 into a small fraction to enhancing the biodegradability of the influent of GAC filter during drinking  
1545 water treatment (Ross et al., 2019). When ozonation combined with GAC filtration, it increases  
1546 the removal efficiency of MPs by approximately 17.2-22.2% (Wang et al., 2020b). So ozonation-  
1547 GAC filtration couple process reduced 74-83.1% MPs from the final effluent of drinking water  
1548 treatment plant. GAC filtration also coupled with sand filtration and sedimentation/flotation  
1549 process for treated drinking water treatment plant (Pivokonsky et al., 2018).

1550 Coagulation/flocculation, sedimentation, sand and GAC filtration hybrid process removed 81%  
1551 MPs and coagulation/flocculation, flotation, sand filtration and GAC filtration hybrid treatment  
1552 reduce the MPs 83% in drinking water treatment.

1553 In an advanced drinking water treatment plant (coagulation+sedimentation+sand  
1554 filtration+ozonation+GAC), the overall MPs removal efficiency was 82.1-88.6%, of which 82.9-  
1555 87.5% 73.1–88.9% fibers fragments and 89.1–92.7% spheres were removed (Wang et al., 2020b).  
1556 With GAC filtration combined with coagulation and sedimentation only, the MPs removal was  
1557 reduced to 56.8-60.9% where 1-5  $\mu\text{m}$  MPs removal was 73.7-98.5%. On the other hand, the  
1558 combination of coagulation and sedimentation removed only fiber types MPs at 40.5-54.5%. In  
1559 the coagulation/sedimentation process, it was found that the larger size MPs had a higher removal  
1560 efficiency. MPs  $> 10 \mu\text{m}$  were almost completely removed, followed by the removal efficiency of  
1561 44.9–75.0% for 5–10  $\mu\text{m}$  in this process. Despite the high removal efficiency of MPs about 99%,  
1562 the conventional WWTP with primary and secondary treatment is not specially designed to  
1563 improve the quality of final effluent. Different technologies could be combined before application  
1564 to remove MPs in the WWTPs. Among them, a few physical and chemical treatment showed better  
1565 performance when they use as combined with another process. The effective removal rates of MPs  
1566 in tertiary stage of different WWTPs can be followed as hybrid MBR with RO or UF  $>$  coagulation-  
1567 membrane disc-filter  $>$  coagulation- ozonation  $>$  Flocculation-DAF  $>$  constructed wetland  $>$   
1568 coagulation-RSF  $>$  ozonation-GAC filtration  $>$  coagulation-sedimentation. In summary, the  
1569 combination of MBR with physical treatment such as RO/UF/NF has been found highly efficient  
1570 in the removal of a wide range of MPs. CWs based hybrid treatment was found highly efficient,  
1571 environmentally friendly and cost-effective. Moreover, coagulation with  
1572 ozonation/GAC/DAF/RSF/ filtration processes are also more cost-effective than MBR based  
1573 hybrid treatment.

1574

## 1575 **7. Recommendations for future work**

1576 Although significant progress has been made in MPs research in terms of their analysis,  
1577 interactions with other contaminants, toxicological effects, and removal by different treatment  
1578 technologies, there are still many gaps. Future research directions on MPs are suggested as follows:

1579 ❖ More reliable methods for the detection and quantitative analysis of MPs are urgently  
1580 needed to cover a wide range of MPs in the aquatic environment.

- 1581 ❖ More studies are needed to determine the toxicological effects of MPs.
- 1582 ❖ In membrane-based treatment, more research is needed to minimize the membrane abrasion  
1583 and fouling to increase the performance in MPs removal.
- 1584 ❖ Application of dynamic membrane technology in MBR treatment should be promoted to  
1585 improve MPs removal performance.
- 1586 ❖ The degradation and/or transformation of MPs in MBR should be investigated.
- 1587 ❖ More research is needed to isolate and amplify the number of MPs-degrading microbes for  
1588 their targeted applications.
- 1589 ❖ Research on MPs removal from the sludge phase produced from biological treatment  
1590 methods is urgently needed.
- 1591 ❖ CWs should be further developed for application in MPs removal.
- 1592 ❖ New materials and cathodic materials should be synthesized for efficient removal of MPs  
1593 in Fenton and electro-Fenton processes.
- 1594 ❖ For commercial-scale photocatalysis treatment plants, the utilization of solar energy should  
1595 be actively explored instead of UV irradiations.
- 1596 ❖ More effective and efficient photo-catalysts are to be synthesized for the photocatalytic  
1597 degradation for MPs removal.
- 1598 ❖ More bio-inspired materials and their cost-efficient synthesis routes for sol-gel  
1599 agglomeration method should be sought.
- 1600 ❖ Development of more viable anodes for EC method is needed.
- 1601 ❖ Hybrid treatments are needed to be specially designed to remove MPs.

1602

## 1603 **8. Conclusions**

1604 A detailed review of MPs abundance and sources in the aquatic environment, their effective  
1605 identification and analytical methods, interactions with major contaminants, potential  
1606 toxicological effects on living organisms, and removal technologies from wastewater have been  
1607 provided. MPs are widely detected in freshwater and marine environments such as water,  
1608 sediments and organisms. A comparative evaluation of different MPs identification and analysis  
1609 methods in environmental samples is presented by describing their applications, advantages, and  
1610 drawbacks. Py-GC-MS, Py-MS methods, Raman spectroscopy, and FT-IR spectroscopy have been  
1611 found as the most promising methods for MPs identification and quantification. WWTPs act as a

1612 significant source of MPs besides the domestic and industrial sources. MPs are found to act as a  
1613 significant vector of different contaminants such as heavy metals, additive mixtures, surfactants,  
1614 antibiotics, pesticides, and pharmaceuticals. Various adverse effects of MPs such as reduced  
1615 fecundity, increased genotoxicity, growth inhibition, the elevation of reactive oxygen species,  
1616 neurotoxic effects, reduction of photosynthesis of producers, and interferences with food intake,  
1617 enzyme activities, plasma activities, lysosome activities and energy balance, are detected in  
1618 different aquatic organisms. Different MPs treatment methods are discussed on their performance,  
1619 together with their advantages and limitations. Filtration methods are reported as the most efficient  
1620 physical treatment method although more developments are still required to implement them in  
1621 large scale MPs treatment. CWs and MBR technologies are the most efficient among the biological  
1622 treatment methods. In chemical treatment, EC, coagulation, sol-gel agglomeration, photo-Fenton  
1623 and electro-Fenton processes show promising results in MPs removal. Hybrid treatment such as  
1624 MBR-UF/RO system; coagulation followed by ozonation, GAC, DAF, RS, filtration; and CWs  
1625 based hybrid technologies have shown highly promising results for effective MPs removal.

1626

#### 1627 **Conflict of interest**

1628 There are no conflicts to declare.

1629

#### 1630 **References**

1631 Abou-Zeid, D.M., Müller, R.J., Deckwer, W.D., 2001. Degradation of natural and synthetic  
1632 polyesters under anaerobic conditions. *J. Biotechnol.* 86, 113-126.

1633 Agasild, H., Nöges, T., 2005. Cladoceran and rotifer grazing on bacteria and phytoplankton in two  
1634 shallow eutrophic lakes: in situ measurement with fluorescent microspheres. *J. Plankton Res.*  
1635 27, 1155-1174.

1636 Ahmed, I., Nipattummakul, N., Gupta, A., 2011. Characteristics of syngas from co-gasification of  
1637 polyethylene and woodchips. *Appl. Energy.* 88, 165-174.

1638 Ahmed, M.B., Zhou, J.L., Ngo, H.H., Guo, W., 2015. Adsorptive removal of antibiotics from water  
1639 and wastewater: progress and challenges. *Sci. Total Environ.* 532, 112-126.

1640 Ahmed, M.B., Zhou J.L., Ngo, H.H., Guo, W., Chen M., 2016. Progress in the preparation and  
1641 application of modified biochar for improved contaminant removal from water and wastewater.  
1642 *Bioresour. Technol.* 214, 836-851.

1643 Ahmed, M.B., Zhou J.L., Ngo, H.H., Guo, W., Thomaidis, N.S., Xu, J., 2017. Progress in the  
1644 biological and chemical treatment technologies for emerging contaminant removal from  
1645 wastewater: a critical review. *J. Hazard. Mater.* 323, 274-298.

1646 Ahmed, T., Shahid, M., Azeem, F., Rasul, I., Shah, A.A., Noman, M., Hammed A., Manzoor N.,  
1647 Manzoor I., Muhammad S., 2018. Biodegradation of plastics: current scenario and future  
1648 prospects for environmental safety. *Environ. Sci. Pollut. Res.* 25, 7287-7298.

1649 Akarsu, C., Deniz, F., 2020. Electrocoagulation/Electroflotation Process for Removal of Organics  
1650 and Microplastics in Laundry Wastewater. *CLEAN–Soil, Air, Water*, 2000146.

1651 Akbal, F, Camcı, S., 2011. Copper, chromium and nickel removal from metal plating wastewater  
1652 by electrocoagulation. *Desalination.* 269, 214-222.

1653 Ali, S.S., Qazi, I.A., Arshad, M., Khan, Z., Voice, T.C., Mehmood, C.T., 2016. Photocatalytic  
1654 degradation of low density polyethylene (LDPE) films using titania nanotubes. *Environ.*  
1655 *Nanotechnol. Monit. Manag.* 5, 44-53.

1656 Alimi, O.S., Fadare, O.O., Okoffo, E.D., 2021. Microplastics in African ecosystems: Current  
1657 knowledge, abundance, associated contaminants, techniques, and research needs.  
1658 *Sci. Total Environ.* 755, 142422.

1659 Alimi, O.S., Farner Budarz, J., Hernandez, L.M., Tufenkji, N., 2018. Microplastics and  
1660 nanoplastics in aquatic environments: aggregation, deposition, and enhanced contaminant  
1661 transport. *Environ. Sci. Technol.* 52, 1704-1724.

1662 Alomar, C., Estarellas, F., Deudero, S., 2016. Microplastics in the Mediterranean Sea: deposition  
1663 in coastal shallow sediments, spatial variation and preferential grain size. *Mar. Environ. Res.*  
1664 115, 1-10.

1665 Alvim, C. B., Mendoza-Roca, J. A., Bes-Piá, A., 2020. Wastewater treatment plant as microplastics  
1666 release source—Quantification and identification techniques. *J. Environ. Manage.* 255, 109739.

1667 Anderson, P.J., Warrack, S., Langen, V., Challis, J.K., Hanson, M.L., Rennie, MD., 2017.  
1668 Microplastic contamination in lake Winnipeg, Canada. *Environ. Pollut.* 225, 223-231.

1669 Andrady, A.L., 2011. Microplastics in the marine environment. *Mar. Pollut. Bull.* 62, 1596-1605.

1670 Anjana, K., Hinduja, M., Sujitha, K., Dharani, G., 2020. Review on plastic wastes in marine  
1671 environment—Biodegradation and biotechnological solutions. *Mar. Pollut. Bull.* 150, 110733.



1672 Ariza-Tarazona, M.C., Villarreal-Chiu, J.F., Barbieri, V., Siligardi, C., Cedillo-González, E.I., 2019.  
1673 New strategy for microplastic degradation: Green photocatalysis using a protein-based porous  
1674 N-TiO<sub>2</sub> semiconductor. *Ceram. Int.* 45, 9618-9624.

1675 Arossa, S., Martin, C., Rossbach, S., Duarte, C.M., 2019. Microplastic removal by red sea giant  
1676 clam (*Tridacna maxima*). *Environ. Pollut.* 252, 1257-1266.

1677 Ashton, K., Holmes, L., Turner, A., 2010. Association of metals with plastic production pellets in  
1678 the marine environment. *Mar. Pollut. Bullet.* 60, 2050-2055.

1679 Auta, H., Emenike, C., Fauziah, S., 2017. Distribution and importance of microplastics in the  
1680 marine environment: a review of the sources, fate, effects, and potential solutions. *Environ.*  
1681 *Int.* 102, 165-176.

1682 Avérous, L., Pollet, E., 2012. Biodegradable polymers. *Environmental silicate nano-biocomposites.*  
1683 Springer, pp 13-39.

1684 Bakir, A., O'Connor, I.A., Rowland, S.J., Hendriks, A.J., Thompson, R.C., 2016. Relative  
1685 importance of microplastics as a pathway for the transfer of hydrophobic organic chemicals to  
1686 marine life. *Environ. Pollut.* 219, 56-65.

1687 Bakir, A., Rowland, S.J., Thompson, R.C., 2014. Enhanced desorption of persistent organic  
1688 contaminants from microplastics under simulated physiological conditions. *Environ. Pollut.*  
1689 185, 16-23.

1690 Basri, H., Ismail, A.F., Aziz, M., 2011. Polyethersulfone (PES)–silver composite UF membrane:  
1691 effect of silver loading and PVP molecular weight on membrane morphology and antibacterial  
1692 activity. *Desalination.* 273, 72-80.

1693 Batel, A., Linti, F., Scherer, M., Erdinger, L., Braunbeck, T., 2016. Transfer of benzo[*a*]pyrene  
1694 from microplastics to *Artemia nauplii* and further to zebrafish via a trophic food web  
1695 experiment: CYP1A induction and visual tracking of persistent organic contaminants. *Environ.*  
1696 *Toxicol. Chem.* 35, 1656-1666.

1697 Bayo, J., Olmos, S., López-Castellanos, J., 2020. Microplastics in an urban wastewater treatment  
1698 plant: The influence of physicochemical parameters and environmental factors. *Chemosphere.*  
1699 238, 124593.

1700 Benson, N. U., Fred-Ahmadu, O. H., 2020. Occurrence and distribution of microplastics-sorbed  
1701 phthalic acid esters (PAEs) in coastal psammitic sediments of tropical Atlantic Ocean, Gulf of  
1702 Guinea. *Sci. Total Environ.* 139013.

- 1703 Besseling, E., Wang, B., Lürling, M., Koelmans, A.A., 2014. Nanoplastic affects growth of *S.*  
1704 *obliquus* and reproduction of *D. magna*. *Environ. Sci. Technol.* 48, 12336-12343.
- 1705 Besseling, E., Wegner, A., Foekema, E.M., Van Den Heuvel-Greve, M.J., Koelmans, A.A., 2013.  
1706 Effects of microplastic on fitness and PCB bioaccumulation by the lugworm *Arenicola marina*  
1707 (L.). *Environ. Sci. Technol.* 47, 593-600.
- 1708 Blair, R.M., Waldron, S., Gauchotte-Lindsay, C., 2019. Average daily flow of microplastics  
1709 through a tertiary wastewater treatment plant over a ten-month period. *Water Res.* 163, 114909.
- 1710 Bläsing, M., Amelung, W., 2018. Plastics in soil: Anal. Methods and possible sources. *Sci. Total*  
1711 *Environ.* 612, 422-435.
- 1712 Brandon, J., Goldstein, M., Ohman, M.D., 2016. Long-term aging and degradation of microplastic  
1713 particles: Comparing in situ oceanic and experimental weathering patterns. *Mar. Pollut. Bull.*  
1714 110, 299-308.
- 1715 Braun, D., 2004. Poly (vinyl chloride) on the way from the 19th century to the 21st century. *J.*  
1716 *Polym. Sci. A Polym. Chem.* 42, 578-586.
- 1717 Browne, M.A., Niven, S.J., Galloway, T.S., Rowland, S.J., Thompson, R.C., 2013. Microplastic  
1718 moves contaminants and additives to worms, reducing functions linked to health and  
1719 biodiversity. *Curr. Biol.* 23, 2388-2392.
- 1720 Burra, K., Gupta, A., 2018a. Kinetics of synergistic effects in co-pyrolysis of biomass with plastic  
1721 wastes. *Appl. Energy.* 220, 408-418.
- 1722 Burra, K., Gupta, A., 2018b. Synergistic effects in steam gasification of combined biomass and  
1723 plastic waste mixtures. *Appl. Energy.* 211, 230-236.
- 1724 Cai, L., Wang, J., Peng, J., Wu, Z., Tan X., 2018. Observation of the degradation of three types of  
1725 plastic pellets exposed to UV irradiation in three different environments. *Sci. Total Environ.*  
1726 628, 740-747.
- 1727 Carlin, J., Craig, C., Little, S., Donnelly, M., Fox, D., Zhai, L., Walters, L., 2020. Microplastic  
1728 accumulation in the gastrointestinal tracts in birds of prey in central Florida, USA. *Environ.*  
1729 *Pollut.* 114633.
- 1730 Carr, S.A., Liu, J., Tesoro, A.G., 2016. Transport and fate of microplastic particles in wastewater  
1731 treatment plants. *Water Res.* 91, 174-182.
- 1732 Carson, H.S., Colbert, S.L., Kaylor, M.J., McDermid, K.J., 2011. Small plastic debris changes  
1733 water movement and heat transfer through beach sediments. *Mar. Pollut. Bull.* 62, 1708-1713.

1734 Castañeda, R.A., Avlijas, S., Simard, M.A., Ricciardi, A., 2014. Microplastic pollution in St.  
1735 Lawrence river sediments. *Can. J. Fish. Aquat.* 71, 1767-1771.

1736 Chamas, A., Moon, H., Zheng, J., Qiu, Y., Tabassum, T., Jang, M., J. H., Abu-Omar, Scott, S. L.,  
1737 Suh, S., 2020. Degradation Rates of Plastics in the Environment. *ACS Sustain. Chem. Eng.* 8,  
1738 3494-3511.

1739 Chang, M., 2015. Reducing microplastics from facial exfoliating cleansers in wastewater through  
1740 treatment versus consumer product decisions. *Mar. Pollut. Bull.* 101, 330-333.

1741 Chen, R., Qi, M., Zhang, G., Yi, C., 2018. Comparative experiments on polymer degradation  
1742 technique of produced water of polymer flooding oilfield. *IOP Conference Series: Earth and  
1743 Environmental Science.* 113, pp. 012208.

1744 Chen, G., Feng, Q., Wang, J., 2020. Mini-review of microplastics in the atmosphere and their risks  
1745 to humans. *Sci. Total Environ.* 703, 135504.

1746 Chua, E.M., Shimeta, J., Nugegoda, D., Morrison, P.D., Clarke, B.O., 2014. Assimilation of  
1747 polybrominated diphenyl ethers from microplastics by the marine amphipod, *Allorchestes  
1748 compressa.* *Environ. Sci. Technol.* 48, 8127-8134.

1749 Claessens, M., Van Cauwenberghe, L., Vandegheuchte, M.B., Janssen, C.R., 2013. New techniques  
1750 for the detection of microplastics in sediments and field collected organisms. *Mar. Pollut. Bull.*  
1751 70, 227-233.

1752 Cole, M., Lindeque, P., Fileman, E., Halsband, C., Galloway, T.S., 2015. The impact of polystyrene  
1753 microplastics on feeding, function and fecundity in the marine copepod *Calanus helgolandicus.*  
1754 *Environ. Sci. Technol.* 49, 1130-1137.

1755 Cole, M., Lindeque, P., Fileman, E., Halsband, C., Goodhead, R., Moger, J., Galloway T.S., 2013.  
1756 Microplastic ingestion by zooplankton. *Environ. Sci. Technol.* 47, 6646-6655.

1757 Cole, M., Lindeque, P., Halsband, C., Galloway, T.S., 2011. Microplastics as contaminants in the  
1758 marine environment: a review. *Mar. Pollut. Bull.* 62, 2588-2597.

1759 Cole, M., Lindeque, P.K., Fileman, E., Clark, J., Lewis, C., Halsband, C., Galloway T.S., 2016.  
1760 Microplastics alter the properties and sinking rates of zooplankton faecal pellets. *Environ. Sci.*  
1761 *Technol.* 50, 3239-3246.

1762 Cooper, D.A., Corcoran, P.L., 2010. Effects of mechanical and chemical processes on the  
1763 degradation of plastic beach debris on the island of Kauai, Hawaii. *Mar. Pollut. Bull.* 60, 650-  
1764 654.

1765 Corcoran, P.L., Biesinger, M.C., Grifi, M., 2009. Plastics and beaches: a degrading relationship.  
1766 Mar. Pollut. Bull. 58, 80-84.

1767 Courtene-Jones, W., Quinn, B., Murphy, F., Gary, S.F., Narayanaswamy, B.E., 2017. Optimisation  
1768 of enzymatic digestion and validation of specimen preservation methods for the analysis of  
1769 ingested microplastics. Anal. Methods. 9, 1437-1445.

1770 Crawford, C., Quinn, B., 2017. Microplastic separation techniques. Microplastic Contaminants.  
1771 Elsevier, pp. 203-218.

1772 da Costa, J.P., Santos, P.S., Duarte, A.C., Rocha-Santos, T., 2016. (Nano) plastics in the  
1773 environment—sources, fates and effects. Sci. Total Environ. 566, 15-26.

1774 Dazzi, A., Prater, C.B., Hu, Q., Chase, D.B., Rabolt, J.F., Marcott, C., 2012. AFM–IR: combining  
1775 atomic force microscopy and infrared spectroscopy for nanoscale chemical characterization.  
1776 Appl. Spectrosc. 66, 1365-1384.

1777 Dazzi, A., Saunier, J., Kjoller, K., Yagoubi, N., 2015. Resonance enhanced AFM-IR: A new  
1778 powerful way to characterize blooming on polymers used in medical devices. Int. J. Pharm.  
1779 484, 109-114.

1780 De la Cruz, N., Giménez, J., Esplugas, S., Grandjean, D., De Alencastro, L., Pulgarin, C., 2012.  
1781 Degradation of 32 emergent contaminants by UV and neutral photo-fenton in domestic  
1782 wastewater effluent previously treated by activated sludge. Water Res. 46, 1947-1957.

1783 de Luna, M.D.G., Veciana, M.L., Su, C.C., Lu, M.C., 2012. Acetaminophen degradation by electro-  
1784 Fenton and photoelectro-Fenton using a double cathode electrochemical cell. J. Hazard. Mater.  
1785 217, 200-207.

1786 Dehaut, A., Cassone, A-L., Frère, L., Hermabessiere, L., Himber, C., Rinnert, E., Riviere G.,  
1787 Lambert C., Soudant P., Huvet A., Duflos G., 2016. Microplastics in seafood: Benchmark  
1788 protocol for their extraction and characterization. Environ. Pollut. 215, 223-233.

1789 Dekiff, J.H., Remy, D., Klasmeier, J., Fries, E., 2014. Occurrence and spatial distribution of  
1790 microplastics in sediments from Norderney. Environ. Pollut. 186, 248-256.

1791 Derraik, J.G., 2002. The pollution of the marine environment by plastic debris: a review. Mar.  
1792 Pollut. Bull. 44, 842-852.

1793 Di, M., Wang, J., 2018. Microplastics in surface waters and sediments of the Three Gorges  
1794 Reservoir, China. Sci. Total Environ. 616, 1620-1627.

1795 Dolar, D., Gros, M., Rodriguez-Mozaz, S., Moreno, J., Comas, J., Rodriguez-Roda, I., Barcelo D.,  
1796 2012. Removal of emerging contaminants from municipal wastewater with an integrated  
1797 membrane system, MBR–RO. *J. Hazard. Mater.* 239, 64-69.

1798 Dong, H., Chen, Y., Wang, J., Zhang, Y., Zhang, P., Li, X., Zou, J., Zhou, A., 2020. Interactions of  
1799 microplastics and antibiotic resistance genes and their effects on the aquaculture environments.  
1800 *J. Hazard. Mater.* 123961.

1801 Doyle, M.J., Watson, W., Bowlin, N.M., Sheavly, S.B., 2011. Plastic particles in coastal pelagic  
1802 ecosystems of the Northeast Pacific ocean. *Mar. Environ. Res.* 71, 41-52.

1803 Dřimal, P., Hoffmann, J., Družbík, M., 2007. Evaluating the aerobic biodegradability of plastics in  
1804 soil environments through GC and IR analysis of gaseous phase. *Polym. Test.* 26, 729-741.

1805 Dris, R., Gasperi, J., Saad, M., Mirande, C., Tassin, B., 2016. Synthetic fibers in atmospheric  
1806 fallout: a source of microplastics in the environment? *Mar. Pollut. Bull.* 104, 290-293.

1807 Duis, K., Coors, A., 2016. Microplastics in the aquatic and terrestrial environment: sources (with  
1808 a specific focus on personal care products), fate and effects. *Environ. Sci. Eur.* 28, 2.

1809 Dümichen, E., Barthel, A-K., Braun, U., Bannick, C.G., Brand, K., Jekel, M., Senz R., 2015.  
1810 Analysis of polyethylene microplastics in environmental samples, using a thermal  
1811 decomposition method. *Water Res.* 85, 451-457.

1812 Dümichen, E., Eisentraut, P., Bannick, C.G., Barthel, A.K., Senz, R., Braun, U., 2017. Fast  
1813 identification of microplastics in complex environmental samples by a thermal degradation  
1814 method. *Chemosphere.* 174, 572-584.

1815 Eerkes-Medrano, D., Thompson, R.C., Aldridge, D.C., 2015. Microplastics in freshwater systems:  
1816 a review of the emerging threats, identification of knowledge gaps and prioritisation of  
1817 research needs. *Water Res.* 75, 63-82.

1818 Enfrin, M., Dumée, L.F., Lee, J., 2019. Nano/microplastics in water and wastewater treatment  
1819 processes – Origin, impact and potential solutions. *Water Res.* 161, 621-638.

1820 Ersahin, M.E., Ozgun, H., Dereli, R.K., Ozturk, I., Roest, K., van Lier, J.B., 2012. A review on  
1821 dynamic membrane filtration: materials, applications and future perspectives. *Bioresour.*  
1822 *Technol.* 122, 196-206.

1823 Eskandarloo, H., Kierulf, A., Abbaspourrad, A., 2017. Light-harvesting synthetic nano-and  
1824 micromotors: a review. *Nanoscale* 9, 12218-12230.

1825 Evangeliou, N., Grythe, H., Klimont, Z., Heyes, C., Eckhardt, S., Lopez-Aparicio, S., Stohl, A.,  
1826 2020. Atmospheric transport is a major pathway of microplastics to remote regions. *Nat.*  
1827 *Commun.* 11, 1-11.

1828 Farias, E.L., Howe, K.J., Thomson, B.M., 2014. Effect of membrane bioreactor solids retention  
1829 time on reverse osmosis membrane fouling for wastewater reuse. *Water Res.* 49, 53-61.

1830 Felsing, S., Kochleus, C., Buchinger, S., Brennholt, N., Stock, F., Reifferscheid, G., 2018. A new  
1831 approach in separating microplastics from environmental samples based on their electrostatic  
1832 behavior. *Environ. Pollut.* 234, 20-28.

1833 Feng, H-M., Zheng, J-C., Lei, N-Y., Yu, L., Kong, K.H-K., Yu, H-Q., et al., 2011. Photoassisted  
1834 Fenton degradation of polystyrene. *Environ. Sci. Technol.* 45, 744-750.

1835 Free, C.M., Jensen, O.P., Mason, S.A., Eriksen, M., Williamson, N.J., Boldgiv, B., 2014. High-  
1836 levels of microplastic pollution in a large, remote, mountain lake. *Mar. Pollut. Bull.* 85, 156-  
1837 163.

1838 Frias, J., Sobral, P., Ferreira, A., 2010. Organic contaminants in microplastics from two beaches of  
1839 the Portuguese coast. *Mar. Pollut. Bull.* 60, 1988-1992.

1840 Fries, E., Dekiff, J.H., Willmeyer, J., Nuelle, M.T., Ebert, M., Remy, D., 2013. Identification of  
1841 polymer types and additives in marine microplastic particles using pyrolysis-GC/MS and  
1842 scanning electron microscopy. *Environ. Sci. Process Impacts.* 15, 1949-1956.

1843 Fu, J., Lee, W-N., Coleman, C., Nowack, K., Carter, J., Huang, C.H., 2019. Removal of  
1844 pharmaceuticals and personal care products by two-stage biofiltration for drinking water  
1845 treatment. *Sci. Total Environ.* 664, 240-248.

1846 Fu, Z., Chen, G., Wang, W., Wang, J., 2020. Microplastic pollution research methodologies,  
1847 abundance, characteristics and risk assessments for aquatic biota in China. *Environ. Pollut.*  
1848 115098.

1849 Ganiyu, S.O., Zhou, M., Martínez-Huitle, C.A., 2018. Heterogeneous electro-Fenton and  
1850 photoelectro-Fenton processes: a critical review of fundamental principles and application for  
1851 water/wastewater treatment. *Appl. Catal. B.* 235, 103-129.

1852 Ganzenko, O., Huguenot, D., Van Hullebusch, E.D., Esposito, G., Oturan, M.A., 2014.  
1853 Electrochemical advanced oxidation and biological processes for wastewater treatment: a  
1854 review of the combined approaches. *Environ. Sci. Pollut. Res.* 21, 8493-8524.

1855 Garcia-Segura, S., Eiband, M.M.S, de Melo, J.V., Martínez-Huitle, C.A., 2017. Electrocoagulation  
1856 and advanced electrocoagulation processes: A general review about the fundamentals,  
1857 emerging applications and its association with other technologies. *J. Electroanal. Chem.* 801,  
1858 267-299.

1859 Gasperi, J., Wright, S.L., Dris, R., Collard, F., Mandin, C., Guerrouache, M., Langlois V., Kelly  
1860 F.J., Tassin B., 2018. Microplastics in air: are we breathing it in? *Curr. Opin. Environ. Sci.*  
1861 *Health.* 1, 1-5.

1862 Godoy, V., Blázquez, G., Calero, M., Quesada, L., Martín-Lara, M., 2019. The potential of  
1863 microplastics as carriers of metals. *Environ. Pollut.* 255, 113363.

1864 Gómez, E.F., Michel Jr., F.C., 2013. Biodegradability of conventional and bio-based plastics and  
1865 natural fiber composites during composting, anaerobic digestion and long-term soil incubation.  
1866 *Polym. Degrad. Stab.* 98, 2583-2591.

1867 Gouin, T., Roche, N., Lohmann, R., Hodges, G., 2011. A thermodynamic approach for assessing  
1868 the environmental exposure of chemicals absorbed to microplastic. *Environ. Sci. Technol.* 45,  
1869 1466-1472.

1870 Grbic, J., Nguyen, B., Guo, E., You, J.B., Sinton, D., Rochman, C.M., 2019. Magnetic extraction  
1871 of microplastics from environmental samples. *Environ. Sci. Technol. Letters* 6, 68-72.

1872 Green, D.S., Boots, B., O'Connor, N.E., Thompson, R., 2017. Microplastics affect the ecological  
1873 functioning of an important biogenic habitat. *Environ. Sci. Technol.* 51, 68-77.

1874 Gu, J.G., Gu, J.D., 2005. Methods currently used in testing microbiological degradation and  
1875 deterioration of a wide range of polymeric materials with various degree of degradability: a  
1876 review. *J. Polym. Environ.* 13, 65-74.

1877 Guo, X., Pang, J., Chen, S., Jia, H., 2018. Sorption properties of tylosin on four different  
1878 microplastics. *Chemosphere.* 209, 240-245.

1879 Guo, X, Wang, J., 2019. Sorption of antibiotics onto aged microplastics in freshwater and seawater.  
1880 *Mar. Pollut. Bull.* 149, 110511.

1881 Hale, R. C., Seeley, M. E., La Guardia, M. J., Mai, L., Zeng, E. Y., 2020. A global perspective on  
1882 microplastics. *J. Geophys. Res. Oceans* 125, e2018JC014719.

1883 Harrison, J.P, Ojeda, J.J., Romero-González, M.E., 2012. The applicability of reflectance micro-  
1884 Fourier-transform infrared spectroscopy for the detection of synthetic microplastics in marine  
1885 sediments. *Sci. Total Environ.* 416, 455-463.

1886 Helcoski, R., Yonkos, L.T., Sanchez, A., Baldwin, A.H., 2020. Wetland soil microplastics are  
1887 negatively related to vegetation cover and stem density. *Environ. Pollut.* 256, 113391.

1888 Herbort, A.F., Schuhen K., 2017. A concept for the removal of microplastics from the marine  
1889 environment with innovative host-guest relationships. *Environ. Sci. Pollut. Res.* 24, 11061-  
1890 11065.

1891 Herbort, A.F., Sturm, M.T., Fiedler, S., Abkai, G., Schuhen, K., 2018. Alkoxy-silyl induced  
1892 agglomeration: a new approach for the sustainable removal of microplastic from aquatic  
1893 systems. *J. Polym. Environ.* 26, 4258-4270.

1894 Hernandez, L.M., Xu, E.G., Larsson, H.C., Tahara, R., Maisuria, V.B., Tufenkji, N., 2019. Plastic  
1895 teabags release billions of microparticles and nanoparticles into tea. *Environ. Sci. Technol.* 53,  
1896 12300-12310.

1897 Hernandez, L.M., Yousefi, N., Tufenkji, N., 2017. Are there nanoplastics in your personal care  
1898 products? *Environ. Sci. Technol. Letters* 4, 280-285.

1899 Hidalgo-Ruz, V., Gutow, L., Thompson, R.C., Thiel, M., 2012. Microplastics in the marine  
1900 environment: a review of the methods used for identification and quantification. *Environ. Sci.*  
1901 *Technol.* 46, 3060-3075.

1902 Hidayaturrahman, H., Lee, T.G., 2019. A study on characteristics of microplastic in wastewater of  
1903 South Korea: Identification, quantification, and fate of microplastics during treatment process.  
1904 *Mar. Pollut. Bull.* 146, 696-702.

1905 Hirai, H., Takada, H., Ogata, Y., Yamashita, R., Mizukawa, K., Saha, M., Kwan C, Moore C, Gray  
1906 H, Laursen D, Zettler E.R., 2011. Organic microcontaminants in marine plastics debris from  
1907 the open ocean and remote and urban beaches. *Mar. Pollut. Bull.* 62, 1683-1692.

1908 Holmes, L.A., Turner, A., Thompson, R.C., 2012. Adsorption of trace metals to plastic resin pellets  
1909 in the marine environment. *Environ. Pollut.* 160, 42-48.

1910 Horton, A.A., Walton, A., Spurgeon, D.J., Lahive, E., Svendsen, C., 2017. Microplastics in  
1911 freshwater and terrestrial environments: evaluating the current understanding to identify the  
1912 knowledge gaps and future research priorities. *Sci. Total Environ.* 586, 127-141.

1913 Hu, L., Fu, J., Wang, S., Xiang, Y., Pan, X., 2020. Microplastics generated under simulated fire  
1914 scenarios: Characteristics, antimony leaching, and toxicity. *Environ. Pollut.* 269, 115905.

1915 Hu, Y., Wang, X.C., Ngo, H.H., Sun, Q., Yang, Y., 2018. Anaerobic dynamic membrane bioreactor  
1916 (AnDMBR) for wastewater treatment: A review. *Bioresour. Technol.* 247, 1107-1118.



1917 Huang, B.C., Guan, Y.F., Chen, W., Yu, H.Q., 2017. Membrane fouling characteristics and  
1918 mitigation in a coagulation-assisted microfiltration process for municipal wastewater  
1919 pretreatment. *Water Res.* 123, 216-223.

1920 Hüffer, T., Hofmann, T., 2016. Sorption of non-polar organic compounds by micro-sized plastic  
1921 particles in aqueous solution. *Environ. Pollut.* 214, 194-201.

1922 Huppertsberg, S., Knepper, T.P., 2018. Instrumental analysis of microplastics—benefits and  
1923 challenges. *Anal. Bioanal. Chem.* 410, 6343-6352.

1924 Ibrar, I., Altaee, A., Zhou, J.L., Naji, O., Khanafer, D., 2020. Challenges and potentials of forward  
1925 osmosis process in the treatment of wastewater. *Crit. Rev. Environ. Sci. Technol.* 50, 1339-  
1926 1383.

1927 Imhof, H.K., Schmid, J., Niessner, R., Ivleva, N.P., Laforsch, C., 2012. A novel, highly efficient  
1928 method for the separation and quantification of plastic particles in sediments of aquatic  
1929 environments. *Limnol. Oceanogr.: Methods.* 10, 524-537.

1930 Ivleva, N.P., Wiesheu, A.C., Niessner, R., 2017. Microplastic in aquatic ecosystems. *Angew. Chem.*  
1931 *Int. Ed.* 56, 1720-1739.

1932 Jang, M., Shim, W. J., Han, G. M., Cho, Y., Moon, Y., Hong, S. H., 2020. Relative importance of  
1933 aqueous leachate versus particle ingestion as uptake routes for microplastic additives  
1934 (hexabromocyclododecane) to mussels. *Environ. Pollut.* 116272.

1935 Jiang, L., Hu, X., Yin, D., Zhang, H., Yu, Z., 2011. Occurrence, distribution and seasonal variation  
1936 of antibiotics in the Huangpu River, Shanghai, China. *Chemosphere.* 82, 822-828.

1937 Jin, Q., Wang, X., Li, S., Mikulčić, H., Bešenić, T., Deng, S., Vujanović M, Tan H, Kumfer B.M.,  
1938 2019. Synergistic effects during co-pyrolysis of biomass and plastic: Gas, tar, soot, char  
1939 products and thermogravimetric study. *J. Energy Inst.* 92, 108-117.

1940 Joo, S., Cho, I.J., Seo, H., Son, H.F., Sagong, H.Y., Shin, T.J., Choi S.Y., Lee S.Y., Kim K.J., 2018.  
1941 Structural insight into molecular mechanism of poly (ethylene terephthalate) degradation. *Nat.*  
1942 *Commun.* 9, 1-12.

1943 Kalčíková, G., Alič, B., Skalar, T., Bundschuh, M., Gotvajn, A.Ž., 2017. Wastewater treatment  
1944 plant effluents as source of cosmetic polyethylene microbeads to freshwater. *Chemosphere.*  
1945 188, 25-31.

1946 Kalčíková, G., Skalar, T., Marolt, G., Kokalj, A. J., 2020. An environmental concentration of aged  
1947 microplastics with adsorbed silver significantly affects aquatic organisms. *Water Res.* 115644.

- 1948 Kale, G., Kijchavengkul, T., Auras, R., Rubino, M., Selke, S.E., Singh, S.P., 2007. Compostability  
1949 of bioplastic packaging materials: an overview. *Macromol. Biosci.* 7, 255-277.
- 1950 Kang, J.H., Kwon, O.Y., Shim, W.J., 2015. Potential threat of microplastics to zooplanktivores in  
1951 the surface waters of the Southern Sea of Korea. *Arch. Environ. Contam. Toxicol.* 69, 340-  
1952 351.
- 1953 Kaposi, K.L., Mos, B., Kelaher, B.P., Dworjanyn, S.A., 2014. Ingestion of microplastic has limited  
1954 impact on a marine larva. *Environ. Sci. Technol.* 48, 1638-1645.
- 1955 K ppler, A., Fischer, D., Oberbeckmann, S., Schernewski, G., Labrenz, M., Eichhorn, K.J., Voit  
1956 B., 2016. Analysis of environmental microplastics by vibrational microspectroscopy: FTIR,  
1957 Raman or both? *Anal. Bioanal. Chem.* 408, 8377-8391.
- 1958 Karami, A., Golieskardi, A., Choo, C.K., Romano, N., Ho, Y.B., Salamatina, B., 2017. A high-  
1959 performance protocol for extraction of microplastics in fish. *Sci. Total Environ.* 578, 485-494.
- 1960 Karlsson, T.M., Hassell v, M., Jakubowicz, I., 2018. Influence of thermooxidative degradation on  
1961 the in situ fate of polyethylene in temperate coastal waters. *Mar. Pollut. Bull.* 135, 187-194.
- 1962 Karlsson, T.M., Vethaak, A.D., Almroth, B.C., Ariese, F., van Velzen, M., Hassell v, M., Leslie  
1963 H.A., 2017. Screening for microplastics in sediment, water, marine invertebrates and fish:  
1964 method development and microplastic accumulation. *Mar. Pollut. Bull.* 122, 403-408.
- 1965 Kijchavengkul, T., Auras, R., Rubino, M., Selke, S., Ngouajio, M., Fernandez, R.T., 2010.  
1966 Biodegradation and hydrolysis rate of aliphatic aromatic polyester. *Polym. Degrad. Stab.* 95,  
1967 2641-2647.
- 1968 Klavarioti, M., Mantzavinos, D., Kassinos, D., 2009. Removal of residual pharmaceuticals from  
1969 aqueous systems by advanced oxidation processes. *Environ. Int.* 35, 402-417.
- 1970 Klein, S., Dimzon, I.K., Eubeler, J., Knepper, T.P., 2018. Analysis, occurrence, and degradation of  
1971 microplastics in the aqueous environment. *Freshwater Microplastics*. Springer, Cham, pp. 51-  
1972 67.
- 1973 Klein, S., Worch, E., Knepper, T.P., 2015. Occurrence and spatial distribution of microplastics in  
1974 river shore sediments of the Rhine-Main area in Germany. *Environ. Sci. Technol.* 49, 6070-  
1975 6076.
- 1976 Koelmans, A.A., Bakir, A., Burton, G.A., Janssen, C.R., 2016. Microplastic as a vector for  
1977 chemicals in the aquatic environment: critical review and model-supported reinterpretation of  
1978 empirical studies. *Environ. Sci. Technol.* 50, 3315-3326.

- 1979 Koelmans, A.A., Besseling, E., Wegner, A., Foekema, E.M., 2013. Plastic as a carrier of POPs to  
1980 aquatic organisms: a model analysis. *Environ. Sci. Technol.* 47, 7812-7820.
- 1981 Koelmans, A.A., Nor, N.H.M., Hermsen, E., Kooi, M., Mintenig, S.M., De France, J., 2019.  
1982 Microplastics in freshwaters and drinking water: critical review and assessment of data quality.  
1983 *Water Res.* 155, 410-422.
- 1984 Kolstad, J.J., Vink, E.T., De Wilde, B., Debeer, L., 2012. Assessment of anaerobic degradation of  
1985 Ingeo™ polylactides under accelerated landfill conditions. *Polym. Degrad. Stab.* 97, 1131-  
1986 1141.
- 1987 Kwon, O. Y., Kang, J. H., Hong, S. H., Shim, W. J., 2020. Spatial distribution of microplastic in the  
1988 surface waters along the coast of Korea. *Mar. Pollut. Bull.* 110729.
- 1989 Lambert, S., Scherer, C., Wagner, M., 2017. Ecotoxicity testing of microplastics: Considering the  
1990 heterogeneity of physicochemical properties. *Integr. Environ. Assess. Manag.* 13, 470-475.
- 1991 Lares, M., Ncibi, M.C., Sillanpää, M., Sillanpää, M., 2018. Occurrence, identification and removal  
1992 of microplastic particles and fibers in conventional activated sludge process and advanced  
1993 MBR technology. *Water Res.* 133, 236-246.
- 1994 Lee, H., Shim, W.J., Kwon, J.H., 2014. Sorption capacity of plastic debris for hydrophobic organic  
1995 chemicals. *Sci. Total Environ.* 470, 1545-1552.
- 1996 Lee, K.E., Morad, N., Teng, T.T., Poh, B.T., 2012. Development, characterization and the  
1997 application of hybrid materials in coagulation/flocculation of wastewater: A review. *Chem.*  
1998 *Eng. J.* 203, 370-386.
- 1999 Lee, Y. K., Murphy, K. R., Hur, J., 2020. Fluorescence signatures of dissolved organic matter  
2000 leached from microplastics: Polymers and additives. *Environ. Sci. Technol.* 54, 11905-11914.
- 2001 Leslie, H., Brandsma, S., Van Velzen, M., Vethaak, A., 2017. Microplastics en route: Field  
2002 measurements in the Dutch river delta and Amsterdam canals, wastewater treatment plants,  
2003 North Sea sediments and biota. *Environ. Int.* 101, 133-142.
- 2004 Levin, I.W., Bhargava, R., 2005. Fourier transform infrared vibrational spectroscopic imaging:  
2005 integrating microscopy and molecular recognition. *Annu. Rev. Phys. Chem.* 56, 429-474.
- 2006 Li, G., Ben, W., Ye, H., Zhang, D., Qiang, Z., 2018a. Performance of ozonation and biological  
2007 activated carbon in eliminating sulfonamides and sulfonamide-resistant bacteria: A pilot-scale  
2008 study. *Chem. Eng. J.* 341, 327-334.

- 2009 Li, J., Liu, H., Chen, J.P., 2018b. Microplastics in freshwater systems: A review on occurrence,  
2010 environmental effects, and methods for microplastics detection. *Water Res.* 137, 362-374.
- 2011 Li, J., Zhang, K., Zhang, H., 2018c. Adsorption of antibiotics on microplastics. *Environ. Pollut.*  
2012 237, 460-467.
- 2013 Li, L., Xu, G., Yu, H., 2018d. Dynamic membrane filtration: formation, filtration, cleaning, and  
2014 applications. *Chem. Eng. Technol.* 41, 7-18.
- 2015 Li, L., Xu, G., Yu, H., Xing, J., 2018e. Dynamic membrane for micro-particle removal in  
2016 wastewater treatment: Performance and influencing factors. *Sci. Total Environ.* 627, 332-340.
- 2017 Li, Q., Wu, J., Zhao, X., Gu, X., Ji, R., 2019a. Separation and identification of microplastics from  
2018 soil and sewage sludge. *Environ. Pollut.* 254, 113076.
- 2019 Li, Q., Xu, Z., Pinnau, I., 2007. Fouling of reverse osmosis membranes by biopolymers in  
2020 wastewater secondary effluent: Role of membrane surface properties and initial permeate flux.  
2021 *J. Membr. Sci.* 290, 173-181.
- 2022 Li, W., Shi, Y., Gao, L., Liu, J., Cai, Y., 2012. Occurrence of antibiotics in water, sediments, aquatic  
2023 plants, and animals from Baiyangdian Lake in North China. *Chemosphere.* 89, 1307-1315.
- 2024 Li, X., Mei, Q., Chen, L., Zhang, H., Dong, B., Dai, X., et al., 2019b. Enhancement in adsorption  
2025 potential of microplastics in sewage sludge for metal contaminants after the wastewater  
2026 treatment process. *Water Res.* 157, 228-237.
- 2027 Liang, W., Luo, Y., Song, S., Dong, X., Yu, X., 2013. High photocatalytic degradation activity of  
2028 polyethylene containing polyacrylamide grafted TiO<sub>2</sub>. *Polym. Degrad. Stab.* 98, 1754-1761.
- 2029 Lin, H., Zhang, M., Wang, F., Meng, F., Liao, B.Q., Hong, H., Chen J., Gao W., 2014. A critical  
2030 review of extracellular polymeric substances (EPSs) in membrane bioreactors: characteristics,  
2031 roles in membrane fouling and control strategies. *J. Membr. Sci.* 460, 110-125.
- 2032 Lithner, D., Damberg, J., Dave, G., Larsson, Å., 2009. Leachates from plastic consumer products—  
2033 screening for toxicity with *Daphnia magna*. *Chemosphere.* 74, 1195-1200.
- 2034 Lithner, D., Larsson, Å., Dave, G., 2011. Environmental and health hazard ranking and assessment  
2035 of plastic polymers based on chemical composition. *Sci. Total Environ.* 409, 3309-3324.
- 2036 Liu, F.F., Liu, G.Z., Zhu, Z.L., Wang, S.C., Zhao, F.F., 2019a. Interactions between microplastics  
2037 and phthalate esters as affected by microplastics characteristics and solution chemistry.  
2038 *Chemosphere.* 214, 688-694.

2039 Liu, F., Vianello, A., Vollertsen, J., 2019b. Retention of microplastics in sediments of urban and  
2040 highway stormwater retention ponds. *Environ. Pollut.* 255, 113335.

2041 Liu, X., Yuan, W., Di, M., Li, Z., Wang, J., 2019c. Transfer and fate of microplastics during the  
2042 conventional activated sludge process in one wastewater treatment plant of China. *Chem. Eng.*  
2043 *J.* 362, 176-182.

2044 Löder, M.G., Gerdt, G., 2015. Methodology used for the detection and identification of  
2045 microplastics—A critical appraisal. *Marine Anthropogenic Litter*. Springer, pp. 201-227.

2046 Löder, M.G.J., Kuczera, M., Mintenig, S., Lorenz, C., Gerdt, G., 2015. Focal plane array detector-  
2047 based micro-Fourier-transform infrared imaging for the analysis of microplastics in  
2048 environmental samples. *Environ. Chem.* 12, 563-581.

2049 Lönnstedt, O.M., Eklöv, P., 2016. Environmentally relevant concentrations of microplastic  
2050 particles influence larval fish ecology. *Science.* 352, 1213-1216.

2051 Lucas, N., Bienaime, C., Belloy, C., Queneudec, M., Silvestre, F., Nava-Saucedo, J-E., 2008.  
2052 Polymer biodegradation: Mechanisms and estimation techniques—A review. *Chemosphere.* 73,  
2053 429-442.

2054 Luís, L.G., Ferreira, P., Fonte, E., Oliveira, M., Guilhermino, L., 2015. Does the presence of  
2055 microplastics influence the acute toxicity of chromium (VI) to early juveniles of the common  
2056 goby (*Pomatoschistus microps*)? A study with juveniles from two wild estuarine populations.  
2057 *Aquat. Toxicol.* 164, 163-174.

2058 Luo, H., Zhao, Y., Li, Y., Xiang, Y., He, D., Pan, X., 2020. Aging of microplastics affects their  
2059 surface properties, thermal decomposition, additives leaching and interactions in simulated  
2060 fluids. *Sci. Total Environ.* 714, 136862.

2061 Luo, J., Chen, Y., Feng, L., 2016. Polycyclic aromatic hydrocarbon affects acetic acid production  
2062 during anaerobic fermentation of waste activated sludge by altering activity and viability of  
2063 acetogen. *Environ. Sci. Technol.* 50, 6921-6929.

2064 Lv, X., Dong, Q., Zuo, Z., Liu, Y., Huang, X., Wu, W-M., 2019. Microplastics in a municipal  
2065 wastewater treatment plant: Fate, dynamic distribution, removal efficiencies, and control  
2066 strategies. *J. Clean. Prod.* 225, 579-586.

2067 Ma, B., Xue, W., Ding, Y., Hu, C., Liu, H., Qu, J., 2019. Removal characteristics of microplastics  
2068 by Fe-based coagulants during drinking water treatment. *J. Environ. Sci.* 78, 267-275.

2069 Macwan, D., Dave, P.N., Chaturvedi, S., 2011. A review on nano-TiO<sub>2</sub> sol–gel type syntheses and  
2070 its applications. *J. Mater. Sci.* 46, 3669-3686.

2071 Maes, T., Jessop, R., Wellner, N., Haupt, K., Mayes, A.G., 2017a. A rapid-screening approach to  
2072 detect and quantify microplastics based on fluorescent tagging with Nile Red. *Sci. Rep.* 7, 1-  
2073 10.

2074 Mahon, A.M., O’Connell, B., Healy, M.G., O’Connor, I., Officer, R., Nash, R., Morrison L., 2017.  
2075 Microplastics in sewage sludge: effects of treatment. *Environ. Sci. Technol.* 51, 810-818.

2076 Majewsky, M., Bitter, H., Eiche, E., Horn, H., 2016. Determination of microplastic polyethylene  
2077 (PE) and polypropylene (PP) in environmental samples using thermal analysis (TGA-DSC).  
2078 *Sci. Total Environ.* 568, 507-511.

2079 Malankowska, M., Echaide-Gorriz, C., Coronas, J., 2021. Microplastics in marine environment:  
2080 a review on sources, classification, and potential remediation by membrane technology.  
2081 *Environ. Sci.: Water Res. Technol.*

2082 Mason, S.A., Garneau, D., Sutton, R., Chu, Y., Ehmann, K., Barnes, J., Fink P., Papazissimos D.,  
2083 Rogers D.L., 2016. Microplastic pollution is widely detected in US municipal wastewater  
2084 treatment plant effluent. *Environ. Pollut.* 218, 1045-1054.

2085 Massardier-Nageotte, V., Pestre, C., Cruard-Pradet, T., Bayard, R., 2006. Aerobic and anaerobic  
2086 biodegradability of polymer films and physico-chemical characterization. *Polym. Degrad.*  
2087 *Stab.* 91, 620-627.

2088 Mathalon, A., Hill, P., 2014. Microplastic fibers in the intertidal ecosystem surrounding Halifax  
2089 Harbor, Nova Scotia. *Mar. Pollut. Bull.* 81, 69-79.

2090 McCormick, A., Hoellein, T.J., Mason, S.A., Schluep, J., Kelly, J.J., 2014. Microplastic is an  
2091 abundant and distinct microbial habitat in an urban river. *Environ. Sci. Technol.* 48, 11863-  
2092 11871.

2093 Mendoza, L.M.R., Jones P.R., 2015. Characterisation of microplastics and toxic chemicals  
2094 extracted from microplastic samples from the North Pacific Gyre. *Environ. Chem.* 12, 611-  
2095 617.

2096 Mendoza, L.M.R., Karapanagioti, H., Álvarez, N.R., 2018. Micro (nanoplastics) in the marine  
2097 environment: current knowledge and gaps. *Curr. Opin. Environ. Sci. Health.* 1, 47-51.

2098 Miao, F., Liu, Y., Gao, M., Yu, X., Xiao, P., Wang, M., et al., 2020. Degradation of polyvinyl  
2099 chloride microplastics via an electro-Fenton-like system with a TiO<sub>2</sub>/graphite cathode. *J.*  
2100 *Hazard. Mater.* 123023.

2101 Michielssen, M.R., Michielssen, E.R., Ni, J., Duhaime, M.B., 2016. Fate of microplastics and other  
2102 small anthropogenic litter (SAL) in wastewater treatment plants depends on unit processes  
2103 employed. *Environ. Sci. Water Res. Technol.* 2, 1064-1073.

2104 Min, K., Cuiffi, J. D., Mathers, R. T., 2020. Ranking environmental degradation trends of plastic  
2105 marine debris based on physical properties and molecular structure. *Nat. Commun.* 11, 1-11.

2106 Mintenig, S., Int-Veen, I., Löder, M.G., Primpke, S., Gerdts, G., 2017. Identification of  
2107 microplastic in effluents of waste water treatment plants using focal plane array-based micro-  
2108 Fourier-transform infrared imaging. *Water Res.* 108, 365-372.

2109 Mohan, D., Sarswat, A., Ok, Y.S., Pittman Jr., C.U., 2014. Organic and inorganic contaminants  
2110 removal from water with biochar, a renewable, low cost and sustainable adsorbent—a critical  
2111 review. *Bioresour. Technol.* 160, 191-202.

2112 Mohanty, S.K., Valenca, R., Berger, A.W., Iris, K., Xiong, X., Saunders, T.M., Tsang D.C., 2018.  
2113 Plenty of room for carbon on the ground: potential applications of biochar for stormwater  
2114 treatment. *Sci. Total Environ.* 625, 1644-1658.

2115 Mohee, R., Unmar, G., Mudhoo, A., Khadoo, P., 2008. Biodegradability of  
2116 biodegradable/degradable plastic materials under aerobic and anaerobic conditions. *Waste*  
2117 *Manage.* 28, 1624-1629.

2118 Moore, C.J., 2008. Synthetic polymers in the marine environment: a rapidly increasing, long-term  
2119 threat. *Environ. Res.* 108, 131-139.

2120 Moore, C.J., Moore, S.L., Leecaster, M.K., Weisberg, S.B., 2001. A comparison of plastic and  
2121 plankton in the North Pacific central gyre. *Mar. Pollut. Bull.* 42, 1297-1300.

2122 Moussa, D.T., El-Naas, M.H., Nasser, M., Al-Marri, M.J., 2017. A comprehensive review of  
2123 electrocoagulation for water treatment: Potentials and challenges. *J. Environ. Manage.* 186,  
2124 24-41.

2125 Müller, R.J., Kleeberg, I., Deckwer, W.D., 2001. Biodegradation of polyesters containing aromatic  
2126 constituents. *J. Biotechnol.* 86, 87-95.

2127 Murphy, F., Ewins, C., Carbonnier, F., Quinn, B., 2016. Wastewater treatment works (WWTW) as  
2128 a source of microplastics in the aquatic environment. *Environ. Sci. Technol.* 50, 5800-5808.

2129 Murray, F., Cowie, P.R., 2011. Plastic contamination in the decapod crustacean *Nephrops*  
2130 *norvegicus* (Linnaeus, 1758). *Mar. Pollut. Bull.* 62, 1207-1217.

2131 Nel, H., Froneman, P., 2015. A quantitative analysis of microplastic pollution along the south-  
2132 eastern coastline of South Africa. *Mar. Pollut. Bull.* 101, 274-279.

2133 Ngo, P.L., Pramanik, B.K., Shah, K., Roychand, R., Pathway., 2019. classification and removal  
2134 efficiency efficienyof microplastics in wastewater treatment plants. *Environ. Pollut.* 113326.

2135 O'Toole, G., Kaplan, H.B., Kolter, R., 2000. Biofilm formation as microbial development. *Annu.*  
2136 *Rev. Microbiol.* 54, 49-79.

2137 Ouattara, J.M.P., Coulibaly, L., Tiho, S., Gourène, G., 2009. Comparison of macrofauna  
2138 communities in sediments of the beds of vertical flow constructed wetlands planted with  
2139 *Panicum maximum* (Jacq.) treating domestic wastewater. *Ecol. Eng.* 2009; 35: 1237-1242.

2140 Paço, A., Duarte, K., da Costa, J.P., Santos, P.S., Pereira, R., Pereira, M.E, Freitas A.C., Duarte  
2141 A.C., Rocha-Santos T.A., 2017. Biodegradation of polyethylene microplastics by the marine  
2142 fungus *Zalerion maritimum*. *Sci. Total Environ.* 586: 10-15.

2143 Padervand, M., Lichtfouse, E., Robert, D., Wang, C., 2020. Removal of microplastics from the  
2144 environment. A review. *Environ. Chem. Lett.* 1-22.

2145 Park, H.B., Kamcev, J., Robeson, L.M., Elimelech, M., Freeman, B.D., 2017. Maximizing the right  
2146 stuff: The trade-off between membrane permeability and selectivity. *Science.* 356, eaab0530.

2147 .

2148 Paul-Pont, I., Lacroix, C., Fernández, C.G., Hégaret, H., Lambert, C., Le Goïc, N., Frère L,  
2149 Cassone AL, Sussarellu R, Fabioux C, Guyomarch J., 2016. Exposure of marine mussels  
2150 *Mytilus* spp. to polystyrene microplastics: toxicity and influence on fluoranthene  
2151 bioaccumulation. *Environ. Pollut.* 216, 724-737.

2152 Perren, W., Wojtasik, A., Cai, Q., 2018. Removal of microbeads from wastewater using  
2153 electrocoagulation. *ACS Omega.* 3, 3357-3364.

2154 Perz, V., Hromic, A., Baumschlager, A., Steinkellner, G., Pavkov-Keller, T., Gruber, K., Bleymaier  
2155 K, Zitzenbacher S, Zankel A, Mayrhofer C, Sinkel C., 2016. An esterase from anaerobic  
2156 *Clostridium hathewayi* can hydrolyze aliphatic–aromatic polyesters. *Environ. Sci. Technol.* 50,  
2157 2899-2907.

2158 Pinto, F., Franco, C., André, R., Miranda, M., Gulyurtlu, I., Cabrita, I., 2002. Co-gasification study  
2159 of biomass mixed with plastic wastes. *Fuel.* 81, 291-297.



2160 Pivokonsky, M., Cermakova, L., Novotna, K., Peer, P., Cajthaml, T., Janda, V., 2018. Occurrence  
2161 of microplastics in raw and treated drinking water. *Sci. Total Environ.* 643, 1644-1651.

2162 Prata, J. C., da Costa, J. P., Lopes, I., Duarte, A. C., Rocha-Santos, T., 2020. Environmental  
2163 exposure to microplastics: An overview on possible human health effects. *Sci. Total Environ.*  
2164 702, 134455.

2165 Qi, K., Cheng, B., Yu, J., Ho, W., 2017. Review on the improvement of the photocatalytic and  
2166 antibacterial activities of ZnO. *J. Alloys Compd.* 727, 792-820.

2167 Qin, J.J., Kekre, K.A., Tao, G., Oo, M.H., Wai, M.N., Lee, T.C., et al.. 2006. New option of MBR-  
2168 RO process for production of NEWater from domestic sewage. *J. Membr. Sci.* 272, 70-77.

2169 Qin, J-J., Oo, M.H., Lee, H., Kolkman, R., 2004. Dead-end ultrafiltration for pretreatment of RO  
2170 in reclamation of municipal wastewater effluent. *J. Membr. Sci.* 243, 107-113.

2171 Quinn, B., Murphy, F., Ewins, C., 2017. Validation of density separation for the rapid recovery of  
2172 microplastics from sediment. *Anal. Methods.* 9, 1491-1498.

2173 Raju, S., Carbery, M., Kuttykattil, A., Senthirajah, K., Lundmark, A., Rogers, Z., et al. 2020.  
2174 Improved methodology to determine the fate and transport of microplastics in a  
2175 secondary wastewater treatment plant. *Water Res.* 173, 115549.

2176 Roch, S., Friedrich, C., Brinker, A., 2020. Uptake routes of microplastics in fishes: practical and  
2177 theoretical approaches to test existing theories. *Sci. Rep.* 10, 1-12.

2178 Rochman, C.M., Hoh, E., Kurobe, T., Teh, S.J., 2013. Ingested plastic transfers hazardous  
2179 chemicals to fish and induces hepatic stress. *Sci. Rep.* 3, 3263.

2180 Ross, P., van der Aa, L., van Dijk, T., Rietveld, L., 2019. Effects of water quality changes on  
2181 performance of biological activated carbon (BAC) filtration. *Sep. Purif. Technol.* 212, 676-  
2182 683.

2183 Ruan, Y., Zhang, K., Wu, C., Wu, R., Lam, P.K., 2019. A preliminary screening of HBCD  
2184 enantiomers transported by microplastics in wastewater treatment plants. *Sci. Total Environ.*  
2185 674, 171-178.

2186 Rummel, C.D., Jahnke, A., Gorokhova, E., Kühnel, D., Schmitt-Jansen, M., 2017. Impacts of  
2187 biofilm formation on the fate and potential effects of microplastic in the aquatic environment.  
2188 *Environ. Sci. Technol. Letters.* 4, 258-267.

2189 Sadri, S.S., Thompson, R.C., 2014. On the quantity and composition of floating plastic debris  
2190 entering and leaving the Tamar Estuary, Southwest England. *Mar. Pollut. Bull.* 81, 55-60.

2191 Sait, S. T., Sørensen, L., Kubowicz, S., Vike-Jonas, K., Gonzalez, S. V., Asimakopoulos, A. G.,  
2192 Booth, A. M., 2020. Microplastic fibres from synthetic textiles: Environmental degradation  
2193 and additive chemical content. *Environ. Pollut.* 268, 115745.

2194 Scherer, C., Weber, A., Lambert, S., Wagner, M., 2018. Interactions of microplastics with  
2195 freshwater biota. *Freshwater Microplastics*. Springer, Cham, pp. 153-180.

2196 Schwaferts, C., Niessner, R., Elsner, M., Ivleva, N.P., 2019. Methods for the analysis of  
2197 submicrometer-and nanoplastic particles in the environment. *Trends Anal. Chem.* 112, 52-65.

2198 Schwaferts, C., Sogne, V., Welz, R., Meier, F., Klein, T., Niessner, R., Elsner M, Ivleva N.P., 2020.  
2199 Nanoplastic Analysis by Online Coupling of Raman Microscopy and Field-Flow Fractionation  
2200 Enabled by Optical Tweezers. *Anal. Chem.* 92, 5813-5820.

2201 Shah, A.A., Hasan, F., Hameed, A., Ahmed, S., 2008. Biological degradation of plastics: a  
2202 comprehensive review. *Biotechnol. Adv.* 26, 246-265.

2203 Shen, M., Ye, S., Zeng, G., Zhang, Y., Xing, L., Tang, W., Wen, X., Liu, S., 2020. Can microplastics  
2204 pose a threat to ocean carbon sequestration?. *Mar. Pollut. Bull.* 150, 110712.

2205 Shi, Y., Huang, J., Zeng, G., Gu, Y., Chen, Y., Hu, Y., Tang B, Zhou J, Yang Y, Shi L., 2017.  
2206 Exploiting extracellular polymeric substances (EPS) controlling strategies for performance  
2207 enhancement of biological wastewater treatments: an overview. *Chemosphere.* 180, 396-411.

2208 Shim, W.J., Hong, S.H., Eo, S.E., 2017. Identification methods in microplastic analysis: a review.  
2209 *Anal. Methods.* 9, 1384-1391.

2210 Shim, W.J., Song, Y.K., Hong, S.H., Jang, M., 2016. Identification and quantification of  
2211 microplastics using Nile Red staining. *Mar. Pollut. Bull.* 113, 469-476.

2212 Siipola, V., Pflugmacher, S., Romar, H., Wendling, L., Koukkari, P., 2020. Low-Cost Biochar  
2213 Adsorbents for Water Purification Including Microplastics Removal. *Appl. Sci.* 10, 788.

2214 Simon, M., van Alst, N., Vollertsen, J., 2018. Quantification of microplastic mass and removal  
2215 rates at wastewater treatment plants applying Focal Plane Array (FPA)-based Fourier  
2216 Transform Infrared (FT-IR) imaging. *Water Res.* 142, 1-9.

2217 Sjollema, S.B., Redondo-Hasselerharm, P., Leslie, H.A., Kraak, M.H., Vethaak, A.D., 2016. Do  
2218 plastic particles affect microalgal photosynthesis and growth? *Aquat. Toxicol.* 170, 259-261.

2219 Solleiro-Villavicencio, H., Gomez-De León, C. T., Del Río-Araiza, V. H., Morales-Montor, J.,  
2220 2020. The detrimental effect of microplastics on critical periods of development in the  
2221 neuroendocrine system. *Birth Defects Res.* 112, 1326-1340.

- 2222 Sol, D., Laca, A., Laca, A., Díaz, M., 2020. Approaching the environmental problem of  
2223 microplastics: Importance of WWTP treatments. *Sci. Total Environ.* 740, 140016.
- 2224 Sommer, F., Dietze, V., Baum, A., Sauer, J., Gilge, S., Maschowski, C., et al., 2018. Tire abrasion  
2225 as a major source of microplastics in the environment. *Aerosol Air Qual. Res.* 18, 2014-2028.
- 2226 Song, Y.K., Hong, S.H., Jang, M., Han, G.M., Rani, M., Lee, J., et al., 2015. A comparison of  
2227 microscopic and spectroscopic identification methods for analysis of microplastics in  
2228 environmental samples. *Mar. Pollut. Bull.* 93, 202-209.
- 2229 Song, Y.K., Hong, S.H., Jang, M., Kang, J-H., Kwon, O.Y., Han, G.M., Shim W.J., 2014. Large  
2230 accumulation of micro-sized synthetic polymer particles in the sea surface microlayer. *Environ.*  
2231 *Sci. Technol.* 48, 9014-9021.
- 2232 Sornalingam, K., McDonagh, A., Zhou, J.L., 2016. Photodegradation of estrogenic endocrine  
2233 disrupting steroidal hormones in aqueous systems: progress and future challenges. *Sci. Total*  
2234 *Environ.* 550, 209-224.
- 2235 Stolte, A., Forster, S., Gerdts, G., Schubert, H., 2015. Microplastic concentrations in beach  
2236 sediments along the German Baltic coast. *Mar. Pollut. Bull.* 99, 216-229.
- 2237 Su, L., Xue, Y., Li, L., Yang, D., Kolandhasamy, P., Li, D., Shi H., 2016. Microplastics in taihu  
2238 lake, China. *Environ. Pollut.* 216, 711-719.
- 2239 Sun, Q., Ren, S. Y., Ni, H. G., 2020. Incidence of microplastics in personal care products: An  
2240 appreciable part of plastic pollution. *Sci. Total Environ.* 140218.
- 2241 Sussarellu, R., Suquet, M., Thomas, Y., Lambert, C., Fabioux, C., Pernet, M.E., Le Goïc N,  
2242 Quillien V, Mingant C, Epelboin Y, Corporeau C., 2016. Oyster reproduction is affected by  
2243 exposure to polystyrene microplastics. *Proc. Natl. Acad. Sci. U.S.A.* 113, 2430-2435.
- 2244 Tagg, A., Harrison, J.P., Ju-Nam, Y., Sapp, M., Bradley, E.L., Sinclair, C.J., Ojeda J.J., 2017.  
2245 Fenton's reagent for the rapid and efficient isolation of microplastics from wastewater. *Chem.*  
2246 *Commun.* 53, 372-375.
- 2247 Tagg, A.S., Sapp, M., Harrison, J.P., Ojeda, J.J., 2015, Identification and quantification of  
2248 microplastics in wastewater using focal plane array-based reflectance micro-FT-IR imaging.  
2249 *Anal. Chem.* 87, 6032-6040.
- 2250 Talvitie, J., Mikola, A., Koistinen, A., Setälä, O., 2017a. Solutions to microplastic pollution—  
2251 Removal of microplastics from wastewater effluent with advanced wastewater treatment  
2252 technologies. *Water Res.* 123, 401-407.

2253 Talvitie, J., Mikola, A., Setälä, O., Heinonen, M., Koistinen, A., 2017b. How well is microlitter  
2254 purified from wastewater?—A detailed study on the stepwise removal of microlitter in a tertiary  
2255 level wastewater treatment plant. *Water Res.* 109, 164-172.

2256 Tao, L., Wang, Y., Zou, Y., Zhang, N., Zhang, Y., Wu, Y., Wang Y, Chen R, Wang S., 2019. Charge  
2257 Transfer Modulated Activity of Carbon-Based Electrocatalysts. *Adv. Energy Mater.* 1901227.

2258 Teuten, E.L., Saquing, J.M., Knappe, D.R., Barlaz, M.A., Jonsson, S., Björn, A., Rowland S.J.,  
2259 Thompson R.C., Galloway T.S., Yamashita R, Ochi D., 2009. Transport and release of  
2260 chemicals from plastics to the environment and to wildlife. *Philos. Trans. R. Soc. Lond., B,*  
2261 *Biol. Sci.* 364, 2027-2045.

2262 Thompson, R.C., Moore, C.J., Vom Saal, F.S., Swan, S.H., 2009. Plastics, the environment and  
2263 human health: current consensus and future trends. *Philos. Trans. R. Soc. Lond., B, Biol. Sci.*  
2264 364, 2153-2166.

2265 Thompson, R.C., Olsen, Y., Mitchell, R.P., Davis, A., Rowland, S.J., John, A.W., McGonigle D,  
2266 Russell A.E., 2004. Lost at sea: where is all the plastic? *Science.* 304, 838.

2267 Tian, L., Kolvenbach, B., Corvini, N., Wang, S., Tavanaie, N., Wang, L., Ma Y, Scheu S, Corvini  
2268 PF, Ji R., 2017. Mineralisation of <sup>14</sup>C-labelled polystyrene plastics by *Penicillium variable*  
2269 after ozonation pre-treatment. *New. Biotechnol.* 38, 101-105.

2270 Tofa, T.S., Kunjali, K.L., Paul, S., Dutta, J., 2019. Visible light photocatalytic degradation of  
2271 microplastic residues with zinc oxide nanorods. *Environ. Chem. Lett.* 17, 1341-1346.

2272 Turner, A., Holmes, L.A., 2015. Adsorption of trace metals by microplastic pellets in fresh water.  
2273 *Environ. Chem.* 12, 600-610.

2274 Van Cauwenberghe, L., Devriese, L., Galgani, F., Robbens, J., Janssen, C.R., 2015. Microplastics  
2275 in sediments: a review of techniques, occurrence and effects. *Mar. Environ. Res.* 111, 5-17.

2276 Van Doren, L.G., Posmanik, R., Bicalho, F.A., Tester, J.W., Sills, D.L., 2017. Prospects for energy  
2277 recovery during hydrothermal and biological processing of waste biomass. *Bioresour. Technol.*  
2278 225, 67-74.

2279 Von Moos, N., Burkhardt-Holm, P., Köhler, A., 2012. Uptake and effects of microplastics on cells  
2280 and tissue of the blue mussel *Mytilus edulis* L. after an experimental exposure. *Environ. Sci.*  
2281 *Technol.* 46, 11327-11335.

2282 Wang, F., Shih, K.M., Li, X.Y., 2015. The partition behavior of perfluorooctanesulfonate (PFOS)  
2283 and perfluorooctanesulfonamide (FOSA) on microplastics. *Chemosphere.* 119, 841-847.

- 2284 Wang, F., Wong, C.S., Chen, D., Lu, X., Wang, F., Zeng, E.Y., 2018a. Interaction of toxic chemicals  
2285 with microplastics: a critical review. *Water Res.* 139, 208-219.
- 2286 Wang, F., Yang, W., Cheng, P., Zhang, S., Zhang, S., Jiao, W., Sun Y., 2019a. Adsorption  
2287 characteristics of cadmium onto microplastics from aqueous solutions. *Chemosphere.* 235,  
2288 1073-1080.
- 2289 Wang, H., Zhang, Y., Wang, C., 2019b. Surface Modification and Selective Flotation of Waste  
2290 Plastics for Effective Recycling-A Review. *Sep. Purif. Technol.* 226, 75-94.
- 2291 Wang, L., Kaeppler, A., Fischer, D., Simmchen, J., 2019c. Photocatalytic TiO<sub>2</sub> micromotors for  
2292 removal of microplastics and suspended matter. *ACS Appl. Mater. Interfaces.* 11, 32937-32944.
- 2293 Wang, Q., Hernández-Crespo, C., Santoni, M., Van Hulle, S., Rousseau, D.P., 2020a. Horizontal  
2294 subsurface flow constructed wetlands as tertiary treatment: Can they be an efficient barrier for  
2295 microplastics pollution? *Sci. Total Environ.* 137785.
- 2296 Wang, Q., Ye, L., Jiang, G., Jensen, P.D., Batstone, D.J., Yuan, Z., 2013. Free nitrous acid (FNA)-  
2297 based pretreatment enhances methane production from waste activated sludge. *Environ. Sci.*  
2298 *Technol.* 47, 11897-11904.
- 2299 Wang, W., Wang, J., 2018. Investigation of microplastics in aquatic environments: An overview of  
2300 the methods used, from field sampling to laboratory analysis. *Trends Anal. Chem.* 108, 195-  
2301 202.
- 2302 Wang, Y., Zhao, J., Wang, D., Liu, Y., Wang, Q., Ni, B.J., Chen F, Yang Q, Li X, Zeng G, Yuan Z.,  
2303 2018b. Free nitrous acid promotes hydrogen production from dark fermentation of waste  
2304 activated sludge. *Water Res.* 145, 113-124.
- 2305 Wang, Z., Lin, T., Chen, W., 2020b. Occurrence and removal of microplastics in an advanced  
2306 drinking water treatment plant (ADWTP). *Sci. Total Environ.* 700, 134520.
- 2307 Wegner, A., Besseling, E., Foekema, E.M., Kamermans, P., Koelmans, A.A., 2012. Effects of  
2308 nanopolystyrene on the feeding behavior of the blue mussel (*Mytilus edulis* L.). *Environ.*  
2309 *Toxicol. Chem.* 31, 2490-2497.
- 2310 Wei, W., Huang, Q.S., Sun, J., Wang, J.Y., Wu, S.L., Ni, B.J., 2019a. Polyvinyl chloride  
2311 microplastics affect methane production from the anaerobic digestion of waste activated  
2312 sludge through leaching toxic bisphenol-A. *Environ. Sci. Technol.* 53, 2509-2517.

2313 Wei, W., Zhang, Y.T., Huang, Q.S., Ni, B.J., 2019b. Polyethylene terephthalate microplastics affect  
2314 hydrogen production from alkaline anaerobic fermentation of waste activated sludge through  
2315 altering viability and activity of anaerobic microorganisms. *Water Res.* 163, 114881.

2316 Witt, U., Einig, T., Yamamoto, M., Kleeberg, I., Deckwer, W.D., Müller, R.J., 2001.  
2317 Biodegradation of aliphatic–aromatic copolyesters: evaluation of the final biodegradability  
2318 and ecotoxicological impact of degradation intermediates. *Chemosphere.* 44, 289-299.

2319 Yang, L., Li, K., Cui, S., Kang, Y., An, L., Lei, K., 2019. Removal of microplastics in municipal  
2320 sewage from China's largest water reclamation plant. *Water Res.* 155, 175-181.

2321 Yang, L., Zhang, Y., Kang, S., Wang, Z., Wu, C., 2021. Microplastics in freshwater sediment: A  
2322 review on methods, occurrence, and sources. *Sci. Total Environ.* 754, 141948.

2323 Yang, Y., Liu, W., Xu, C., Wei, B., Wang, J., 2017. Antibiotic resistance genes in lakes from middle  
2324 and lower reaches of the Yangtze River, China: effect of land use and sediment characteristics.  
2325 *Chemosphere.* 178, 19-25.

2326 Yang, Y., Yang, J., Wu, W.M., Zhao, J., Song, Y., Gao, L., Yang R., Jian L., 2015. Biodegradation  
2327 and mineralization of polystyrene by plastic-eating mealworms: Part 1. chemical and physical  
2328 characterization and isotopic tests. *Environ. Sci. Technol.* 49, 12080-12086.

2329 Yin, L., Wen, X., Du, C., Jiang, J., Wu, L., Zhang, Y., et al., 2020. Comparison of the abundance  
2330 of microplastics between rural and urban areas: A case study from East Dongting Lake.  
2331 *Chemosphere* 244: 125486.

2332 Yoshida, S., Hiraga, K., Takehana, T., Taniguchi, I., Yamaji, H., Maeda, Y., Toyohara K, Miyamoto  
2333 K, Kimura Y, Oda K., 2016. A bacterium that degrades and assimilates poly (ethylene  
2334 terephthalate). *Science.* 351, 1196-1199.

2335 Yu, F., Yang, C., Zhu, Z., Bai, X., Ma, J., 2019. Adsorption behavior of organic contaminants and  
2336 metals on micro/nanoplastics in the aquatic environment. *Sci. Total Environ.* 694, 133643.

2337 Zhang, K., Shi, H., Peng, J., Wang, Y., Xiong, X., Wu, C., Lam P.K., 2018. Microplastic pollution  
2338 in China's inland water systems: A review of findings, methods, characteristics, effects, and  
2339 management. *Sci. Total Environ.* 630, 1641-1653.

2340 Zhang, X., Chen, J., Li, J., 2020a. The removal of microplastics in the wastewater treatment  
2341 process and their potential impact on anaerobic digestion due to contaminants association.  
2342 *Chemosphere.* 126360.

- 2343 Zhang, X., Zhang, H., Yu, K., Li, N., Liu, Y., Liu, X., Zhang H, Yang B, Wu W, Gao J, Jiang J.,  
2344 2020b. Rapid monitoring approach for microplastics using portable pyrolysis-mass  
2345 spectrometry. *Anal. Chem.* 92, 4656-4662.
- 2346 Zhang, Y., Diehl, A., Lewandowski, A., Gopalakrishnan, K., Baker, T., 2020c. Removal efficiency  
2347 of micro-and nanoplastics (180 nm–125 µm) during drinking water treatment. *Sci. Total*  
2348 *Environ.* 720, 137383.
- 2349 Zhao, S., Zhu, L., Li, D., 2015. Microplastic in three urban estuaries, China. *Environ. Pollut.* 206,  
2350 597-604.
- 2351 Zhao, S., Zhu, L., Li, D., 2016. Microscopic anthropogenic litter in terrestrial birds from Shanghai,  
2352 China: Not only plastics but also natural fibers. *Sci. Total Environ.* 550, 1110-1115.
- 2353 Zheng, Y., Yanful, E.K., Bassi, A.S., 2005. A review of plastic waste biodegradation. *Crit. Rev.*  
2354 *Biotechnol.* 25, 243-250.
- 2355 Ziajahromi, S., Drapper, D., Hornbuckle, A., Rintoul, L., Leusch, F.D., 2020. Microplastic  
2356 pollution in a stormwater floating treatment wetland: Detection of tyre particles in sediment.  
2357 *Sci. Total Environ.* 713, 136356.
- 2358 Ziajahromi, S., Neale, P.A., Rintoul, L., Leusch, F.D., 2017. Wastewater treatment plants as a  
2359 pathway for microplastics: development of a new approach to sample wastewater-based  
2360 microplastics. *Water Res.* 112, 93-99.
- 2361 Ziajahromi, S., Neale, P.A., Telles Silveira, I., Chua, A., Leusch, F.D.L., 2021. An audit of  
2362 microplastic abundance throughout three Australian wastewater treatment plants.  
2363 *Chemosphere* 263, 128294.