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2 impacted groundwater: effects of PFASs properties and groundwater chemistry

3 Hoang Nhat Phong Vo^{1,2}, Thi Minh Hong Nguyen², Huu Hao Ngo^{1*}, Wenshan Guo¹, Pradeep Shukla²

4 ¹Centre for Technology in Water and Wastewater, School of Civil and Environmental Engineering, University of

5 Technology Sydney, Sydney, NSW 2007, Australia

6 ² Queensland Alliance for Environmental Health Sciences (QAEHS), The University of Queensland, 20 Cornwall

- 7 Street, Woolloongabba, Queensland 4102, Australia
- 8

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9 * Corresponding author: *E-mail address: <u>ngohuuhao121@gmail.com</u>*

10 Abstract

The widespread use of per- and polyfluoroalkyl substances (PFASs)-related products such as 11 aqueous film-forming foams (AFFF) has led to increasing contamination of groundwater 12 systems. The concentration of PFASs in AFFF-impacted groundwater can be several orders of 13 magnitude higher than the drinking water standard. There is a need for a sustainable and 14 effective sorbent to remove PFASs from groundwater. This work aims to investigate the 15 16 sorption of PFASs in groundwater by biochar column. The specific objectives are to understand 17 the influences of PFASs properties and groundwater chemistry to PFASs sorption by biochar. The PFASs-spiked Milli-Q water (including 19 PFASs) and four aqueous film-forming foams 18 19 (AFFF)-impacted groundwater were used. The partitioning coefficients (log K_d) of long chain PFASs ranged from 0.77 to 4.63 while for short chain PFASs they remained below 0.68. For 20 21 long chain PFASs (C \geq 7), log K_d increased by 0.5 and 0.8 for each CF₂ moiety of PFCAs and PFSAs, respectively. Dissolved organic matter (DOM) was the most influential factor in 22 PFASs sorption over pH, salinity, and specific ultraviolet absorbance (SUVA). DOM contained 23 24 hydrophobic compounds and metal ions which can form DOM-PFASs complexes to provide

25	more sorption sites for PFASs. The finding is useful for executing PFASs remediation by
26	biochar filtration column, especially legacy long chain PFASs, for groundwater remediation.

27

28 Keywords

29 AFFF, PFASs, biochar, sorption, groundwater, DOM

30 1. Introduction

Aqueous film-forming foams (AFFF) are surfactant products comprising of per- and 31 polyfluoroalkyl substances (PFASs). They are used for extinguishing flammable liquid fuel 32 fires in training and fire-fighting activities at airports and military bases. PFASs seep into 33 groundwater through the soil layer and then in some cases entering the drinking water 34 reservoirs (Baduel et al., 2017; Zhang et al., 2019c). A variety of PFASs has been detected in 35 AFFF-impacted groundwater throughout the world (Dauchy et al., 2019; Sammut et al., 2019; 36 Xu et al., 2021). The concentration of perfluoroalkyl carboxylates (PFCAs) and perfluoroalkyl 37 sulfonates (PFSAs) in groundwater near the firefighting training grounds has been found 38 several orders of magnitude above the drinking water standard of Australia (0.08 µg/L) 39 40 (Bräunig et al., 2017; Sevices, 2016). Exposure to the AFFF-impacted groundwater, has led to the accumulation of certain types of PFASs within humans. Recent reports have linked 41 42 PFASs with thyroid hormone disruption, low activity sperm, diabetes and cancer, especially in the blood serum of firefighters and local people who are frequently or acutely exposed to 43 PFASs (Barton et al., 2020; Donat-Vargas et al., 2019a; Donat-Vargas et al., 2019b; Lin et al., 44 2019; Rotander et al., 2015). 45

A sustainable method is needed for removing PFASs from groundwater. Anion-exchange
resins, polymers and a range of synthetic materials are widely used for the removal of pollutants
by adsorption, however they are costly and not a viable solution for developing countries (Lu

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49 et al., 2020). Biochar is a sustainable carbonaceous sorbent that can be produced locally using agriculture and timber waste, compared to traditional activated carbons which are generally 50 51 produced from fossil fuels (Sørmo et al., 2021). Biochar exhibits comparable PFASs sorption 52 to traditional activated carbon sorbents, and offers substantial benefits such as sustainable values and being an environmentally-friendly alternative (Silvani et al., 2019; Zhang et al., 53 2019b). Biochar has been applied for a wide range of environmental applications such as 54 55 groundwater and soil remediation (e.g., pesticide, heavy metals) and amendment (Singh et al., 2014; Xiao et al., 2017). An agriculture waste-based biochar has been chosen to investigate the 56 57 sorption of PFASs in AFFF-impacted groundwater.

PFASs co-exist in groundwater with other contaminants. The behavior and mechanism of 58 PFASs sorption mainly rely on the characteristics of sorbent, the properties of PFASs, and the 59 matrices of groundwater. Perfluoroalkyl acids (PFAAs) comprise of PFSAs and PFCAs and 60 61 are commonly detected in groundwater (Bräunig et al., 2017; Yong et al., 2021). Studies that cover mechanistic sorption of PFAAs in groundwater are still lacking and the differences of 62 PFCAs and PFSAs in terms of sorption onto biochar sorbents have not been adequately 63 documented (Park et al., 2020; Sørmo et al., 2021). The effect of functional groups and carbon 64 chain length on the sorption of PFASs to biochar needs further elaboration, particularly in the 65 context of groundwater matrices. 66

Some studies that have reported PFASs sorption in groundwater covered sorption to different biochars and a semi pilot test of the process of remediating groundwater by biochar (Kundu et al., 2021; Xiao et al., 2017). However, a study of real-world PFASs-impacted groundwater is lacking because actual groundwater contains several co-contaminants which may influence the effectiveness of the process such as pH, salinity, specific ultraviolet absorbance (SUVA) and dissolved organic matter (DOM). These factors are inter-related, co-exist in groundwater and affect PFASs sorption together in a complex way. For instance, the change of pH in

groundwater partly depends on the deprotonation of organic matter surface charge and ions
(i.e., Ca²⁺, Na⁺). Most of the published studies investigated the effects of those factors without
considering their co-presence in groundwater but using PFASs-spiked MQ water with either
pH, salinity, synthetic DOM (i.e., fulvic and humic) and DOM-free solutions (Gagliano et al.,
2020; Jeon et al., 2011; Nguyen et al., 2020; Yu et al., 2012).

Whether salinity influences PFASs sorption onto carbonaceous materials remains a topic of 79 controversy, potentially due to the complexity of groundwater matrices (Dontsova and Bigham, 80 81 2005; Du et al., 2015; Wu et al., 2020). The molecular structure of hydrophobic DOM is similar to that of PFASs to a certain extent, due to the net negative charge and molecular weight (200 82 - 1,000 Da) (Kothawala et al., 2017). The aromatic DOM contains a hydrophobic backbone 83 like PFASs. These features result in two contradictory influences of DOM on PFASs: 84 competition with PFASs for sorbent sites and providing additional PFAS binding sites on their 85 86 own (Kothawala et al., 2017; McCleaf et al., 2017; Wu et al., 2020; Yu et al., 2012). The competition or mutual support of DOM for PFASs sorption strongly depends on the 87 concentration and composition of DOM in groundwater (i.e. hydrophobicity). Groundwater 88 89 may consist of various organic compounds such as hydrophilic acids, proteins, phenolic groups, amino acids and Fe/Al oxides. DOM is able to form complexes with PFASs either by 90 electrostatic interaction or cation bridging with multivalent ions such as Ca²⁺, Fe³⁺ and Al³⁺ 91 (Gagliano et al., 2020). Overall, these factors (i.e., pH, salinity, SUVA and DOM) have posed 92 93 challenges in interpreting the effects of groundwater chemistry on PFASs sorption. It is 94 necessary to understand the effects of PFASs properties and groundwater chemistry on PFASs sorption to assess the biochar column for practical application. 95

96 Therefore, this work aims to investigate the effects of PFASs properties and groundwater
97 chemistry on PFASs sorption in a rapid small-scale column testing (RSSCT) biochar column.
98 The specific objectives were to (i) determine the influences of PFASs functional groups and

99 carbon chain length of PFCAs and PFSAs, and (ii) study the synergistic/competitive effects of
100 pH, salinity, SUVA and DOM from different AFFF-impacted groundwater sources on PFASs
101 sorption. This study provides important insight into the sorption of PFASs in a biochar column.
102 The sorption behaviour of PFSAs and PFCAs subgroups in a sophisticated groundwater matrix
103 were explained in detail.

104 2. Materials and methods

105 2.1 Standards and reagents

A total of 19 PFAAs were studied, including 11 PFCAs (C₃-C₁₃) and 8 PFSAs (C₄-C₁₂) (Table S1). For quantification, 17 isotopically labelled PFAS standards were used (Table S1). Five other isotopically labelled PFAS standards including 13C₃-PFHxS, 13C₈-PFOA, 13C₃-PFBA, 13C₅-PFPeA, and 13C₈-PFOS were spiked for instrument recovery calculations. All PFAAs (\geq 98% purity) and labelled standards (\geq 99% purity) were purchased from Wellington Laboratories (Ontario, Canada). All the used chemicals were purchased from Sigma-Aldrich of suitable analytical grade and high purity.

113 2.2 Sorbents and groundwater

The biochar was derived from agriculture-related waste and used for the PFASs sorption study. 114 115 The biochar was synthesized from sugarcane bagasse. Sugarcane biochar was chosen for the column study based on the sorption results of the preliminary batch experiment (data not 116 117 shown). The raw material was dried at room temperature and cut into small pieces (1 - 2 cm), 118 washed by distilled water to phase out contaminants, and dried overnight at 55 °C. Next, the 119 material was modified to increase the surface area and pore volume. 10 g of the material was incubated in 1 L MgCl₂ solution (20 % v/v) by shaking at 120 rpm for 24 h. The mixture was 120 121 heated at 80 °C to evaporate the liquid phase. The solid phase was subsequently collected, washed by distilled water and dried at 105 °C over a 6 h period. Finally, the materials were 122

pyrolyzed at 550 °C for 1 h under N_2 gas. The final product was ground, homogenized, and sieved to achieve the effective size (0.5 mm). It was stored in sterile conditions prior to use for this study's experiments.

This study experimented with both PFASs-spiked MQ and AFFF-impacted groundwater. In 126 practice, concentrations of PFASs in the groundwater of Australia can be highly variable 127 ranging from below the limit of detection to 100 µg/L (Phong Vo et al., 2020). The initial 128 concentration of each PFASs in PFASs-spiked MQ was chosen at 10 µg/L. Some groundwater 129 sources possessed long chain PFASs up to C_{14} (Wei et al., 2018), but most of them did not 130 because long chain PFASs were retained by the soil (Bräunig et al., 2017; Murray et al., 2019; 131 Yong et al., 2021). The short chain and long chain PFASs have been defined previously (Buck 132 et al., 2011). The short chain PFCAs and PFSAs have less than six and five perfluorinated 133 carbons, respectively; and vice versa for the long chain PFCAs and PFSAs. 134

We prepared the PFASs-spiked MQ having carbon chain length up to C_{13} for two reasons. The first one is to calculate the log partitioning coefficient (log K_d) of PFCAs and PFSAs in the ideal condition. Then, the log K_d results are used to validate the effect of groundwater chemistry on PFASs sorption. The groundwater samples were collected at four different bores (i.e., B1, B2, B3 and B4) near the firefighting training ground in Queensland (Australia) (Table 1).

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groundwaters (n=2). Numbers in brackets are standard deviation (SD)

Table 1. Concentration of PFASs (µg/L) and other key parameters in four AFFF-impacted

	B1	B2	B3	B4
	24(0.00)	LOD	LOD	LOD
PFBA (C3)	2.4 (0.06)	< LOD	< LOD	< LOD
PFPeA (C4)	4.4 (0.45)	< LOD	< LOD	< LOD
PFHxA (C5)	11.1 (0.05)	< LOD	< LOD	1 (0.01)
PFHpA (C6)	3.3 (0.2)	< LOD	< LOD	< LOD
PFOA (C7)	6.1 (0.22)	0.3 (0.002)	< LOD	0.2 (0.01)

PFNA (C8)	0.4 (0.02)	0.5 (0.003)	< LOD	< LOD
PFDA (C9)	< LOD	0.4 (0.03)	< LOD	< LOD
PFUnDA (C10)	< LOD	< LOD	< LOD	< LOD
PFDoDA (C11)	< LOD	< LOD	< LOD	< LOD
PFTriDA (C12)	< LOD	< LOD	< LOD	< LOD
PFTeDA (C13)	< LOD	< LOD	< LOD	< LOD
PFHxDA (C15)	< LOD	< LOD	< LOD	< LOD
PFODA (C17)	< LOD	< LOD	< LOD	< LOD
PFBS (C4)	2.9 (0.08)	< LOD	< LOD	0.6 (0.01)
PFPeS (C5)	2.6 (0.1)	< LOD	< LOD	0.6 (0.03)
PFHxS (C6)	22.2 (1.2)	0.26 (0.02)	0.7 (0.05)	5.5 (0.2)
PFHpS (C7)	2.4 (0.08)	< LOD	< LOD	0.2 (0.02)
PFOS (C8)	72.1 (0.01)	2.1 (0.03)	0.8 (0.04)	2.8 (0.13)
PFNS (C9)	< LOD	< LOD	< LOD	< LOD
PFDS (C10)	< LOD	< LOD	< LOD	< LOD
PFDoDS (C12)	< LOD	< LOD	< LOD	< LOD
рН	7.8	7.6	7.1	7.1
Salinity (%)	0.26	0.02	0.02	0.22
DOM (mg/L)	4.6 (0.1)	6.5 (0.1)	6.6 (0.1)	10.9 (0.2)
SUVA (L/mg.m)	8.3 (0.1)	4.8 (0.1)	4.8 (0.1)	0.8 (0.1)

142 *Limit of detection (LOD) = $0.1 \, \mu g/L$

143 2.3 Column sorption study

We conducted a PFASs sorption analysis using the biochar-based RSSCT experiments. RSSCT 144 145 is an alternative method which is based on the simulation of a full-scale column. RSSCT is a scaled-down version of the full-scale system and has been applied successfully to predict the 146 breakthrough profiles of PFASs (McCleaf et al., 2017; Park et al., 2020). The RSSCT was 147 designed following the suggestion of previous studies (McCleaf et al., 2017; Park et al., 2020). 148 A syringe (5 ml) was filled with 500 mg sugarcane biochar. The syringe type was selected to 149 150 eliminate the channelling effects because the ratio of column diameter to particle diameter was greater than 50 (Park et al., 2020). The biochar bed was compressed by a piston until it was 151 sufficiently tight enough to create a packed bed of 2.25 cm in height. Five identical columns 152 153 were created for five groundwaters. Characteristics of the groundwater are summarized in

Table 2. The column was activated by MQ water which functioned to remove air in the media
bed, keep it water-saturated and release the potential DOM and alkalinity presented in biochar
(Smebye et al., 2016). A low flow rate of 0.5 ml/min was set to suppress any dispersion effect.
The Reynolds number at this flowrate was 0.05. The linear velocity in the column is 0.438
cm/min.

159 The concentrations of sorbates in the column effluent were used to develop the breakthrough profiles of PFASs as a function of eluted bed volumes. The value of 1 bed volume (BV) was 160 approximately 2.0 mL. Fractions were collected every 20 min into 15 ml polypropylene tubes. 161 The total experiment time was 8 h. Before sampling, all the tubes were centrifuged at 3,000 162 rpm for 20 min at 20 °C, 250 µL of each sample was collected into microcentrifugation tubes 163 and then further centrifuged at 14,000 rpm for 20 min at 20 °C to eliminate residual biochar. 164 The final solute (200 µL) was transferred to a vial containing 200 µL methanol (99.8%, 165 166 LiChrosolv[®], Sigma-Aldrich). The mass-labelled and recovery internal standards were spiked at 4 ng/ml prior to performing LC-MS analysis. 167

168 2.4 Surface area, pore volume, salinity and dissolved organic matter and specific UVA169 analysis

The surface area and pore volume of the sorbents were determined using Brunauer-Emmett-Teller (BET) analyzer (Quantachrome Autosorb IQ, USA) with the sorption/desorption isotherm of nitrogen at 77 K. Firstly, degassing of the sorbents was carried out in vacuum conditions at 473 K and lasted 6 h. Surface area and pore size distribution were calculated via the Density Functional Theory algorithm. The pore size in the sorbent is divided into three groups: micropore (< 2 nm), mesopore (2 – 50 nm) and macropore (>50 nm). This sorbent comprises macropore and mesopore only (Table 2). 178

Numbers in brackets are standard deviation (SD).

Pore size	Surface area distribution (m ² /g)	Pore volume distribution (mm ³ /g)
Micropore (<2 nm)	81.3 (0.15)-	76.25 (4)
Mesopore (2 – 50 nm)	14.8 (0.1)	45.81 (5)
Macropore (>50 nm)	-	-

The salinity of groundwater was measured using a salt meter (Horiba, Japan). Concentrations of DOM were analysed by Multi N/C 3100 (Analytik Jena, Germany). The groundwater samples were pre-filtered before analysis using 0.45 μ m syringe filters (PhenexTM-GF 28 mm) to eliminate the particulate organic matter. In this study, we assumed that DOM was equivalent to dissolved organic carbon (DOC) due to the negligible amount of dissolved organic nitrogen and other compounds in the groundwater (5 – 7%) (Connolly et al., 2020). The SUVA was calculated from Eq. 1 below.

186 SUVA = $\frac{UV_{254}}{DOC} \times 100$ (Eq. 1)

187 Where SUVA is specific UVA (L/mg.m), UV₂₅₄ is UV absorbance at 254 nm (1/cm) and DOC
188 is dissolved organic carbon (mg/L).

189 2.5 Partitioning coefficient of PFASs onto biochar in the column

190 The partitioning coefficient (K_d) was estimated by using breakthrough column 191 chromatographic data. This was achieved by keeping the volumetric flowrate and the carrier 192 flowrate constant. This was possible since the concentration of adsorbed species was 193 significantly small. The PFASs concentration in the pores is assumed to equal to the bulk 194 groundwater concentration. In detail, the partitioning coefficient was calculated by the mass 195 balance equation (Eq. 2) below:

¹⁷⁷ Table 2. Surface area and pore volume distribution of the MgCl₂ modified sugarcane biochar.

196 $\left[\varepsilon_b + (1 - \varepsilon_b)\left(\varepsilon_p + \left(1 - \varepsilon_p\right)K_d\rho_s\right)\right]V = F \int_0^\infty \frac{(c_0 - C)}{C} dt$

197

198 Where K_d (L/kg) is the partitioning coefficient, V is the column volume, F is the volumetric 199 flowrate, C is the equilibrium or near-equilibrium concentration (at 8h), C₀ is the feed 200 concentration, ε is the porosity (b for bed and p for particle) and ρ_s is the density of sorbent.

201 2.6 Quantitative LC-MS/MS analysis

202 The PFASs analysis method was described recently elsewhere (Bräunig et al., 2019; Nguyen et al., 2020). PFASs were analysed using high performance liquid chromatography (Nexera 203 HPLC, Shimadzu Corp., Japan) coupled to a tandem mass spectrometer (QTrap 5500 AB-204 Sciex, Ontario, Canada). The polar C18 Luna Omega column (100 x 2.1 mm) had a particle 205 size 1.6 µm, and pore size of 100 Å. The mobile phases consist of A phase of 10 mM 206 ammonium acetate in Milli-Q water, acetonitrile and acetic acid (94.9:5:0.1, v/v), and B phase 207 of 0.5 mM ammonium acetate in acetonitrile, Milli-Q water and acetic acid (94.9:5:0.1, v/v). 208 The negative electrospray ionization mode was employed. The injection volume was 10 µL 209 holding at a temperature of 50 °C. A pretreatment column (C18, 50 \times 4.6 mm, 5 μ m, 210 211 Phenomenex, Lane Cove, Australia) was set up between the solvent and injector to delay PFASs background. PFASs quantification was calculated by mass-labelled internal standard. 212 An eight-point calibration standard was prepared with concentrations ranging from 0.1 to 100 213 214 $\mu g/L.$

For quantitative analysis, the calibration standards were weighted linearly through 1/xregression. The requirement for regression coefficients was ≥ 0.993 . Analytical replications were performed once in every five injections together with two points of the calibration standards being reanalysed after 10 samples. The full calibration standards were analysed both at the beginning and the end of each analytical batch. A maximum variation of 10% among

analytical replications was considered acceptable. Procedural blanks and samples spiked with

a known amount of PFASs were also performed in duplicate. The limits of detection (LODs),

and limits of quantification (LOQs) were reported elsewhere (Nguyen et al., 2020).

223 2.7 Principal component analysis

Origin Pro 2017 (Origin Lab) was used to perform principal component analysis (PCA) analysis and evaluate the correlation of partitioning coefficients K_d of PFCAs/PFSAs and selected groundwater chemistry factors (i.e., pH, salinity, SUVA and DOM). The dataset for PCA analysis is summarized in Table S5.

228 3. Results and discussion

3.1 Partitioning coefficients of PFASs onto biochar in PFAS-spiked MQ water and AFFF-impacted groundwater

To elaborate on the sorption behaviour of PFASs, partitioning coefficients (presented as log 231 232 K_d) of PFASs between biochar and the aqueous phase were calculated (Fig. 1). Generally, the PFSAs demonstrated higher log K_d than PFCAs given a similar perfluorinated carbon chain 233 length. For example, in the PFASs-spiked MQ samples, log K_d of PFOS was 1.57 log units 234 235 which were 1.3 times higher than the log K_d of PFNA (1.18 log units). Also, the long chain PFASs exhibited better partitioning on the sorbent compared to short chain PFASs. There is a 236 trend of increasing log K_d with the increasing numbers of perfluorinated carbons. Log K_d of 237 long chain PFASs ranged from 0.77 to 4.63 log units, while for short chain PFASs, log Kd 238 stayed below 0.68 log units. The trends were consistent for both PFASs-spiked MQ and AFFF-239 impacted groundwater and agreed with previous studies (Oliver et al., 2020; Xiao et al., 2017; 240 Zhang et al., 2019a). Further discussion on the influence of functional groups and carbon chain 241 length on PFASs sorption is provided in section 3.2. 242



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Fig. 1. Log K_d for sorption of PFCAs and PFSAs in PFAS-spiked MQ water and AFFF-impacted groundwater onto biochar. PFASs in each groundwater type were examined in distinct colors and symbols. All measures show significant differences (p<0.01). The K_d values of the PFASs-spiked MQ and AFFF-impacted groundwater are given in detail in Table S4. All measures showed significant differences (p < 0.05) amongst log K_d of groundwater sources.

249 3.2 Effects of PFASs properties

250 3.2.1 Functional groups

The functional groups consist of the carboxylic group for PFCAs and the sulfonic group for 251 PFSAs. The PFSAs possessed higher log K_d values than PFCAs given a similar number of 252 perfluorinated carbons (Fig. 1 and Table S4). From Fig. 2, it shows a linear correlation of log 253 K_d and retention time of PFASs in the LC-MS column (Adjusted $R^2 = 0.79 - 0.83$). The 254 retention time of PFAS in the LC-MS column is a typical proxy indicating the hydrophobicity 255 of PFASs (Xiao et al., 2017). The slope of PFCAs linear fit was 1.18 which was slightly less 256 257 than the value of PFCAs, being 1.45. It indicated that PFSAs were retained much more than PFCAs; in fact, it was 1.3-fold due to the higher hydrophobicity. The smaller size of the 258

259 carboxylic functional group of PFCAs possibly made it less hydrophobic than the sulfonic functional group (Oliver et al., 2020). Recently, Wang et al. (2019) stated that PFSAs exhibited 260 higher sorption affinity than PFCAs. Park et al. (2020) established the correlation between half-261 breakthrough bed volume (BV₅₀) and octanol-water partition coefficient (log D_{ow}) of PFASs 262 to predict the sorption behaviour, but it was monotonic and did not reflect the discrepancy of 263 PFCAs and PFSAs that we found. Higgins and Luthy (2006) reported that PFSAs showed 1.7-264 fold higher hydrophobicity than PFCAs which was slightly higher than our value. This is 265 probably attributed to the discrepancies of soil and groundwater matrices, and properties of 266 267 sorbents and feedstocks used for producing the biochar. For instance, the surface area of pinewood biochar of 351 m²/g was 4 times higher than the sugarcane biochar of this study 268 (Xiao et al., 2017). The pinewood biochar of Xiao et al. (2017) was prepared by gasification 269 270 method, which differs from the pyrolysis method of this study. The other types of biochar prepared by pyrolysis method showed less surface area than our biochar. Higher surface area 271 could provide more binding sites for PFAS even other larger sorbates such as DOM could block 272 the micropore (Ando et al., 2010; Wu et al., 2020). The finding indicates the characteristics of 273 biochar which defines the PFAS sorption efficiency also depends on the methods used to 274 produced biochar (Muvhiiwa et al., 2019). 275



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Fig. 2. Correlation of PFASs retention time in the LC-MS column and log K_d of PFCAs and PFSAs groups. The
details of PFASs retention time in a LC-MS column can be found in Table S5. The log K_d values were obtained
from PFAS-spiked MQ water.

280 3.2.2 Carbon chain length

281 The longer chain PFASs of the same group showed higher log K_d values resulting from the extensive dominance of long chain PFASs on sorbent sites and pores over short chain PFASs 282 (Xiao et al., 2017). The increase of $\log K_d$ per CF₂ moiety also differs between the short chain 283 and long chain PFASs. For short chain PFASs, the increasing trend of log K_d per CF₂ moiety 284 is not clearly observed which is potentially due to the low sorption efficiency and 285 286 hydrophobicity of short chain PFASs. For long chain PFASs, log K_d increases 0.5 and 0.8 for each CF₂ moiety of PFCAs and PFSAs respectively. It is caused by the increase of 287 hydrophobicity of long chain PFASs proportional to the amount of perfluorinated carbons, 288 289 which refers to the rule of additivity (Park et al., 2020). The observation of increasing log K_d overall agrees with other studies, but log K_d values are sorbent specific (Park et al., 2020; Xiao 290 et al., 2017). Some sorbents exhibit higher sorption affinity with more porosity. For instance, 291

commercial activated carbon (F300) achieved 4 to 5.5 log K_d for short chain PFASs which is much higher than the biochar from this study (Xiao et al., 2017). However, the flow patterns of sorption experiment also pose certain influence on the value of log K_d . Although the K_d calculation of this study primes by the equilibrium sorption, the K_d values might be underestimated by higher flow rates which would restrain the diffusion of PFAS to the micropore.

The breakthrough profiles of selected PFASs in PFASs-spiked MQ water and AFFF-impacted 298 groundwater are shown in Fig. 4 and Fig. S1. The shorter chain PFASs demonstrated shallower 299 breakthrough curves and smaller BV₅₀ values. For example, in the B4 samples, the order of 300 BV₅₀ values was PFOS>PFHxS>PFHxA which agreed with the effects of PFASs properties on 301 sorption behavior. The data reported in other studies agreed with what we have found 302 (Dalahmeh et al., 2019; Liu et al., 2019; Park et al., 2020). However, the amount of BV in this 303 304 study was less than what other researches documented (McCleaf et al., 2017; Park et al., 2020). The threshold of BV in this RSSCT column was 120 while the noted studies reported BV up 305 to 150,000. The operation of RSSCT and BV₅₀ depended on various engineering factors such 306 as initial PFASs concentrations, numbers of PFASs, depth of the packed bed, and 307 characteristics of the sorbent (Murray et al., 2019; Xiao et al., 2017). For instance, we 308 experimented with 19 PFASs substances, having an initial concentration of 10 µg/L. Park et al. 309 (2020) and McCleaf et al. (2017) used the initial PFASs concentration ranging from 0.1 to 0.35 310 μ g/L which was less than our values by 30- to 100-fold. 311

312 3.3 Effects of groundwater chemistry

313 3.3.1 Contributions of factors in groundwater to PFASs sorption

314 The matrix of groundwater is diverse and has a complex effect on the sorption of PFAS to

biochar. Groundwater chemistry has been widely recognized as exerting effects on the sorption

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of the charged organic molecules (Higgins and Luthy, 2006; Park et al., 2020). To evaluate the
effects of groundwater chemistry on PFASs sorption, we conducted PCA analysis to describe
the relative correlation between K_d of PFCAs/PFSAs groups and the groundwater chemistry
factors (i.e., pH, salinity, SUVA and DOM) (Fig. 3).

The first factor is pH which ranged from 7.1 to 7.8 in the four groundwater samples. The pKa values of all PFASs in this study were below 2 which meant that the speciation of PFASs was unchanged (Ahrens et al., 2012; Goss, 2008). It means any pH influences observed are attributed to the pH-related changes in the sorbents, such as organic matter charge, and less likely due to protonation/deprotonation of PFASs (Higgins and Luthy, 2006). The salinity posed certain effects on PFASs sorption and was strongly related to the presence of divalent cations in groundwater such as Ca^{2+} and Mg^{2+} (Wu et al., 2020).

Monovalent ions did not reveal a particular influence on anionic PFASs sorption but divalent 327 ions did (Higgins and Luthy, 2006). The divalent cations were present at high concentrations 328 in B1 and B4 groundwater samples (i.e., 7 - 419 mg Ca^{2+}/L and 1 - 277 mg Mg^{2+}/L) (Army 329 Aviation Centre Oakey, 2019). The increase in Ca^{2+} enhanced sorption of anionic sorbate onto 330 the sorbent due to the salting out and cation-bridging effects (Dontsova and Bigham, 2005; Du 331 332 et al., 2015). Salinity itself does not adequately explain the increase of K_d but it should be placed in a joint context with SUVA and DOM. The divalent cations inter-correlated with 333 DOM by influencing the deprotonation of DOM (Jeon et al., 2011). B1, B2 and B3 samples 334 contained high aromaticity (SUVA>2 L/mg.m) whereas B4 possessed lower aromaticity 335 (SUVA<2 L/mg.m). The aromatic substances are hydrophobic which attract PFASs and 336 337 increase PFASs sorption (Park et al., 2020; Wu et al., 2020). However, our PCA analysis shows that SUVA influences PFASs sorption to a lesser extent than salinity and DOM. 338

The PCA analysis suggests that DOM influences PFASs sorption the most critically in this study. According to Appleman et al. (2013), a DOM concentration of 1.7 mg/L could increase

the breakthrough time of PFASs compared to deionized water. The DOM concentrations of 341 four AFFF-impacted groundwater sources in this study were more than 2 mg/L, indicating a 342 significant impact of DOM on the sorption behaviour of PFASs. In Fig. 1, log K_d values of 343 PFASs in the AFFF-impacted groundwaters exceeded the PFASs-spiked MQ 1 - 1.5 logs. The 344 higher DOM concentration in groundwater samples resulted in higher log K_d values. Kothawala 345 et al. (2017) and McCleaf et al. (2017) agreed with our result that the increase of DOM in water 346 347 samples facilitates PFASs sorption. McCleaf et al. (2017) has found that after operating the column for 50d, the removal efficiency of PFAS increased which might be attributed to the 348 349 loading of DOM on sorbent and agglomeration of PFAS. Some studies reported different findings that DOM did not affect PFAS sorption or could reduce it (Wu et al., 2020; Yu et al., 350 2012). For instance, Wu et al. (2020) performed a PCA analysis and concluded that DOM did 351 352 not perform inhibition to PFAS sorption. However, the analysis of Wu et al. (2020) used the data of both positive charged (β-cyclodextrin polymers) and negative charged (activated 353 carbons) sorbents. Wu et al. (2020) also indicated that DOM may only influence PFAS sorption 354 on negative charged sorbent like our case. Yu et al. (2012) investigated that DOM impaired the 355 PFAS sorption from 15% to 30% due to the pore blockage and competition of sorbent sites by 356 DOM. The degree of DOM effect depended on the molecular size distribution of DOM. DOM 357 having size larger than 30 kDa showed similar sorption efficiency to DOM-free solution; in 358 turn, low molecular weight DOM (<1 kDa) demonstrated greater effect. It implied that the 359 360 effect of DOM on PFAS sorption was also subjected to the specific characteristic of groundwater. In addition, Yu et al. (2012) used the DOM-spiked artificial groundwater which 361 ignored other matrices, while our study, Kothawala et al. (2017) and McCleaf et al. (2017) 362 experimented with the real surface and groundwater. The potential reason for the disparity of 363 those studies is the pollutants transport patterns of sorption studies (i.e., batch and column, flow 364 rate) which undermine the influence of DOM. Another important underlying reason is that 365

multivalent cations presented in real groundwater can form DOM-PFASs complexes whichretain much more PFASs in the column (section 3.3.2).

This PCA analysis includes 6 variables and 4 observations which are similar to Wu et al. (2020) consisting of 7 variables and 12 observations (i.e., 3 sorbents in 4 groundwater sources). This study focuses on only one type of sorbent so the number of observations is less than Wu et al. (2020), which is a limit that needs to be addressed in the future.

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Fig. 3. PCA biplot showing the correlations of the groundwater chemistry and the partitioning coefficients per CF₂ moiety of PFCAs and PFSAs (calculated by Eq. S1 and Eq. S2). The PCA was created by the multidimensional dataset with 6 variables and 4 observations. The variables are plotted by the straight arrows. Correlations between the variables are shown through the angles in between the two arrows. The observations were plotted as points in different colors and symbols. The degrees of observations for the variables are demonstrated via the relative distance of the points to the arrows.

380 3.3.2 How DOM influence PFASs sorption?

381 The effects of DOM on the sorption behavior of PFASs are indicated through the breakthrough

382 profiles (Fig. 4). For PFHxA, PFHxS and PFOA, the effect of DOM on the breakthrough

profiles was not significantly different (Fig. 4a, b, c). Importantly, the breakthrough profiles of PFOS did not follow the general trend like other PFASs and this requires further elaboration (Fig. 4d). In B4 sample, PFOS reached BV₅₀ more slowly and did not reach the point of exhaustion within the studied timeframe. The discrepancy in the PFOS breakthrough profiles was strongly attributed to the variation of DOM levels presented in four groundwater sources.



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Fig. 4. Breakthrough profiles of selected PFASs in the AFFF-impacted groundwater sources: PFHxA (panel A),
PFHxS (panel B), PFOA (panel C), and PFOS (panel D). The PFASs collected in the similar location are denoted
with an identical symbol and color pattern. Concentrations of the selected PFASs in the panels are beyond the
detection limit (>0.1 µg/L).

393 DOM demonstrated a significant influence on the breakthrough behavior of PFASs showed 394 through the correlation of BV_{50} and DOM concentrations (Appleman et al., 2013; McCleaf et 395 al., 2017). The BV_{50} was selected for establishing the correlation with DOM because it 396 reflected the apparent PFASs sorption capacity of biochar (Corwin and Summers, 2011). 397 Interestingly, we found that the concentration of DOM correlated significantly to BV_{50} of PFOS

398 (Fig. 5a) ($R^2=0.68$). This was despite the difference in the initial concentration of PFASs and 399 other types of interference, for example pH. This correlation has confirmed the significant 400 effect of DOM on sorption of PFASs in groundwater, rather than pH, at least in the scope of 401 this research.

402 The composition of DOM also played an important role in the sorption of PFASs. This phenomenon has not been fully explained in previous studies. With UV scanning, we 403 discovered that groundwater samples contained large amounts of hydrophobic DOM (Fig. 5b). 404 405 The Bz band (190-240 nm) presented the aromatic compounds based on the transition and vibration perturbation of the II-e system (Korshin et al., 1997). The absorbances of all samples 406 are higher than 0.5, possibly containing hydrophobic DOM which can form DOM-PFASs 407 complexes and increase PFAS sorption. The ET band (240-400 nm) exhibited hydrophilic 408 functional groups such as hydroxyl, carboxyl, carbonyl and ester varieties. The absorbances of 409 410 the hydrophilic regions are all lower than 0.5, thus demonstrating the minor hydrophilic DOM fraction of the groundwater samples. 411

DOM competes with PFASs and occupies the sorption sites within the pores of the sorbent. At 412 neutral pH, DOM was deprotonated so that it was negatively charged creating electrostatic 413 414 repulsion between the negatively charged functional group of PFASs (Choppin and Kullberg, 1978). The short chain PFASs were subject to electrostatic repulsion and consequently 415 absorbed less than the long chain PFASs. However, DOM could provide more sorption sites 416 for PFASs on their own surfaces. Those DOM were hydrophobic which meant they were able 417 to entrap hydrophobic PFASs and form DOM-PFASs complexes on the uncharged sites. DOM 418 can also create cation bridging with multivalent ions such as Fe³⁺ and Al³⁺ (Gagliano et al., 419 2020). For example, the carboxyl group of PFOA could create a complex structure with Fe and 420 Al oxide in groundwater (Xiao et al., 2019). DOM is present in groundwater in much higher 421 422 concentrations than PFASs and therefore retained PFASs extensively, proportional to the

423 increase of DOM. This was the solid-phase dissolution process and the main reason for the increase of K_d values in higher DOM concentrations. In this study, we exclude the possibility 424 of DOM desorption in the eluent because biochar was washed and activated with MQ water. 425 426 Previous work has shown that washing reduced alkalinity of biochar leading to the reduction of final pH in the mixture of biochar and groundwater (Smebye et al., 2016). In addition, the 427 biochar of this study was modified with MgCl₂ to have positive charge sites for sorption of 428 DOM and PFASs, even at high pH (Fang et al., 2020). Hence, the effect of DOM desorption 429 on the result is not apparent. 430

To further verify this assertion, sorbent was collected at the top and middle of the RSSCT 431 media bed and processed through FTIR (Fig. 5c, d). The spectra of 1390 cm⁻¹ in both 432 groundwater samples indicated the presence of monodentate which is a complex of iron 433 structure and the PFASs carboxyl group. On the FTIR spectra, several bands could be observed, 434 indicating the sorption of PFASs and organic matter on sorbents. Bands observed at 2992 cm⁻ 435 ¹ and 3672 cm⁻¹ depicted C-H and O-H stretches of aromatic and phenol substances, 436 respectively. Those bands coincided with the data produced by UV scanning and confirmed 437 438 the competitive sorption of PFASs and hydrophobic organic matter in groundwater samples. Likewise, the bands of 1078 cm⁻¹, 1250 cm⁻¹, 1580 cm⁻¹ and 873 cm⁻¹ represent sulfonic group, 439 C-F stretching, C=O stretching of carboxylic group and absorbance of CF_2 and CF_3 , 440 respectively (Guo et al., 2017). 441



Fig. 5. Correlation of DOM in groundwater samples and BV₅₀ of PFOS in RSSCT (panel A), UV scan profiles of
groundwater samples (panel B), FTIR scan of sorbent on top and middle of RSSCT media bed of the B1 sample
(panel C) and B4 sample (panel D). The dashed line and shaded area in panel A show linear fit and 95% confidence
band, respectively. The y axis presents the DOM concentration (panel A), absorbance (panel B) and transmittance
(panel C, D).

448 3.4 Environmental implications

The finding of this study is subjected to only one biochar (sugarcane bagasse pyrolyzed at 550 449 °C). In essence, future studies need to investigate the effect of biochar produced from various 450 feedstocks (e.g., corn straw, timber, biosolid) and processes (e.g., pyrolyzed temperature and 451 pretreatment methods). For example, corn straw-derived biochar showed higher PFAS sorption 452 453 efficiency at higher pyrolysis temperatures (700 °C and 550 °C)(Guo et al., 2017). The higher pyrolysis temperature could increase surface area and pore volume of biochar up to 10-fold 454 (Guo et al., 2017). At 500 °C, pyrolysis can be combined with biochar pretreatment by chemical 455 456 activation, such as ZnCl₂ and H₂SO₄, to increase microporosity (Martin et al., 1996). Those

experiments, together with this study, will provide adequately the sorption capacity of biocharand relevant mechanisms of PFAS sorption.

The data presented demonstrate a potential control alternative to alleviate the risk for 459 communities who are exposed to PFASs-impacted groundwater. It provides an insight into the 460 understanding and applicability of PFASs sorption by biochar which could be used for 461 462 upscaling and field deployment. The removal of long chain PFASs by biochar is viable. The application can be further extended for sorption of emerging PFASs such as other anionic, 463 nonionic and zwitterionic variations whose toxicity and bioaccumulation potential in AFFF-464 impacted groundwater are not fully investigated. Further validating research is also required. 465 Although other sorbents (i.e., ion exchange resin, β -cyclodextrin, Rembind®) and technologies 466 (membrane, advanced oxidation process) might be more effective in PFASs remediation, 467 biochar is more beneficial in cost and sustainable values (Phong Vo et al., 2020). For instance, 468 469 the commercial price of biochar is 246 - 500/tonne which was 3-5 times less than activated 470 carbon (\$1500 US/tonne) (Ahmad et al., 2014; Mohan et al., 2014). Sørmo et al. (2021) applied 5% biochar dose to immobilize PFASs leaching from the soil, while Hale et al. (2017) did for 471 472 3% activated carbon. In practice, the applied dose of biochar depended on several factors (e.g., sorbent characteristic and water matrix) which would decide the ultimate cost of using biochar 473 for PFAS remediation. Apart from soil immobilization, biochar can also be reused in thermal 474 treatment. The addition of activated biochar significantly enhances the sorption of gas-phase 475 476 PFAS on biochar and increases PFAS decomposition (Sasi et al., 2021). The application of this 477 biochar depends on the availability of raw material, environmental protection strategy of local government and expectation of end-users. This sugarcane bagasse biochar is potential for 478 regions where sugarcane is the main agricultural product. 479

Effects of groundwater matrix, particularly DOM, is an important point to look at beforedesigning field scale works. This study provides mechanistic interaction of DOM and PFASs

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482 in various groundwater matrices. Some of the breakthrough profiles are however nearequilibrium conditions which needs further elaboration as, in practice, the near one is not 483 expected to happen. Batch experiments do not completely show the effect of DOM on the 484 sorption of pollutants due to different pollutants transport pattern. For instance, flow rate in the 485 column is an important factor determining the diffusion of pollutants into micropore. It is 486 challenging to use the results of batch conditions for upscaling, but the result of a column study 487 488 could work (Kothawala et al., 2017). This would be a motivation for future studies to uncover the effects of groundwater matrix to PFASs sorption. In addition, attention to the engineering 489 490 factors of the column such as sorbent bed depth, hydraulic retention time, recirculation potential and sorbent regeneration is required. 491

492 4. Conclusions

This study investigates the effects of PFASs properties and groundwater chemistry on PFASs
sorption in a biochar column. By using PFAS-spiked MQ and AFFF-impacted groundwater,
the following important conclusions can be made:

The sorption of PFSAs towards biochar sorbent was 1.3-fold higher than that of PFCAs.
 Log K_d of long chain PFASs ranged from 0.77 to 4.63 log units while it was below 0.68
 log units for short chain PFAS. Log K_d values of PFASs in the real AFFF-impacted
 groundwater were higher than the PFASs-spiked MQ water 1 - 1.5 log magnitude due
 to the effect of the groundwater matrix.

Amongst the selected factors in groundwater (i.e., pH, salinity, SUVA, DOM), DOM
 had the most significant influence on PFAS sorption determined via PCA analysis.
 DOM contained hydrophobic compounds and complexes (e.g., monodentate) which
 could compete with PFASs and also provide sorption sites for PFAS.

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505 Further works are suggested for a field-scale study to fully evaluate the effects of 506 environmental matrix and applicability of biochar for AFFF-impacted water sources.

507

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512 References

AECOM, 2016. Stage 2C Environmental Investigation. Available from:
 https://www.defence.gov.au/Environment/PFAS/Docs/Oakey/FactSheets/OakeyFactSheetJuly2016.pdf.
 Accessed 21 May 2021

Ahmad, M., Rajapaksha, A.U., Lim, J.E., Zhang, M., Bolan, N., Mohan, D., Vithanage, M., Lee, S.S., Ok,
 Y.S., 2014. Biochar as a sorbent for contaminant management in soil and water: A review. *Chemosphere*,
 99, 19-33.

Ahrens, L., Harner, T., Shoeib, M., Lane, D.A., Murphy, J.G., 2012. Improved Characterization of Gas–
 Particle Partitioning for Per- and Polyfluoroalkyl Substances in the Atmosphere Using Annular Diffusion
 Denuder Samplers. *Environ. Sci. Technol.*, 46(13), 7199-7206.

Ando, N., Matsui, Y., Kurotobi, R., Nakano, Y., Matsushita, T., Ohno, K., 2010. Comparison of natural
 organic matter adsorption capacities of super-powdered activated carbon and powdered activated Carbon.
 Water Res., 44(14), 4127-4136.

- 5. Appleman, T.D., Dickenson, E.R.V., Bellona, C., Higgins, C.P., 2013. Nanofiltration and granular activated
 carbon treatment of perfluoroalkyl acids. *J. Hazard. Mater.*, 260, 740-746.
- 527 6. Army Aviation Centre Oakey, A. 2019. Groundwater and Surface Water Monitoring: April/May 2019.
- 528 7. Baduel, C., Mueller, J.F., Rotander, A., Corfield, J., Gomez-Ramos, M.-J., 2017. Discovery of novel per-
- 529 and polyfluoroalkyl substances (PFASs) at a fire fighting training ground and preliminary investigation of
- their fate and mobility. *Chemosphere*, 185, 1030-1038.

- 8. Barton, K.E., Starling, A.P., Higgins, C.P., McDonough, C.A., Calafat, A.M., Adgate, J.L., 2020.
 Sociodemographic and behavioral determinants of serum concentrations of per- and polyfluoroalkyl
 substances in a community highly exposed to aqueous film-forming foam contaminants in drinking water.
 Int. J. Hyg. Environ. Health, 223(1), 256-266.
- 9. Bräunig, J., Baduel, C., Barnes, C.M., Mueller, J.F., 2019. Leaching and bioavailability of selected
 perfluoroalkyl acids (PFAAs) from soil contaminated by firefighting activities. *Sci. Total Environ.*, 646, 471479.
- 538 10. Bräunig, J., Baduel, C., Heffernan, A., Rotander, A., Donaldson, E., Mueller, J.F., 2017. Fate and
 539 redistribution of perfluoroalkyl acids through AFFF-impacted groundwater. *Sci. Total Environ.*, 596-597,
 540 360-368.
- 541 11. Buck, R.C., Franklin, J., Berger, U., Conder, J.M., Cousins, I.T., de Voogt, P., Jensen, A.A., Kannan, K.,
 542 Mabury, S.A., van Leeuwen, S.P., 2011. Perfluoroalkyl and polyfluoroalkyl substances in the environment:
 543 terminology, classification, and origins. *Integr. Environ. Assess. Manag.*, 7(4), 513-41.
- 544 12. Choppin, G.R., Kullberg, L., 1978. Protonation thermodynamics of humic acid. *J. Inorg. Nucl. Chem.*, 40(4),
 545 651-654.
- 546 13. Connolly, C.T., Cardenas, M.B., Burkart, G.A., Spencer, R.G.M., McClelland, J.W., 2020. Groundwater as
 547 a major source of dissolved organic matter to Arctic coastal waters. *Nature Comm.*, 11(1), 1479.
- 548 14. Corwin, C.J., Summers, R.S., 2011. Adsorption and desorption of trace organic contaminants from granular
 549 activated carbon adsorbers after intermittent loading and throughout backwash cycles. *Water Res.*, 45(2),
 550 417-426.
- 551 15. Dalahmeh, S.S., Alziq, N., Ahrens, L., 2019. Potential of biochar filters for onsite wastewater treatment:
 552 Effects of active and inactive biofilms on adsorption of per- and polyfluoroalkyl substances in laboratory
 553 column experiments. *Environ. Pollut.*, 247, 155-164.
- 16. Dauchy, X., Boiteux, V., Colin, A., Hémard, J., Bach, C., Rosin, C., Munoz, J.-F., 2019. Deep seepage of
 per- and polyfluoroalkyl substances through the soil of a firefighter training site and subsequent groundwater
 contamination. *Chemosphere*, 214, 729-737.

- 557 17. Donat-Vargas, C., Bergdahl, I.A., Tornevi, A., Wennberg, M., Sommar, J., Kiviranta, H., Koponen, J.,
 558 Rolandsson, O., Åkesson, A., 2019a. Perfluoroalkyl substances and risk of type II diabetes: A prospective
- nested case-control study. *Environ. Int.*, 123, 390-398.
- 560 18. Donat-Vargas, C., Bergdahl, I.A., Tornevi, A., Wennberg, M., Sommar, J., Koponen, J., Kiviranta, H.,
- Åkesson, A., 2019b. Associations between repeated measure of plasma perfluoroalkyl substances and
 cardiometabolic risk factors. *Environ. Int.*, 124, 58-65.
- 563 19. Dontsova, K.M., Bigham, J.M., 2005. Anionic Polysaccharide Sorption by Clay Minerals. *Soil Sci. Soc. Am.*564 *J.*, 69(4), 1026-1035.
- 20. Du, Z., Deng, S., Chen, Y., Wang, B., Huang, J., Wang, Y., Yu, G., 2015. Removal of perfluorinated
 carboxylates from washing wastewater of perfluorooctanesulfonyl fluoride using activated carbons and
 resins. J. Hazard. Mater., 286, 136-143.
- 568 21. Fang, L., Li, J.-s., Donatello, S., Cheeseman, C.R., Poon, C.S., Tsang, D.C.W., 2020. Use of Mg/Ca modified
 569 biochars to take up phosphorus from acid-extract of incinerated sewage sludge ash (ISSA) for fertilizer
 570 application. *J. Cleaner Prod.*, 244, 118853.
- 571 22. Gagliano, E., Sgroi, M., Falciglia, P.P., Vagliasindi, F.G.A., Roccaro, P., 2020. Removal of poly- and
 572 perfluoroalkyl substances (PFAS) from water by adsorption: Role of PFAS chain length, effect of organic
 573 matter and challenges in adsorbent regeneration. *Water Res.*, 171, 115381.
- 574 23. Goss, K.-U., 2008. The pKa Values of PFOA and Other Highly Fluorinated Carboxylic Acids. *Environ. Sci.*575 *Technol.*, 42(2), 456-458.
- 576 24. Guo, W., Huo, S., Feng, J., Lu, X., 2017. Adsorption of perfluorooctane sulfonate (PFOS) on corn straw577 derived biochar prepared at different pyrolytic temperatures. *J. Taiwan Inst. Chem. E.*, 78, 265-271.
- 578 25. Hale, S.E., Arp, H.P.H., Slinde, G.A., Wade, E.J., Bjørseth, K., Breedveld, G.D., Straith, B.F., Moe, K.G.,
- 579 Jartun, M., Høisæter, Å., 2017. Sorbent amendment as a remediation strategy to reduce PFAS mobility and
- leaching in a contaminated sandy soil from a Norwegian firefighting training facility. *Chemosphere*, 171, 918.
- 582 26. Higgins, C.P., Luthy, R.G., 2006. Sorption of Perfluorinated Surfactants on Sediments. *Environ. Sci.*583 *Technol.*, 40(23), 7251-7256.

- 584 27. Jeon, J., Kannan, K., Lim, B.J., An, K.G., Kim, S.D., 2011. Effects of salinity and organic matter on the
 585 partitioning of perfluoroalkyl acid (PFAs) to clay particles. *J. Environ. Monit.*, 13(6), 1803-1810.
- 586 28. Korshin, G.V., Li, C.-W., Benjamin, M.M., 1997. Monitoring the properties of natural organic matter
 587 through UV spectroscopy: A consistent theory. *Water Res.*, 31(7), 1787-1795.
- 588 29. Kothawala, D.N., Köhler, S.J., Östlund, A., Wiberg, K., Ahrens, L., 2017. Influence of dissolved organic
 589 matter concentration and composition on the removal efficiency of perfluoroalkyl substances (PFASs)
 590 during drinking water treatment. *Water Res.*, 121, 320-328.
- 30. Kundu, S., Patel, S., Halder, P., Patel, T., Hedayati Marzbali, M., Pramanik, B.K., Paz-Ferreiro, J., de
 Figueiredo, C.C., Bergmann, D., Surapaneni, A., Megharaj, M., Shah, K., 2021. Removal of PFASs from
 biosolids using a semi-pilot scale pyrolysis reactor and the application of biosolids derived biochar for the
 removal of PFASs from contaminated water. *Environ. Sci.: Water Res. Technol.*, 7(3), 638-649.
- 595 31. Lin, P.-I.D., Cardenas, A., Hauser, R., Gold, D.R., Kleinman, K.P., Hivert, M.-F., Fleisch, A.F., Calafat,
 596 A.M., Webster, T.F., Horton, E.S., Oken, E., 2019. Per- and polyfluoroalkyl substances and blood lipid levels
 597 in pre-diabetic adults—longitudinal analysis of the diabetes prevention program outcomes study. *Environ*.
 598 *Int.*, 129, 343-353.
- 599 32. Liu, C.J., Werner, D., Bellona, C., 2019. Removal of per- and polyfluoroalkyl substances (PFASs) from
 600 contaminated groundwater using granular activated carbon: a pilot-scale study with breakthrough modeling.
 601 *Environ. Sci. Water Res. Technol.*, 5(11), 1844-1853.
- 33. Lu, D., Sha, S., Luo, J., Huang, Z., Zhang Jackie, X., 2020. Treatment train approaches for the remediation
 of per- and polyfluoroalkyl substances (PFAS): A critical review. *J. Hazard. Mater.*, 386, 121963.
- 34. Martin, M.J., Balaguer, M.D., Rigola, M., 1996. Feasibility of Activated Carbon Production from Biological
 Sludge by Chemical Activation with ZnCl2 and H2SO4. *Environ. Technol.*, 17(6), 667-671.
- 35. McCleaf, P., Englund, S., Östlund, A., Lindegren, K., Wiberg, K., Ahrens, L., 2017. Removal efficiency of
 multiple poly- and perfluoroalkyl substances (PFASs) in drinking water using granular activated carbon
 (GAC) and anion exchange (AE) column tests. *Water Res.*, 120, 77-87.
- 36. Mohan, D., Sarswat, A., Ok, Y.S., Pittman, C.U., 2014. Organic and inorganic contaminants removal from
 water with biochar, a renewable, low cost and sustainable adsorbent A critical review. *Bioresour. Technol.*,
 160, 191-202.

- 612 37. Murray, C.C., Vatankhah, H., McDonough, C.A., Nickerson, A., Hedtke, T.T., Cath, T.Y., Higgins, C.P.,
- 613 Bellona, C.L., 2019. Removal of per- and polyfluoroalkyl substances using super-fine powder activated
- 614 carbon and ceramic membrane filtration. *J. Hazard. Mater.*, 366, 160-168.
- 615 38. Muvhiiwa, R., Kuvarega, A., Llana, E.M., Muleja, A., 2019. Study of biochar from pyrolysis and gasification
 616 of wood pellets in a nitrogen plasma reactor for design of biomass processes. *J. Environ. Chem. Eng.*, 7(5),
 617 103391.
- 39. Nguyen, T.M.H., Bräunig, J., Thompson, K., Thompson, J., Kabiri, S., Navarro, D.A., Kookana, R.S.,
 Grimison, C., Barnes, C.M., Higgins, C.P., McLaughlin, M.J., Mueller, J.F., 2020. Influences of Chemical
 Properties, Soil Properties, and Solution pH on Soil–Water Partitioning Coefficients of Per- and
 Polyfluoroalkyl Substances (PFASs). *Environ. Sci. Technol.*, 54(24), 15883-15892.
- 622 40. Oliver, D.P., Li, Y., Orr, R., Nelson, P., Barnes, M., McLaughlin, M.J., Kookana, R.S., 2020. Sorption
 623 behaviour of per- and polyfluoroalkyl substances (PFASs) in tropical soils. *Environ. Pollut.*, 258, 113726.
- 41. Park, M., Wu, S., Lopez, I.J., Chang, J.Y., Karanfil, T., Snyder, S.A., 2020. Adsorption of perfluoroalkyl
 substances (PFAS) in groundwater by granular activated carbons: Roles of hydrophobicity of PFAS and
 carbon characteristics. *Water Res.*, 170, 115364.
- 42. Phong Vo, H.N., Ngo, H.H., Guo, W., Hong Nguyen, T.M., Li, J., Liang, H., Deng, L., Chen, Z., Hang
 Nguyen, T.A., 2020. Poly- and perfluoroalkyl substances in water and wastewater: A comprehensive review
 from sources to remediation. *J. Water Proc. Eng.*, 36, 101393.
- 43. Rotander, A., Kärrman, A., Toms, L.-M.L., Kay, M., Mueller, J.F., Gómez Ramos, M.J., 2015. Novel
 Fluorinated Surfactants Tentatively Identified in Firefighters Using Liquid Chromatography Quadrupole
 Time-of-Flight Tandem Mass Spectrometry and a Case-Control Approach. *Environ. Sci. Technol.*, 49(4),
 2434-2442.
- 44. Sammut, G., Sinagra, E., Sapiano, M., Helmus, R., de Voogt, P., 2019. Perfluoroalkyl substances in the
 Maltese environment (II) sediments, soils and groundwater. *Sci. Total Environ.*, 682, 180-189.
- 45. Sasi, P.C., Alinezhad, A., Yao, B., Kubátová, A., Golovko, S.A., Golovko, M.Y., Xiao, F., 2021. Effect of
- 637 granular activated carbon and other porous materials on thermal decomposition of per- and polyfluoroalkyl
- 638 substances: Mechanisms and implications for water purification. *Water Res.*, 200, 117271.

- 46. Silvani, L., Cornelissen, G., Botnen Smebye, A., Zhang, Y., Okkenhaug, G., Zimmerman, A.R., Thune, G.,
 Sævarsson, H., Hale, S.E., 2019. Can biochar and designer biochar be used to remediate per- and
 polyfluorinated alkyl substances (PFAS) and lead and antimony contaminated soils? *Sci. Total Environ.*,
 694, 133693.
- 47. Singh, B., Macdonald, L.M., Kookana, R.S., van Zwieten, L., Butler, G., Joseph, S., Weatherley, A., Kaudal,
 B.B., Regan, A., Cattle, J., Dijkstra, F., Boersma, M., Kimber, S., Keith, A., Esfandbod, M., 2014.
 Opportunities and constraints for biochar technology in Australian agriculture: looking beyond carbon
 sequestration. *Soil Res.*, 52(8), 739-750.
- 647 48. Smebye, A., Alling, V., Vogt, R.D., Gadmar, T.C., Mulder, J., Cornelissen, G., Hale, S.E., 2016. Biochar
 648 amendment to soil changes dissolved organic matter content and composition. *Chemosphere*, 142, 100-105.
- 49. Sørmo, E., Silvani, L., Bjerkli, N., Hagemann, N., Zimmerman, A.R., Hale, S.E., Hansen, C.B., Hartnik, T.,
- 650 Cornelissen, G., 2021. Stabilization of PFAS-contaminated soil with activated biochar. *Sci. Total Environ.*,
 651 763, 144034.
- 50. Wang, W., Mi, X., Zhou, Z., Zhou, S., Li, C., Hu, X., Qi, D., Deng, S., 2019. Novel insights into the
 competitive adsorption behavior and mechanism of per- and polyfluoroalkyl substances on the anionexchange resin. J. Colloid Interface Sci., 557, 655-663.
- 51. Wei, C., Wang, Q., Song, X., Chen, X., Fan, R., Ding, D., Liu, Y., 2018. Distribution, source identification
 and health risk assessment of PFASs and two PFOS alternatives in groundwater from non-industrial areas. *Ecotox. Environ. Safety*, 152, 141-150.
- 52. Wu, C., Klemes, M.J., Trang, B., Dichtel, W.R., Helbling, D.E., 2020. Exploring the factors that influence
 the adsorption of anionic PFAS on conventional and emerging adsorbents in aquatic matrices. *Water Res.*,
 182, 115950.
- 53. Xiao, F., Jin, B., Golovko, S.A., Golovko, M.Y., Xing, B., 2019. Sorption and Desorption Mechanisms of
 Cationic and Zwitterionic Per- and Polyfluoroalkyl Substances in Natural Soils: Thermodynamics and
 Hysteresis. *Environ. Sci. Technol.*, 53(20), 11818-11827.
- 54. Xiao, X., Ulrich, B.A., Chen, B., Higgins, C.P., 2017. Sorption of Poly- and Perfluoroalkyl Substances
 (PFASs) Relevant to Aqueous Film-Forming Foam (AFFF)-Impacted Groundwater by Biochars and
 Activated Carbon. *Environ. Sci. Technol.*, 51(11), 6342-6351.

- 55. Xu, B., Liu, S., Zhou, J.L., Zheng, C., Weifeng, J., Chen, B., Zhang, T., Qiu, W., 2021. PFAS and their
 substitutes in groundwater: Occurrence, transformation and remediation. *J. Hazard. Mater.*, 412, 125159.
- 56. Yong, Z.Y., Kim, K.Y., Oh, J.-E., 2021. The occurrence and distributions of per- and polyfluoroalkyl
 substances (PFAS) in groundwater after a PFAS leakage incident in 2018. *Environ. Pollut.*, 268, 115395.
- 671 57. Yu, J., Lv, L., Lan, P., Zhang, S., Pan, B., Zhang, W., 2012. Effect of effluent organic matter on the
- adsorption of perfluorinated compounds onto activated carbon. J. Hazard. Mater., 225-226, 99-106.
- 58. Zhang, D., He, Q., Wang, M., Zhang, W., Liang, Y., 2019a. Sorption of perfluoroalkylated substances
 (PFASs) onto granular activated carbon and biochar. *Environ. Technol.*, 1-12.
- 59. Zhang, D.Q., Zhang, W.L., Liang, Y.N., 2019b. Adsorption of perfluoroalkyl and polyfluoroalkyl substances

676 (PFASs) from aqueous solution - A review. *Sci. Total Environ.*, 694, 133606.

- 677 60. Zhang, W., Zhang, D., Liang, Y., 2019c. Nanotechnology in remediation of water contaminated by poly-
- and perfluoroalkyl substances: A review. *Environ. Pollut.*, 247, 266-276.

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680