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1 **Characterization and flocculation performance of a newly green flocculant derived**  
2 **from natural bagasse cellulose**

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31 **Abstract**

32 A newly green natural polymer bagasse cellulose based flocculant (PBCF) was  
33 synthesized utilizing a grafting copolymerization method for effectively enhancing humic  
34 acid (HA) removal from natural water. This work aims to investigate flocculation  
35 behavior of PBCF in synthetic water containing HA, and the effects of flocculant dose  
36 and initial solution pH on flocculation performance. Results showed that PBCF  
37 functioned well at a flocculant dose of 60 mg/L and pH ranging from 6.0 to 9.0. The  
38 organic removal efficiency in synthetic water in terms of HA ( $UV_{254}$ ) and chemical  
39 oxygen demand ( $COD_{Mn}$ ) were up to 90.6% and 91.3%, respectively. Furthermore, the  
40 charge neutralization and adsorption bridging played important roles in HA removal.  
41 When applied for lake water, PBCF removed 91.6% turbidity and 50.0% dissolved  
42 organic matter, respectively. In short, PBCF demonstrates great potential in water  
43 treatment in a safe and environmentally friendly or 'green' way.

44 **Keywords:** Bagasse cellulose; Cationic flocculant; Flocculation behavior; Humic acid

## 45 1. Introduction

46 The world's supply of surface water has been severely polluted over many  
47 generations due to industrial development, rapid urbanization, rising levels of  
48 contamination, and abuse of chemical fertilizers and pesticides. Eutrophication of water  
49 bodies has worsened, and the species and quantities of organic matter in water continue  
50 to grow, leading to excessive proliferation of algae. These have ensured that the natural  
51 organic matter (NOM) content in water exceeds government standards/regulations  
52 required for drinking water sources and quality. It is estimated humus accounts for 50%-  
53 90% of natural organic matter (NOM) in water, and humic acid (HA) is an essential  
54 component of humus and dissolved organic matter (DOM) in water (Malczewska et al.,  
55 2022). HA in natural water bodies will increase the chromaticity and smell of water,  
56 and it can even form more toxic complexation with organic matter and heavy metals  
57 (Libeck et al., 2020). Moreover, HA plays a protective role for colloid particles in water  
58 by enhancing their stability, leading to challenges on how to treat water. HA can react  
59 with chlorine during chlorination to produce toxic precursors of disinfection by-  
60 products, for instance trichloromethanes which are carcinogenic and extremely harmful  
61 to human health (Lou et al., 2021).

62 For this reason, it is of great importance to seek effective methods to remove HA  
63 from water. To date, membrane filtration, adsorption, oxidation, biotechnology,  
64 coagulation/flocculation methods, etc., have been the subject of much attention (Du et  
65 al., 2021; Jang et al., 2021; Ryu et al., 2021; Yang et al., 2021; Zhang et al., 2021). The  
66 process of coagulation/flocculation is widely used owing to its advantages of low cost,

67 low energy consumption, easy operation, effective solid-liquid separation of colloidal  
68 suspension and high efficiency in removing natural organic matter from water (Wang  
69 et al., 2019).

70 The coagulants/flocculants for HA removal mainly include inorganic coagulants,  
71 organic polymer flocculants, and natural polymer flocculants. Of these, inorganic  
72 coagulants are widely used because they are inexpensive, yet excessive use of inorganic  
73 coagulants can introduce a lot of metal ions to water and in this way cause secondary  
74 pollution (Vereb et al., 2019). In addition, organic synthetic polymer flocculants can  
75 form stronger and denser flocs at small doses, thus increasing the rate of floc settling  
76 and obtaining better removal efficiency, whereas acrylamide is harmful to the human  
77 body (Li et al., 2017). In contrast, natural polymer flocculants are widely respected due  
78 to their environmental compatibility and renewable properties. Based on different raw  
79 materials, they can be roughly divided into five categories: chitosan flocculants, starch  
80 flocculants, lignin flocculants, plant gum flocculants and cellulose flocculants (Li et al.,  
81 2018; Chen et al., 2020b; Liu et al., 2020; Wang et al., 2021; Wu et al., 2021).

82 Of these, cellulose flocculants have been widely researched because their raw  
83 materials are abundant in nature. Bagasse, orange peel and pomelo peel are high in  
84 cellulose content. A large amount of bagasse wastes every year is produced by the sugar  
85 industry since sugarcane with a short growing cycle is the main cash crop grown in  
86 south China. However, bagasse is rarely reused as a resource (Ma and Wen, 2020).  
87 Therefore, the extraction of cellulose from bagasse and the utilization of cellulose can  
88 reduce the waste of resources and curb environmental pollution. As well, cellulose has

89 the advantages of porous structure and large specific surface area. The surface has a  
90 large number of hydrophilic hydroxyl groups, which can provide more active sites for  
91 modification. The required flocculants can be prepared by modifying cellulose and  
92 introducing functional groups (Li et al., 2018). A new flocculant BPC-g-PAM was  
93 prepared by grafting polyacrylamide (PAM) onto bamboo pulp cellulose (BPC) in  
94 homogeneous aqueous solution (Liu et al., 2014). It can remove as much as 98% of  
95 kaolin with a negative surface charge in water under acidic and neutral conditions, due  
96 to the enhanced charge neutralization and bridging effect in the flocculation process. In  
97 addition to good removal efficiency, it is in fact friendly to the environment. A natural  
98 cellulose dicarboxylic acid flocculant (DCCs) was synthesized from dialdehyde  
99 cellulose via the one-step method. It can improve the removal efficiency of kaolin and  
100 papermaking wastewaters by improving neutralization in the flocculation process (Zhu  
101 et al., 2015). By preparing a flocculant (CMCND) with strong cationic properties and  
102 unique hydrophobic properties, CMCND demonstrated superior efficiency in removing  
103 trace nonylphenol (NP), turbidity and humus from synthetic surface water (Yang et al.,  
104 2016).

105 As is well known, the surface of HA in natural water is negatively charged and can  
106 be removed by electro-neutralization. Therefore, by introducing cationic groups on the  
107 surface of cellulose, the electrostatic repulsion and adsorption ability is enhanced.  
108 Interestingly, inspired by previous research, acryloyloxyethyl trimethyl ammonium  
109 chloride (DAC) is a commonly used graft monomer for quaternary amination of natural  
110 polymer flocculant (Liu et al., 2019; Jiang et al., 2020). DAC can be introduced to the

111 bagasse cellulose surface by graft copolymerization to improve the removal of HA.

112 In this study, a new green flocculant namely PBCF was derived from sugarcane  
113 bagasse with potassium persulfate serving as initiator and DAC as grafting monomer.  
114 The characteristics of PBCF were analyzed, as well as the performance and mechanism  
115 of the flocculant on HA in water. This helps to explain the possibilities for improving  
116 the pretreatment process of micro-polluted water resources.

## 117 **2. Materials and methods**

### 118 **2.1 Reagents**

119 Bagasse was collected from the farmers' markets. Humic acid (HA) was provided  
120 by Sigma-Aldrich Corporation. DAC (80% aqueous solution) was purchased from  
121 Shanghai D&B Biological Science and Technology Co., Ltd. Meanwhile, sodium  
122 hydroxide (NaOH, AR) and potassium persulfate ( $K_2S_2O_8$ , AR) were purchased from  
123 Shanghai Macklin Biochemical Co., Ltd.

### 124 **2.2. Preparation of PBCF**

#### 125 2.2.1 Preparation of alkaline bagasse cellulose

126 First, the bagasse purchased from the market was cleaned, soaked, and washed to  
127 remove all excess impurities. Next, the bagasse was dried until a constant weight was  
128 achieved in a 55 °C oven. Then, the bagasse was pulverized into 40 mesh powder for  
129 use. Following this, bagasse powder weighing 20 g was put it into a beaker, 100 mL  
130 20% NaOH solution was added to it, and then it was soaked at room temperature for 24  
131 h. After that, the sample was filtered and washed with distilled water until it was slightly  
132 alkaline ( $pH \approx 8.0$ ), dried to a constant weight, and ground into powder to obtain alkaline

133 bagasse cellulose (Abc).

### 134 2.2.2 Preparation of PBCF

135 As displayed in Fig. S1, 1 g of ( $W_0$ ) Abc and a specific amount of distilled water  
136 were added to a four-necked flask equipped with a gas guide and a stirring device,  
137 heated and stirred at 90 °C for one hour to allow sufficient wetting. When the  
138 temperature was reduced to the reaction temperature of 45 °C, a certain amount of  
139 potassium persulfate (KPS) initiator was added. After it was fully dissolved, a specific  
140 mass ratio of DAC was added, and nitrogen was passed to ensure the reaction could  
141 proceed under nitrogen protection; the reaction was stopped after 3 h at 45 °C. After the  
142 reaction finished, the product was precipitated with anhydrous ethanol, washed with  
143 anhydrous ethanol and distilled water, respectively, and dried in an oven until constant  
144 weight ( $W_2$ ) was achieved. Each sample was prepared three times in duplicate. The  
145 synthesis line of the PBCF is shown in Fig. S2.

### 146 2.2.3 Determination of grafting percentage of PBCF

147 The grafting percentage ( $G$ ) is the percentage of the mass of DAC monomer that  
148 has been grafted onto Abc to the mass of alkaline bagasse cellulose monomer. It  
149 determines the level of reaction according to the effective number of monomers in the  
150 grafting reaction (Chen et al., 2020c). It can be calculated using the following equation:

$$151 \quad G = [(W_2 - W_0) / W_0] \times 100\%$$

152 Where  $W_0$  denotes the initial weight of Abc, and  $W_2$  stands for the weight of PBCF  
153 after the reaction.

### 154 **2.3 Raw water and coagulation experiment**

155 200 mg HA and 0.4 g sodium hydroxide were dissolved in deionized water and  
156 then stirred. After the HA was dissolved, the solution was transferred to a 1.0 L  
157 volumetric flask with a constant volume. The reserve solution was stored in a brown  
158 reagent bottle, while the HA solution was diluted to 5 mg/L, 10 mg/L and 15 mg/L with  
159 deionized water when used, and the pH of the solution was adjusted near to neutral. The  
160 lake water sample was taken from the central lake at Tianjin Chengjian University  
161 (Xiqing District, Tianjin). The pH, turbidity, DOM (UV<sub>254</sub>), and COD of the lake water  
162 are 8.0 - 9.0, 31.2 - 33.4 NTU, 6.6 - 6.9 mg/L, and 12.1 - 12.8 mg/L, respectively.

163 The flocculation process of PBCF was as follows: adding flocculant, stirring at a  
164 speed of 200 r/min for 1 min, stirring at a speed of 100 r/min for 10 min, stirring at a  
165 speed of 30 r/min for 20 min, left to precipitate for 40 min, taking supernatant and  
166 finally, UV<sub>254</sub> (DOM concentration) and COD<sub>Mn</sub> were measured. Three parallel  
167 samples were set up for each set of experiments.

### 168 **2.4 Characterization of PBCF**

169 The functional groups and surface composition of PBCF were characterized by  
170 Fourier transform infrared (FT-IR) (Nicolet iS10, Thermo Fisher Technology Co., Ltd.)  
171 and X-ray photoelectron spectroscopy (XPS) ((K-alpha, Thermo Scientific). The  
172 surface morphology of PBCF was analyzed using a scanning electron microscope  
173 (SEM) (JSM-7800F, Japan Electronics Co., Ltd.). The crystal structure of PBCF was  
174 investigated by X-ray diffractometer (XRD) (D8 ADVANCE, Bruker, Germany). The  
175 zeta potential of PBCF at different pH values was measured using a zeta potential

176 analyzer (Nano-ZS, Malvern Company, UK).

## 177 **2.5 Flocculation performance analysis**

178 Organic concentration of water samples before and after coagulation was  
179 measured in terms of  $UV_{254}$  and COD using UV-visible spectrophotometer (UV-2600,  
180 Shimadzu, Japan) and COD<sub>Mn</sub> analyzer (Q-COD<sub>MN</sub>, Qing Shi Jie, Shanghai),  
181 respectively. This served to determine the flocculant efficiency in removing HA.  
182 Furthermore, Zeta potential analyzer (Nano-ZS, Malvern Company, UK) measured the  
183 change in the zeta potential of water bodies before and after flocculation, while Optical  
184 flocculation tester (IPDA-100, ECONOVEL, South Korea) estimated the change in  
185 extent of flocculation during the flocculation process. On the basis of these analyses,  
186 the flocculating mechanism of the PBCF in the flocculation process was explored.

## 187 **3. Results and discussion**

### 188 **3.1 Optimization of the flocculant preparation process**

189 The PBCF cationic cellulose-based grafting flocculant was optimized by changing  
190 the KPS dosage and mass ratio of Abc to DAC, with the removal effect of 5 mg/L HA  
191 solution and the grafting rate of flocculant as evaluation indices.

#### 192 **3.1.1 Effect of KPS dosage**

193 As shown in Fig. S3a, when the KPS dosage increased the removal efficiencies of  
194 HA by PBCF also increased firstly but then diminished. When the dosage of KPS was  
195 1.8 g, the removal efficiency of HA reached a maximum of  $90.5 \pm 0.41\%$ . As the KPS  
196 dosage rose further, the removal efficiency of HA gradually declined. This was because  
197 KPS can stimulate hydroxyl groups on basic cellulose and enable them to provide more

198 active sites for graft copolymerization. However, when the dosage of KPS was too high  
199 and exceeded 1.8 g, the reaction rate occurred too rapidly, resulting in the implosion  
200 reaction between Abc and DAC (Chen et al., 2021). That caused an increase in the  
201 monomer mass and graft rate after the reaction; nevertheless, it was not conducive to  
202 the growth of the chain and generation of graft polymer (Chen et al., 2020c).  
203 Subsequently, this led to waning HA removal efficiency.

204 Meanwhile, the DAC copolymer adhered to the Abc surface and covered the pores  
205 and tentacle-like structures on Abc. It reduced the bridging adsorption of PBCF in the  
206 flocculation process. This was the main reason why the grafting rate of PBCF increased  
207 while the  $UV_{254}$  removal rate fell when the amount of KPS was more than 1.8 g.

### 208 3.1.2 Effect of m (Abc): m (DAC)

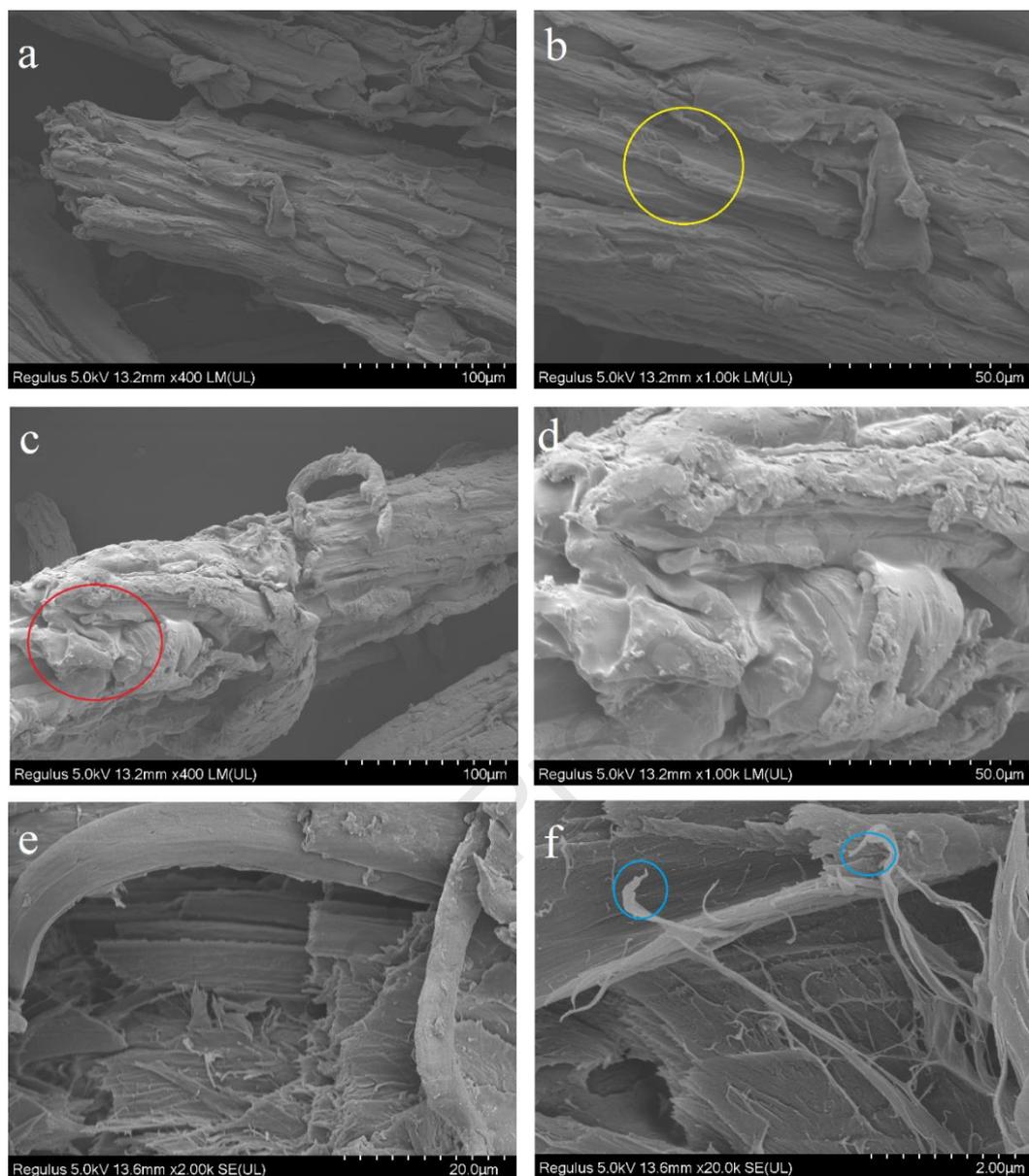
209 The influence of the mass ratio of Abc on DAC regarding HA removal and graft  
210 rate is depicted in Fig. S3b. It can be seen that as the ratio of m (Abc) to m (DAC)  
211 increased gradually, the rate of removing 5 mg/L HA by PBCF flocculant increased first  
212 but then decreased. When m (Abc): m (DAC) was 1:3, the maximum HA removal rate  
213 can reach up to  $90.5 \pm 0.41\%$ . With the increase in the DAC ratio, the positive charge on  
214 the polymer chain rose and thus the electric neutralization effect of flocculant was  
215 enhanced, which helped flocculation sedimentation. However, when the proportion of  
216 DAC in the reaction system was too large, it caused too much of a positive charge on  
217 the polymer chain. As a result, the remaining positive charge reversed the properties of  
218 the particle surface charge and augmented the repulsive force between the particles.  
219 This led to the repulsive force between the flocculant and the colloidal particles and

220 reduction of the flocculation effect. Moreover, excessive DAC gave rise to self-  
221 polymerization, and increased the grafting of PBCF first and then decreased slowly.

## 222 **3.2 Characterization of PBCF**

### 223 3.2.1 Scanning electron microscope

224 The SEM images of Abc, PBCF showed obvious structural changes (Fig 1). After  
225 soaking in NaOH solution, the surface of alkaline bagasse cellulose was noticeably  
226 etched (Fig. 1a). According to the yellow area shown in Fig. 1b, the surface of Abc is  
227 coarse with a large number of wrinkles and pores. This unique structure provided  
228 suitable conditions for the graft copolymerization reaction (Fig. 1a and b). However,  
229 the product surface (PBCF) looked smoother, and many protrusions on the surface of  
230 PBCF can be observed (Fig. 1c and d). Observations suggested that this surface showed  
231 abundant lamellar and fiber-like structures, which could undoubtedly increase the  
232 surface area of the product. In this way, it would be more conducive to boosting the  
233 adsorption and bridging effect and enhancing removal of colloidal particles in the  
234 flocculation (Fig. 1e and f).



235

236

**Fig. 1.** SEM micrographs of Abc (a, b) and PBCF (c, d, e, f)

237

### 3.2.2 Fourier transform infrared

238

239

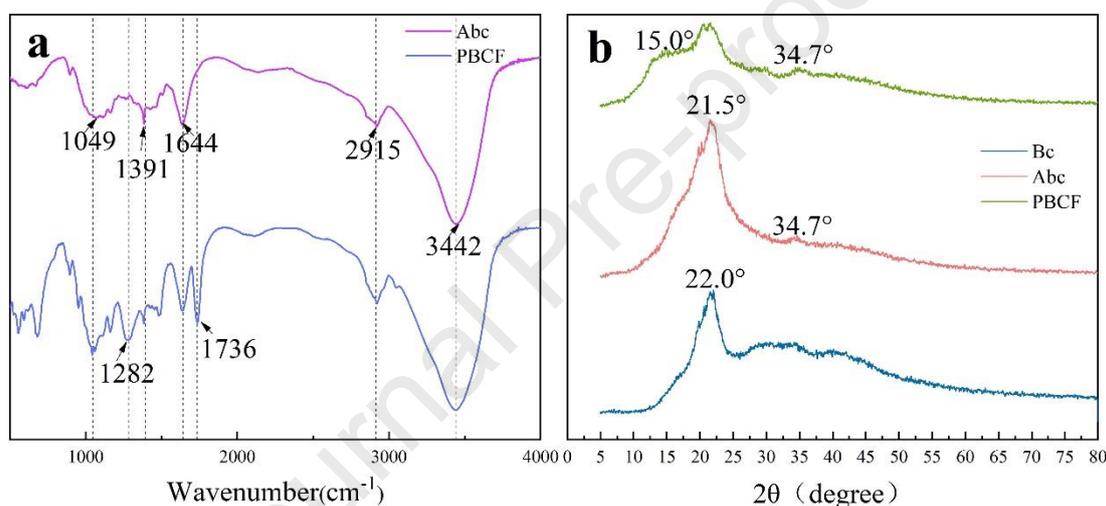
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242

Fig. 2a indicated that the absorptive peaks of Abc at  $3442\text{ cm}^{-1}$ ,  $2915\text{ cm}^{-1}$ ,  $1391\text{ cm}^{-1}$  and  $1049\text{ cm}^{-1}$  could be assignable to O-H stretching vibration, C-H symmetric stretching vibration, C-H bending vibration, and -CH stretching vibration on cellulose skeleton, respectively (Jiang et al., 2020; Tang et al., 2020). The band at  $1644\text{ cm}^{-1}$  was attributed to OH bending vibration in absorbed water. Apart from the original

243 characteristic peaks of Abc, some new absorption peaks at  $1282\text{ cm}^{-1}$  and  $1736\text{ cm}^{-1}$   
 244 emerged in PBCF, which belonged to the stretching vibration of C-N, and COOR,  
 245 respectively (Wang et al., 2019). The absorption peaks at  $1049\text{ cm}^{-1}$  were significantly  
 246 enhanced. This is caused by the reaction of DAC with the -OH on the cellulose skeleton,  
 247 resulting in a stretching vibration of -CH (Liu et al., 2019; Tang et al., 2021). Generally,  
 248 structural changes confirmed that the DAC monomer was successfully grafted onto the  
 249 Abc molecular chain.



250

251 **Fig. 2.** (a) FTIR of Abc and PBCF; (b), XRD of Bc, Abc, and PBCF

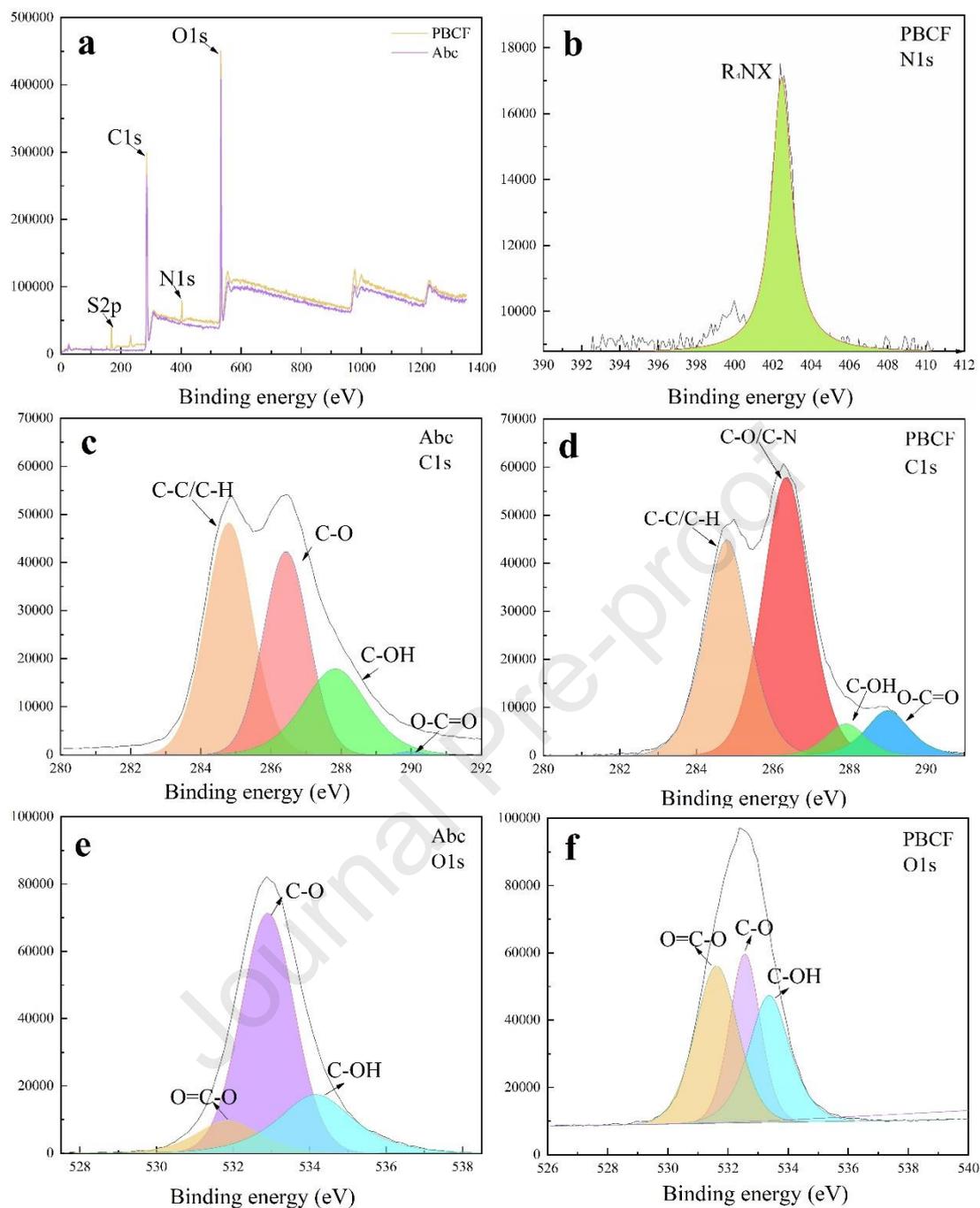
### 252 3.2.3 X-ray diffraction

253 The crystal structures of bagasse cellulose, alkaline bagasse cellulose, and PBCF  
 254 were characterized by XRD. The XRD spectra of Bc, Abc, and PBCF are shown in Fig.  
 255 2b. A wide peak appeared at  $2\theta = 22^\circ$ , which belonged to the crystallization peak of  
 256 natural cellulose I (Badawy et al., 2021). The XRD spectrum of Abc showed that the  
 257 characteristic peak of Bc shifted to  $21^\circ$  from  $22^\circ$  and a small diffraction peak appeared  
 258 at  $2\theta = 34.7^\circ$ , which matched well with the crystal structure of cellulose I  $\beta$  (Tewatia  
 259 et al., 2021). The diffraction peak intensity of Abc at  $2\theta = 21.5^\circ$  increased, and the

260 crystallinity reduced from 86.89% to 65.74%, meaning that the glycosidic bonds and  
261 molecular chains of network structure in Bc after NaOH treatment gradually broke up  
262 (Rizwan et al., 2021). Due to the hydrolysis of the amorphous regions of cellulose, the  
263 hydrogen bonds among molecular chains of cellulose were severely broken.  
264 Amorphous regions and disordered crystals were further destroyed, whereas the regular  
265 crystals did not change, thus crystallinity of Abc decreased. Based on this, PBCF  
266 exhibited a small diffraction peak at  $15^\circ$  and the crystallinity was 75.1%. This suggested  
267 that DAC was successfully grafted onto Abc, which greatly reduced the hydrogen  
268 bonding of cellulose and changed the original structure.

#### 269 3.2.4 X-ray photoelectron spectroscopy

270 XPS analysis further confirmed that the DAC monomer was successfully grafted  
271 to the alkaline bagasse cellulose monomer. The XPS full scan spectra of Abc and PBCF  
272 are shown in Fig. 3. The PBCF contained C, O, N, and S, while Abc contained only C  
273 and O and less N. The peak of PBCF at 402.5 eV is the characteristic peak of N1s  
274 quaternary ammonium salt derived from DAC (Liu et al., 2017). The main elements C  
275 1s and O 1s of Abc and PBCF were analyzed. Through the spectra of C 1s and O 1s, it  
276 emerged that C-O, C-OH and C-H were transformed into O-C=O and C-N through a  
277 graft copolymerization reaction, resulting in more O-C=O and C-N. It strongly proved  
278 the occurrence of graft copolymerization (Liu et al., 2019).



279

280

**Fig. 3.** XPS of Abc (a, c, e) and PBCF (a, b, d, f)

### 281 3.2.5 Zeta potential

282 The zeta potentials of BPCF at different pH values are exhibited in Fig. S4. As  
 283 shown in Fig. S4, the PBCF exhibited a positive charge at pH ranging from 3 to 10. In  
 284 addition, the zeta potential of PBCF presented a decrease trend, which was resulted  
 285 from the restricted ionization of the quaternary amine group on the PBCF molecular

286 chain under alkaline conditions (Kuncoro et al., 2018a; Kuncoro et al., 2018b). This  
287 allowed the PBCF to provide good removal of HA in a wide pH range.

### 288 **3.3 Flocculation of synthetic water**

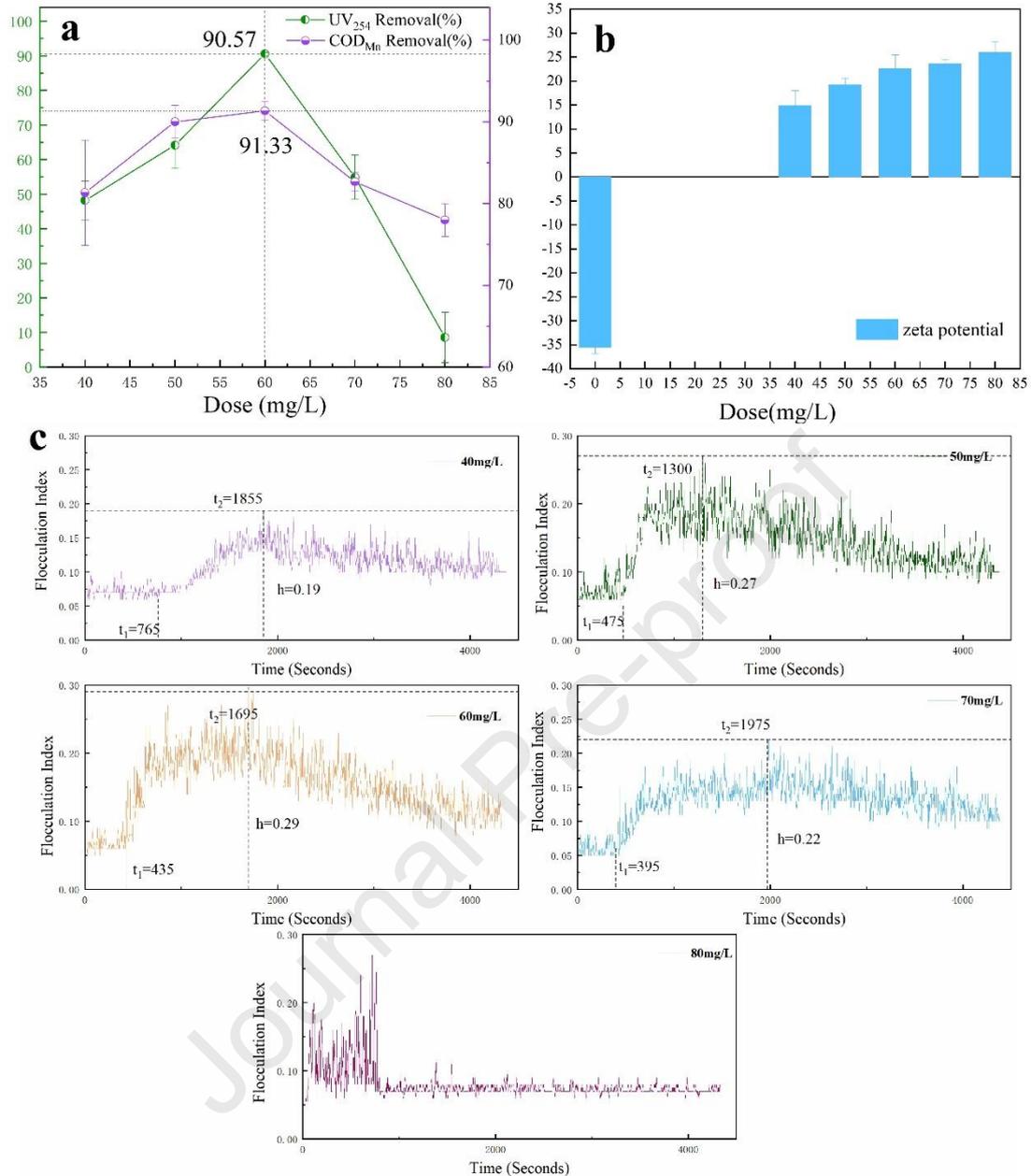
#### 289 3.3.1 Effect of flocculant dose

290 The flocculant dose plays a decisive role in water treatment, and the optimized  
291 dose can ensure the rational utility of resources. In this study, flocculation performance  
292 was investigated in the simulated synthetic water with pH of 7 at 25 °C. As displayed  
293 in Fig. 4a, removal rates of organic matter based UV<sub>254</sub> and COD<sub>Mn</sub> rapidly increased  
294 and then decreased. When the dose of PBCF was up to 60.0 mg/L, removal rates of  
295 UV<sub>254</sub> and COD<sub>Mn</sub> reached the maxima of 90.57±0.41% and 91.33±1.15%, respectively.  
296 However, when the PBCF dose continued increasing, the removal rate abated gradually.  
297 This phenomenon can be attributed to the adsorption charge neutralization in the  
298 flocculation process (Ma et al., 2021).

299 Changes in the zeta potential and Flocculation Index (FI) values in the flocculation  
300 process are depicted in Fig. 4b and c. The flocculation appearance time  $t_1$  gradually  
301 decreased when the PBCF dose increased (Fig. 4c). When the dose of PBCF was 80  
302 mg/L,  $t_1$  was minimal at 720 s. However, due to the overdosing of flocculant, repulsion  
303 between flocs occurred, causing the flocs decomposed quickly. In addition, the value  
304 of FI would gradually increase when the flocs were aggregated, and therefore the higher  
305 value of FI caused the larger floc particle size. The floc particle size reached its  
306 maximum value at the PBCF dosage of 60 mg/L.  $t_2$  and the corresponding time required  
307 were smallest at the flocculant dose of 50 mg/L. This proved that the coagulation rate

308 was at its largest when the flocculant dose was 50 mg/L. When the flocculant dose was  
309 greater than 60 mg/L, the zeta potential gradually tended to stabilize.

310 It proved that when the dose of PBCF was less than 50 mg/L, electrical  
311 neutralization tended to dominate the flocculation process. At this time, the floc size  
312 was small and the removal effect was poor. When the flocculant dose increased  
313 continuously, the adsorption bridging capacity was gradually enhanced and took the  
314 leading role. When the dose of PBCF was beyond 70 mg/L, the flocculant proved to be  
315 excessive, leading to a repulsive interaction among the flocculant molecules. Thus, the  
316 adsorption bridging interaction gradually weakened, gradually shrinking the floc size  
317 and the removal performance as well. It can be seen that the adsorption bridging effect  
318 of PBCF played a dominant role in removals of  $UV_{254}$  and  $COD_{Mn}$ . The size of the  
319 formed floc was largest when PBCF dose was 60 mg/L, under which the as-prepared  
320 PBCF exhibited the best HA removal performance.



321

322 **Fig. 4.** Effects of dose on removal efficiencies (a) of UV<sub>254</sub>, COD<sub>Mn</sub>, zeta potential323 (b), and Flocculation Index (c) (h and t<sub>2</sub> represent the final size of flocs and the time

324 when the floc size reaches the maximum value, respectively)

325 3.3.2 Effects of pH

326 The effect of synthetic water pH at 4.0 – 10.0 was examined at a dose of 60.0 mg/L

327 of PBCF, as shown in Fig. 5. As can be seen from Fig. 5a, the removal efficiencies of

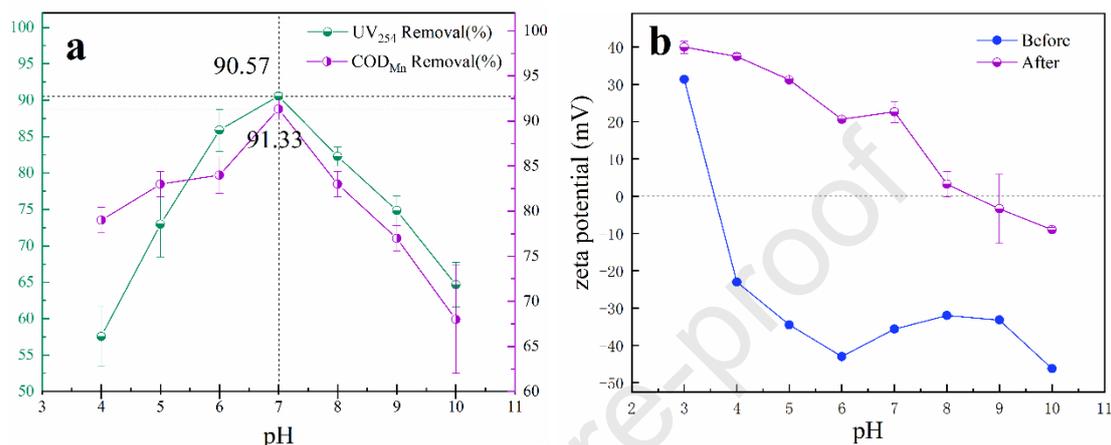
328 UV<sub>254</sub> and COD<sub>Mn</sub> were improved as pH increased from 4.0 to 7.0. The best UV<sub>254</sub> and

329 COD<sub>Mn</sub> removals reached the maximum value of 90.57±0.41% and 91.33±1.15%,  
330 respectively, at pH of 7.0. When the pH level was more than 7.0, the removal rate  
331 decreased slightly with an increase in the pH. This was mainly due to the protonation  
332 and deprotonation of HA at different pH (Liu et al., 2020). At a low pH, it was difficult  
333 for protons to depart from HA, which made HA more stable in water. Moreover, the  
334 surface of PBCF was positively charged. Since the electrical neutralization effect  
335 weakened during the flocculation process, it was hard to remove HA under acidic  
336 conditions. In addition to HA protonation, functional groups belonging to the  
337 carboxylic or phenolic acids of the HA group would be more likely to deprotonate at  
338 higher pH, which resulted in more charges on the HA surface. This could be explained  
339 by the fact of poor removal efficiency of PBCF for UV<sub>254</sub> and COD<sub>Mn</sub> in the acid range.

340 Fig. 5b shows the changes occurring in zeta potential before and after flocculation.  
341 Due to the protonation of HA and the interaction of PBCF, the colloidal particles in  
342 water experienced a charge reversal behavior. The colloidal suspension appeared to be  
343 stable when the pH changed from 4.0 to 6.0. When pH was about 8.0, the zeta potential  
344 of the supernatant after flocculation was close to 0 mV, and the removal efficiency  
345 diminished when the pH increased. It is evidence that charge neutralization and  
346 adsorption bridging both existed in the flocculation. With the increase of pH, more  
347 negative charges of HA would be electrically neutralized with PBCF, and the degree of  
348 deprotonation of HA would be reinforced. Furthermore, the number of flocculants used  
349 for adsorption bridge would be reduced.

350

351 The small flocs generated by adsorption charge neutralization cannot form large  
 352 flocs by effective collision. This also led to the poor removal of dissolved organic matter.  
 353 Fine flocs cannot easily settle but they can be filtered through a filter membrane which  
 354 is 0.45  $\mu\text{m}$  in size.



355

356 **Fig. 5.** Effects of pH on (a) removal efficiencies of UV<sub>254</sub>, COD<sub>Mn</sub> (b) zeta potential  
 357 of water before and after flocculation.

### 358 3.3.3 Effects of initial HA concentrations

359 As shown in Fig. S5a and b, when the HA concentration was 10 mg/L, the  
 360 maximum removal rates of UV<sub>254</sub> and COD<sub>Mn</sub> were 94.03±0.65% and 88.00±1.41%,  
 361 respectively, at the dose of PBCF of 80 mg/L. With the increase of PBCF dose, the  
 362 removal rate gradually improved. When the HA concentration was 15 mg/L, the UV<sub>254</sub>  
 363 removal rate reached the highest (94.11±0.40%) at the dose of PBCF of 80 mg/L. When  
 364 the dosage was more than 80 mg/L, the UV<sub>254</sub> removal rate gradually stabilized to more  
 365 than 92.52%. At the PBCF dose of 70 mg/L, the maximum removal rate of COD<sub>Mn</sub>  
 366 amounted to 90.67±0.47%. With the gradual increase of the PBCF dose, the removal  
 367 rate fell slightly.

368 Combined with the changes in zeta potential of the PBCF before and after  
369 flocculation (Fig. S5b), it can be seen that as the zeta potential of the supernatant  
370 gradually approached the isoelectric point, the efficiency of the flocculant in removing  
371 HA gradually increased, presenting a positive correlation trend. A slight decrease was  
372 observed when the zeta potential became positive.

373 When the HA solution was 10 mg/L, and the dose of flocculant was 80 mg/L, the  
374 adsorption bridging ability of the flocculant was not fully utilized. Therefore, with the  
375 increase in the flocculant mass, the simulated water exhibited strong positive electricity.  
376 The flocculant repelled and this in turn reduced the removal efficiency. However, when  
377 the concentration of HA increased to 15 mg/L, the adsorption bridging capacity was  
378 fully utilized, and the simulated water presented weak positive electricity. Consequently,  
379 the removal efficiency tended to be stable.

380 Under the same conditions, the flocculant's effectiveness in removing large  
381 concentrations of HA was not significantly reduced. This is mainly due to a dominant  
382 role of PBCF in adsorption bridging throughout the HA removal process. With the  
383 increase of HA concentration, the collision probability of flocculant and HA in solution  
384 was greatly strengthened. The number of colloidal particles produced after flocculation  
385 rose. In the flocculation stage the colloid particles constantly collided, which elevated  
386 the volume of flocs. With the rising number of alum blooms, the settling speed became  
387 faster and the removal efficiency improved.

388

### 389 3.4 Flocculation of lake water

#### 390 3.4.1 Dose optimization

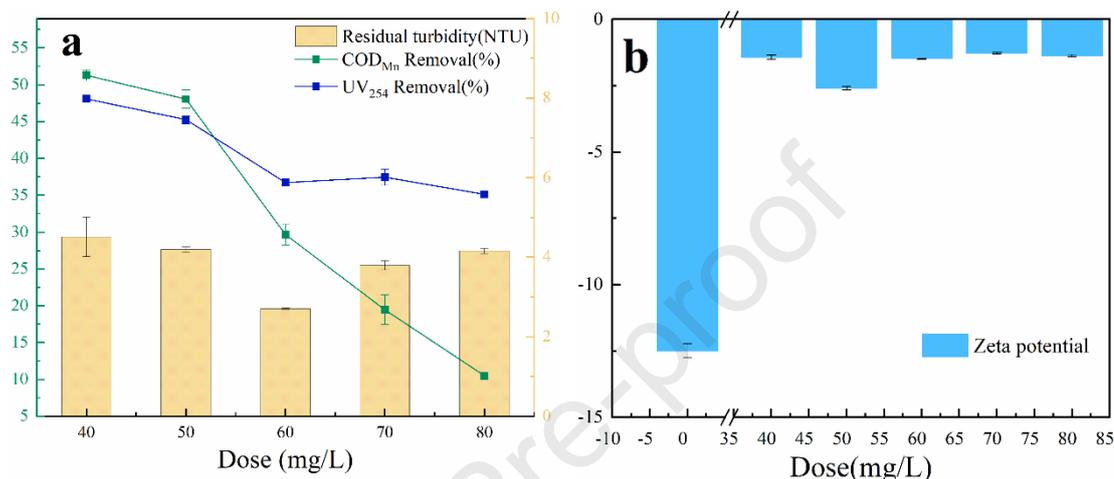
391 Taking turbidity,  $UV_{254}$ , and  $COD_{Mn}$  removal rates as indices, the influence of  
392 PBCF dosage on coagulation performance in natural lake water was investigated. The  
393 flocculant dosage of 60 mg/L achieved the best removal efficiency for turbidity with  
394 residual turbidity of only  $2.79 \pm 0.03$  NTU and a removal rate of  $91.61 \pm 0.09\%$ . When  
395 the dose of PBCF was 40 mg/L, the efficiencies in removing  $UV_{254}$  and  $COD_{Mn}$  reached  
396 the maximum of  $48.08 \pm 0.34\%$  and  $51.27 \pm 0.71\%$ , respectively. However, both amounts  
397 were smaller than the best removal efficiency in synthetic water (Fig. 6a and b).

398 This phenomenon indicated that the turbidity removal efficiency by PBCF is better  
399 than that of  $UV_{254}$  and  $COD_{Mn}$ . This is mainly because insoluble colloidal particles in  
400 water were more easily removed by coagulation compared with DOM. Additionally, it  
401 was gradually enhanced with the increase of flocculant dose but nevertheless, the  
402 repulsive force appeared when too much flocculant was added to the water sample. This  
403 led to the gradual decline in  $UV_{254}$  and  $COD_{Mn}$  removal. No other pollutants were  
404 added to the synthetic water, allowing the flocculant to target HA for removal so better  
405 removal efficiencies of  $UV_{254}$  and  $COD_{Mn}$  were obtained. It is important to note that  
406 the insoluble colloidal particles in water would combine with these particles formed by  
407 flocculation, triggering an increase in colloidal particles weight and more conducive  
408 conditions for the removal of the remaining turbidity in water.

409 PBCF can effectively remove turbidity,  $UV_{254}$  and  $COD_{Mn}$  from lake water.

410 Sugarcane bagasse is cheap and readily available compared to other biomass, which

411 considers a renewable resource while the PBCF preparation process does not cause  
 412 secondary pollution to the environment. Moreover, the sludge floc formed by  
 413 flocculation is small in size, which facilitates its application in practical  
 414 engineering(Chen et al., 2020a).



415

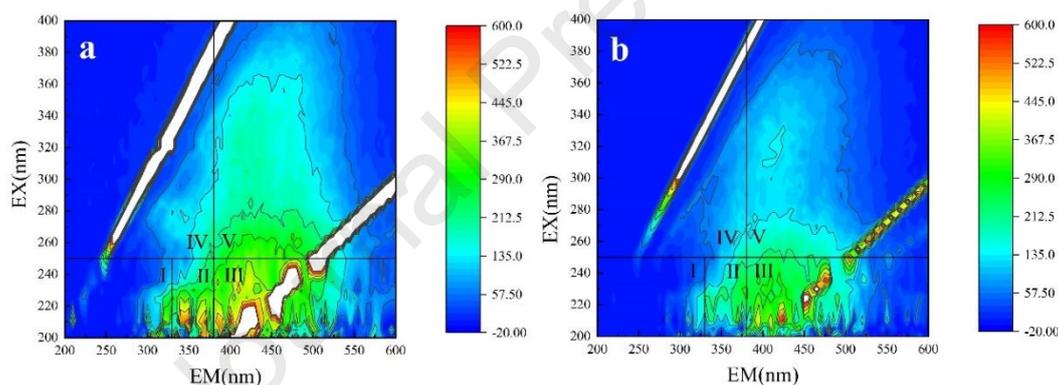
416 **Fig. 6.** Effects of dose of (a) PBCF on removal efficiencies of UV<sub>254</sub>, COD<sub>Mn</sub>,  
 417 and residual turbidity (b) zeta potential.

#### 418 3.4.2 Three-dimensional fluorescence analysis.

419 Fluorescence spectrophotometer (LS55, USA PE) was implemented to conduct  
 420 three-dimensional fluorescence analysis on water samples before and after treatment.  
 421 The PBCF's effectiveness was further explored for treating DOM in lake water. The  
 422 EEM spectra were divided into five regions, which represented separately the different  
 423 compositions of organic compounds in water before and after coagulation (Chen et al.,  
 424 2003). In general, regions I and II represent aromatic proteins; region III stands for  
 425 fulvic acids; and regions IV and V are related to soluble microbial by-products and HA,  
 426 respectively.

427 Fig. 7 showed that after coagulation, the concentration of organic compounds in

428 the supernatant of zones III, IV, and V decreased significantly, and the concentrations  
 429 of organic compounds in zones I and II fell slightly. This outcome indicated that PBCF  
 430 was very efficient in removing xanthogenic acid and HA organic compounds, whereas  
 431 it exhibited poor removal efficiency of aromatic protein organic compounds. The main  
 432 reason for this situation is the high molecular weight of fulvic acid and HA etc. (regions  
 433 III, IV and V), which readily charge neutralization and bridging with PBCF.  
 434 Nevertheless, the aromatic proteins (regions I and II) are stable due to the presence of  
 435 their own benzene ring structure. Therefore, they are not easily removed (Loganathan  
 436 et al., 2020; Tian et al., 2020).



437  
 438 **Fig. 7.** (a) EEM of lake water before and (b) after treatment by PBCF

#### 439 4. Conclusions

440 A newly green flocculant (PBCF) was derived from natural polymer bagasse  
 441 cellulose. PBCF exhibited excellent performance in removing HA from water. It was  
 442 found that adsorption bridging and electric neutralization played a key role in removing  
 443 HA. Remarkably, PBCF can effectively reduce DOM and turbidity in natural lake water  
 444 via adsorption bridging interaction. Overall, this will offer a substitute flocculant for  
 445 water pretreatment and new insights into the flocculating process.

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- 571

## Highlights

- A cationic flocculant (PBCF) was prepared for using agricultural waste as a resource.
- PBCF was effective in removing HA at different concentrations.
- PBCF is highly efficient for  $UV_{254}$ ,  $COD_{Mn}$  and turbidity in lake water.
- Dose-pH graphs and correlation analyses confirmed flocculation during HA removal.
- Electrical neutralization and adsorption bridging dominated HA removal.