

Seroprevalence of Severe Acute Respiratory Syndrome Coronavirus 2-Specific Antibodies in Australia After the First Epidemic Wave in 2020: A National Survey

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Background. As of mid-2021, Australia's only nationwide coronavirus disease 2019 (COVID-19) epidemic occurred in the first 6 months of the pandemic. Subsequently, there has been limited transmission in most states and territories. Understanding community spread during the first wave was hampered by initial limitations on testing and surveillance. To characterize the prevalence of severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2)-specific antibody seroprevalence generated during this time, we undertook Australia's largest national SARS-CoV-2 serosurvey.

Methods. Between June 19 and August 6, 2020, residual specimens were sampled from people undergoing general pathology testing (all ages), women attending antenatal screening (20–39 years), and blood donors (20–69 years) based on the Australian population's age and geographic distributions. Specimens were tested by Wantai total SARS-CoV-2-antibody assay. Seroprevalence estimates adjusted for test performance were produced. The SARS-CoV-2 antibody-positive specimens were characterized with microneutralization assays.

Results. Of 11 317 specimens (5132 general pathology; 2972 antenatal; 3213 blood-donors), 71 were positive for SARS-CoV-2-specific antibodies. Seroprevalence estimates were 0.47% (95% credible interval [CrI], 0.04%–0.89%), 0.25% (CrI, 0.03%–0.54%), and 0.23% (CrI, 0.04%–0.54%), respectively. No seropositive specimens had neutralizing antibodies.

Conclusions. Australia's seroprevalence was extremely low (<0.5%) after the only national COVID-19 wave thus far. These data and the subsequent limited community transmission highlight the population's naivety to SARS-CoV-2 and the urgency of increasing vaccine-derived protection.

Keywords. Australia; COVID-19; SARS-CoV-2; seroprevalence; serosurvey.

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In the first 18 months of the coronavirus disease 2019 (COVID-19) pandemic, Australia has successfully pursued an aggressive suppression strategy [1]. As of July 2021, approximately 30 000 cumulative cases were reported nationally among the population of 26 million [2, 3]. The cumulative rate of COVID-19 in Australia, at 129 cases per 100 000 population, is in the lowest 20% of countries globally [4], and the testing rate per confirmed case is among the highest in the world [5]. In the first year of the pandemic, there were 2 distinct epidemic waves [3]. The first wave, in March–April 2020, affected all 8 Australian states and territories (hereafter jurisdictions), with approximately 7000 cases reported. During this time, the majority of reported cases were acquired overseas [6], but testing criteria were strongly focused on international travelers and their contacts [7], and a high number of the cases detected outside these groups could not be linked to a specific infection source [8]. Therefore, it was

possible that case notifications did not capture the full extent of virus transmission during this period. The second wave, in July–September 2020, was confined to the state of Victoria, with approximately 19 000 cases mainly concentrated in the state’s capital, Melbourne [7, 9]. In June 2021, a third COVID-19 outbreak began in Australia’s largest city, Sydney, with approximately 2000 cases reported to the end of July [3]. Outside of these periods, the vast majority of cases have been among quarantined international travelers and infrequently among short-lived community clusters with limited chains of transmission [10]. This reflects Australia’s successful control of COVID-19 through strict public health measures, including international and domestic border closures, mandatory 14-day quarantine for international arrivals, stay-at-home orders, extensive contact tracing and isolation activities, and comprehensive community testing and surveillance systems [1, 7, 10].

Tracking of the pandemic has been largely based on reported COVID-19 cases detected through virus-specific nucleic acid testing (NAT). Reported cases underestimate the true number of infections in the population [11], because they do not account for individuals who do not present for testing for various reasons including the following: having no or minimal symptoms [12], being ineligible (particularly early in the pandemic when NAT reagents were limited), or simply not presenting despite being symptomatic [13]. Since severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) infection

generates detectable antibodies in the vast majority of people [14], measuring population prevalence of SARS-CoV-2-specific antibodies provides an alternative means of estimating cumulative infections [15]. Such studies help to provide a more complete understanding of SARS-CoV-2 spread in diverse settings, which is a key global public health priority [15].

In addition to helping determine the extent to which case reporting has missed infections, serosurveys are key to estimating population susceptibility and hence reliance on vaccination to protect from SARS-CoV-2. Accordingly, we implemented Australia’s largest national SARS-CoV-2 serosurvey after the country’s first (and to date only) national COVID-19 epidemic period.

METHODS

The Australian COVID-19 Serosurveillance Network

The Australian COVID-19 Serosurveillance Network (<https://www.ncirs.org.au/our-work/serosurveillance>) was formed with representation from health departments in all 8 Australian jurisdictions, 5 major reference laboratories, Australian Red Cross Lifeblood (Lifeblood), and private and public laboratories involved in specimen collection (Figure 1). This structure allowed procedures to be aligned nationally and established a framework for repeat serosurveys. A laboratory expert subgroup was convened to advise on testing. In consultation with jurisdictional health departments and with the goal of achieving broad

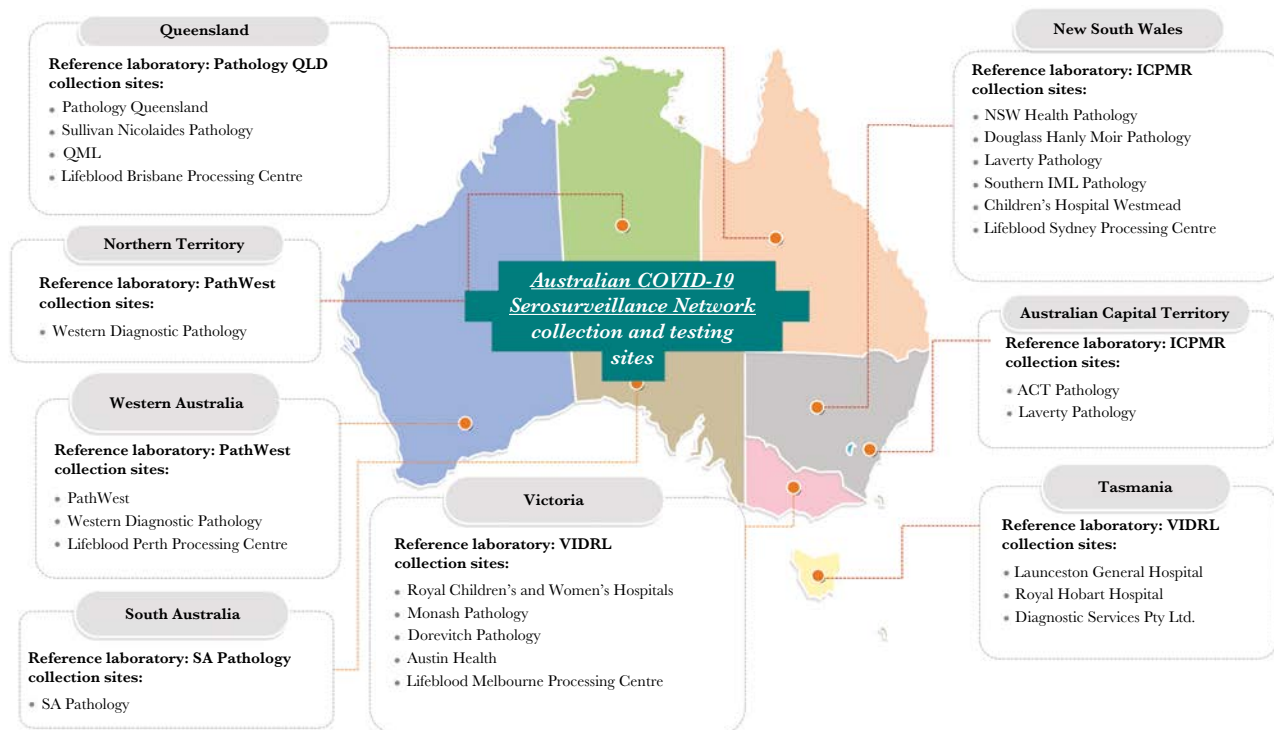


Figure 1. Australian COVID-19 Serosurveillance Network collection sites and reference laboratories by jurisdiction. ACT, Australian Capital Territory; ICPMR, Institute of Clinical Pathology and Medical Research; NSW, New South Wales; SA, South Australia; VIDRL, Victorian Infectious Diseases Reference Laboratory.

geographical coverage and population diversity, public and private laboratories were approached to participate in specimen collection for general pathology and antenatal screening populations. Nineteen representative public and private laboratories provided specimens (Figure 1).

Specimen Selection

Specimens were selected from 3 populations for this study: (1) general pathology - outpatients (all ages) who presented for a diagnostic blood test at participating public or private pathology services; (2) antenatal screening - pregnant women (20–39 years) who provided blood for antenatal screening at participating public or private pathology services; and (3) blood donors - Lifeblood donors (20–69 years) who provided blood at any Lifeblood donor center nationwide.

Deidentified residual blood specimens (serum, heparinized plasma, or EDTA plasma) were collected from the 3 populations. Eligible specimens were selected randomly or sequentially, depending on feasibility at each site (Table 1). Selection occurred upon receipt or before routine disposal until target numbers were achieved. Data were collected for each specimen on age, sex, and postcode, and, where permitted, initials and date of birth.

Sample Size and Distribution

A minimum sample size of 350 per subgroup (jurisdiction, age group, or sex) was adopted to allow exclusion of seroprevalence above 2.0% at the 95% confidence level in each subgroup, if observed seropositivity was at most 0.6%. To achieve geographic spread, we collected specimens proportional to the distribution of the Australian population in each of the 89 national Statistical Area 4 (SA4) regions [2, 16]. The SA4s were assigned according to the sample's residential postcode.

Testing

Specimens were tested at a reference laboratory using the Wantai SARS-CoV-2 total antibody enzyme-linked immunosorbent

assay ([ELISA] Beijing Wantai Biological Pharmacy Enterprise Co. Ltd., Beijing, China). An in-house validation found that the test sensitivity was 95.1% (95% confidence interval [CI], 88.9–98.4) among individuals with a positive SARS-CoV-2 NAT at least 14 days postillness onset (range, 14–130 days). Specificity among pre-pandemic blood specimens was 99.6% (95% CI, 98.9–99.9) (see Supplementary Material).

Specimens were considered ELISA-positive if they returned a positive (signal to cut-off ratio [S/CO] ≥ 1.1 on first and repeat Wantai testing) or equivocal result (S/CO = 0.9 to < 1.1 on first or repeat testing, or positive with nonpositive result upon repeat). To provide additional information on antibody profiles, ELISA-positive specimens were tested quantitatively for neutralizing antibody using a microneutralization assay ([MN] ICPMR, Westmead, New South Wales, Australia) [17]. The MN titers of 10 or more were considered positive.

Analysis

The timing of specimen collection was compared graphically to COVID-19 case notifications reported by a national public data-aggregation website [8]. Sex, age group, and geographic distribution was compared with the following reference populations: (1) - general pathology - Australian-estimated residential population, March 2020 [2]; (2) antenatal screening - the estimated population of women, 20–39 years, undergoing antenatal care (based on births by mother's age in 2019 [18]); and (3) blood donors - Lifeblood donors, 20–69 years, in 2019 (supplied by Lifeblood). Crude ELISA positivity and the distribution of S/CO values were summarized for each population.

Using ELISA results, we estimated the seroprevalence in each demographic subgroup using Bayesian methods to adjust for test sensitivity and specificity, incorporating the statistical uncertainty in these estimated values (see Supplementary Material). We summarized seroprevalence estimates using the median (point estimate) and 95% credible interval (CrI) of the corresponding posterior probability distribution.

Table 1. National Serosurvey Collection Populations and Specimen Eligibility Criteria

Collection Population	Eligible Age Range	Inclusion Criteria	Exclusion Criteria
General pathology	All ages	Specimens collected for routine pathology testing at public and private laboratories (including hospital outpatient and emergency departments)	Specimens from hospital inpatients and specimens submitted for a SARS-CoV-2 antibody diagnostic test
Antenatal screening	20–39 years	Specimens submitted for antenatal testing from pregnant females regardless of stage of pregnancy	Nil
Blood donors	20–69 years	Specimens from blood donors	Specimens from donors with a previous, self-reported COVID-19 infection who enrolled to donate for the first time after recovery from COVID-19 (ie, may have been recruited for convalescent plasma) ^a

Abbreviations: COVID-19, coronavirus disease 2019; SARS-CoV-2, severe acute respiratory syndrome coronavirus 2.

^aExclusion of these blood-donor specimens only occurred after a positive result on the Wantai SARS-CoV-2 total antibody assay.

Our primary analysis specified a “noninformative” prior distribution for seroprevalence that assumed uniform probability density between a predetermined lower bound and 100% for each demographic subgroup within each collection population. A sensitivity analysis was conducted using an alternative prior distribution that assigned a higher probability to low seroprevalence values, a beta distribution with parameters 0.2 and 10 (prior median prevalence ~1%). A separate analysis was also undertaken using the noninformative prior distribution, with equivocal ELISA results classified as negative.

Lower bounds for the prior distribution were calculated separately for demographic subgroups in each collection population, based on the cumulative number of nationally notified COVID-19 cases [19] reported at 14 days before the median specimen collection date, as a proportion of the corresponding estimated resident population [2].

The cumulative number of infections in the population during the first wave was estimated by multiplying the general pathology seroprevalence estimate by the Australian population [2]. The general pathology estimate was used because it encompasses all ages [20]. The infection-to-case-ratio was estimated using total estimated infections compared with the cumulative number of notified COVID-19 cases [19] reported at 14 days before the median serosurvey specimen collection date.

Ethics Approval

Ethics approvals were granted by the Sydney Children’s Hospital Network Human Research Ethics Committee ([SCHN HREC] HREC/17/SCHN/245); Lifeblood Ethics Committee (2020#07); University of Tasmania Health and Medical HREC (21875); Australian Capital Territory Health HREC (2020.LRE.00099); the Northern Territory Department of Health and Menzies School of Health Research HREC (2020-3768); and the Western Australian Department of Health HREC (RGS0000003804).

Patient Consent Statement

Because specimens were residual and deidentified, a waiver of consent was granted by all approving HRECs.

RESULTS

Sample Details

From June 19 to August 6, 2020, 11 317 eligible specimens were collected, comprising 5132 general pathology, 2972 antenatal screening, and 3213 blood donor specimens (Table 2). Sample size targets for each population were exceeded overall and by age group and sex, and at least 90% of targets were reached in all 8 jurisdictions, except for the antenatal screening collection in the 3 smallest jurisdictions: the Australian Capital Territory, Northern Territory, and Tasmania.

The distribution of age, sex, and jurisdiction among specimens broadly mirrored that of the corresponding reference populations (Table 2), except that our sample overrepresented

smaller jurisdictions and was more equally distributed across jurisdictions and age groups, to achieve minimum sample sizes by subgroup.

The overall median specimen collection date was July 2 (interquartile range [IQR] June 29–July 9), approximately 9 weeks after the approximate end of the first epidemic wave, and 15 weeks after its peak (Figure 2). Median collection dates for each population were within 1 week of each other: general pathology June 30, IQR June 25–July 6; antenatal screening July 3, IQR June 30–July 16; blood donors July 5, IQR July 2–July 8.

Antibody Testing Outcomes

The Wantai total SARS-CoV-2-antibody ELISA detected 71 positive specimens (29 general pathology, 15 antenatal screening, and 27 blood donor specimens) (Table 3). These were recorded in all jurisdictions and across multiple Australian SA4s without apparent clustering of infections (data not shown).

The median S/CO among ELISA positives was 2.1 (range, 0.9–21.1; IQR, 1.4–5.3). Blood donors had a broader distribution of S/CO values than other populations (median S/CO in general pathology 1.8, IQR 1.2–3.3; antenatal screening 2.3, IQR 1.3–6.4; blood donors 3.2, IQR = 1.5–15.7). Of the 71 ELISA positives, 13 had S/CO readings in the equivocal range, most frequently in the general pathology population (8 of 29 positives), followed by blood donors (3 of 27) and antenatal screening (2 of 15). No ELISA-positive specimens had quantifiable neutralizing antibodies.

Seroprevalence Estimates

After adjustment, primary estimates of SARS-CoV-2-specific antibody prevalence for the general pathology, antenatal screening, and blood donor populations were as follows: 0.25% (95% CrI, 0.03–0.54), 0.23% (95% CrI, 0.04–0.54), and 0.47% (95% CrI, 0.04–0.89), respectively. There were no statistically discernible differences in seroprevalence estimates between age groups, sex, or jurisdiction (Figure 3). In sensitivity analysis using the alternative prior distribution, seroprevalence estimates were expectedly lower at 0.15% (95% CrI, 0.03–0.45), 0.14% (95% CrI, 0.04–0.44), and 0.31% (95% CrI, 0.04–0.78), respectively. When equivocal ELISA results were classified as negative seroprevalence estimates were 0.17% (95% CrI, 0.03–0.39), 0.20% (95% CrI, 0.04–0.47), and 0.39% (95% CrI, 0.04–0.79), respectively.

Cumulative Severe Acute Respiratory Syndrome Coronavirus 2 Infections

As of June 16, 2020, there were 7636 nationally notified COVID-19 cases [19]. Based on the seroprevalence estimate of 0.25% in the general pathology population, cumulative SARS-CoV-2 infections nationally were estimated to be approximately 64 100 (95% CrI, 7700–138 500). The corresponding estimated infection-to-case-ratio was 8 (CrI, 1–18). Based on the lower seroprevalence estimate from the sensitivity analysis in

Table 2. Target Sample Size and Distribution of Demographic Characteristics in Each Collection and Their Australian Reference Populations

Demographic Group	General Pathology Sample						2020 Australian Population			Antenatal Screening Sample			Antenatal Population			Blood Donor Sample			Blood Donor Population				
	Target		Collected		Distribution of Collection (%)		Reference Population (n)	Reference Population (%)	Target (n)	Collected (n)	Distribution of Collection (%)	Reference Population (n)	Reference Population (%)	Target (n)	Collected (n)	Distribution of Collection (%)	Reference Population (n)	Reference Population (%)	Target (n)	Collected (n)	Distribution of Collection (%)		
	(n)	(n)	(n)	(n)	(%)	(%)	(n)	(%)	(n)	(n)	(%)	(n)	(%)	(n)	(n)	(%)	(n)	(%)	(n)	(n)	(%)		
Age group (years)	399	424	8.3	1 562 936	6.1																		
0 to <5	401	423	8.2	1 629 015	6.3																		
5 to <10	477	584	11.4	3 089 130	12.0																		
10 to <20	514	598	11.7	3 643 811	14.2			1395	1434	48.3	113 507	39.7	628	643	20.0	118 208	26.2						
20 to <30	518	609	11.9	3 747 564	14.6			1405	1538	51.7	172 304	60.3	632	658	20.5	105 927	23.4						
30 to <40	492	571	11.1	3 295 654	12.8																		
40 to <50	482	552	10.8	3 112 102	12.1																		
50 to <60	451	526	10.2	2 674 295	10.4																		
60 to <70	399	467	9.1	1 856 527	7.2																		
70 to <80	355	378	7.4	1 046 715	4.1																		
80+	2244	2787	54.3	12 938 247	50.4			2800	2972	100.0	285 811	100.0	1558	1576	49.1	229 386	50.8						
Female	2244	2345	45.7	12 719 502	49.6																		
Male	350	439	8.6	429 941	1.7			350	311	10.5	5 187	1.8	350	351	10.9	14 295	3.2						
ACT	1121	1295	25.0	8 160 113	31.8			350	732	24.6	92 235	32.3	557	581	18.1	131 055	29.0						
NSW	350	319	6.2	245 522	1.0			350	134	4.5	3309	1.2	350	368	11.5	2982	0.7						
NT	702	871	17.0	5 161 409	20.1			350	397	13.4	57 498	20.1	350	386	12.0	88 704	19.6						
QLD	350	354	6.9	1 768 045	6.9			350	352	11.8	18 299	6.4	350	343	10.7	35 940	8.0						
SA	350	426	8.3	539 732	2.1			350	287	9.7	5367	1.9	350	334	10.4	14 761	3.3						
TAS	905	1070	20.8	6 690 180	26.1			350	389	13.1	72 454	25.4	459	503	15.7	123 147	27.3						
VIC	360	368	7.2	2 658 135	10.4			350	370	12.4	31 439	11.0	350	347	10.8	40 854	9.0						
WA	4488	5132	100.0	25 653 077	100.0			2800	2972	100.0	285 788	100.0	3116	3213	100.0	451 738	100.0						
Total																							

Abbreviations: ACT, Australian Capital Territory; NSW, New South Wales; NT, Northern Territory; SARS-CoV-2, severe acute respiratory syndrome coronavirus 2; QLD, Queensland; SA, South Australia; TAS, Tasmania; VIC, Victoria; WA, Western Australia.

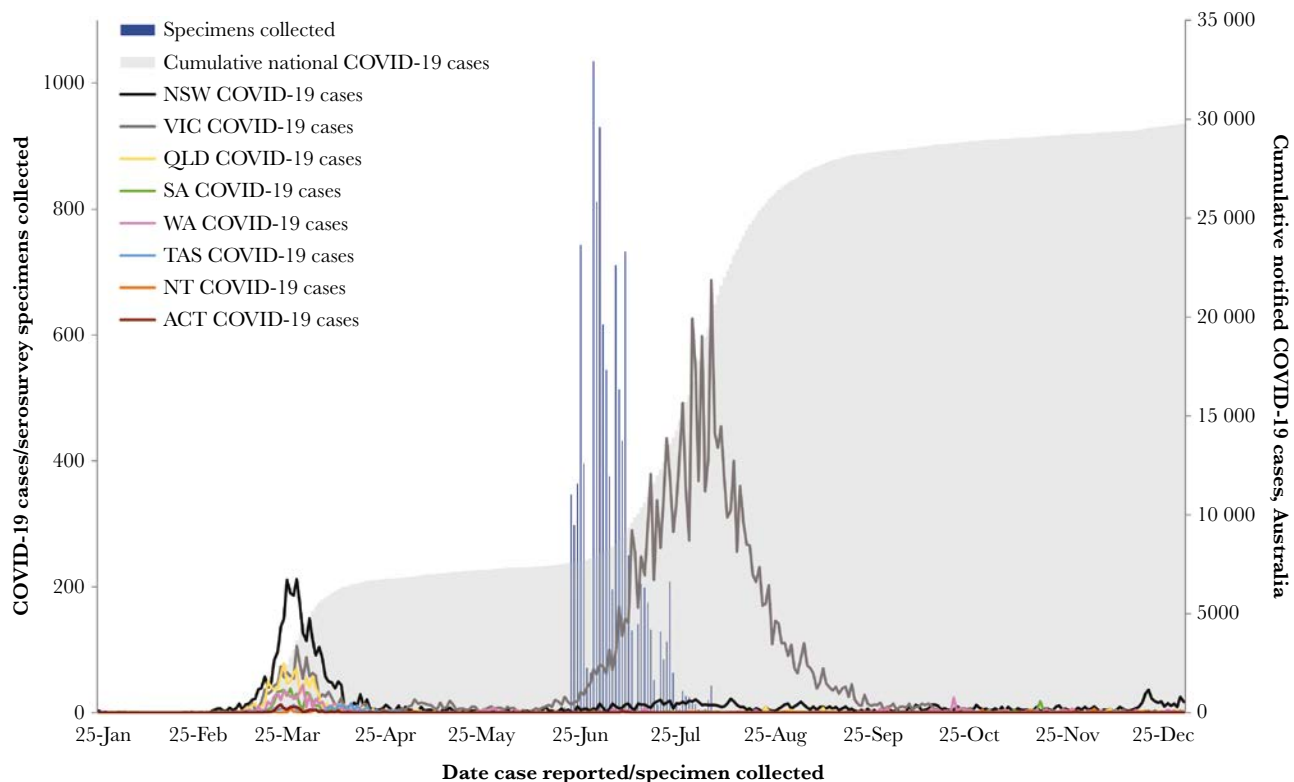


Figure 2. Australian coronavirus disease 2019 (COVID-19) cases and serosurvey specimens by date of report and jurisdiction or specimen collection, January 25 to December 31, 2020. ACT, Australian Capital Territory; NSW, New South Wales; NT, Northern Territory; QLD, Queensland; SA, South Australia; TAS, Tasmania; VIC, Victoria; WA, Western Australia.

the general pathology population of 0.15%, cumulative SARS-CoV-2 infections were estimated to be 38 500 (CrI, 7700–115 400), indicating an infection-to-case-ratio of 5 (CrI, 1–15).

DISCUSSION

We performed Australia’s largest national SARS-CoV-2 serosurvey to measure the spread of COVID-19 infection after the first epidemic wave in March–April 2020. As part of this, we generated important baseline seroprevalence data before the rollout of COVID-19 vaccinations in Australia. We showed that seroprevalence was very low (<0.5%), with no discernible differences between age groups, sexes, or jurisdictions. Furthermore, the 3 populations surveyed provided very similar results, increasing confidence in the generalizability of the findings to the broader community. Our findings suggest that routine surveillance may have missed approximately 7 infections for every notified case, with a credible range of no missed infections to 17 missed infections per case. Even at the upper limit of the credible intervals, the very low seroprevalence confirms that community transmission in Australia during the first wave was limited, despite being dispersed across all 8 jurisdictions. This demonstrates the success of the control measures implemented in March 2020, including the strict public health restrictions in place nationally until mid-May 2020 [6]. It also shows that the vast majority of the Australian population has

no infection-derived protection against SARS-CoV-2, emphasizing the crucial role of vaccination in increasing immunity.

Severe acute respiratory syndrome coronavirus 2 seroprevalence and vaccination coverage elsewhere in the world highlights Australia’s comparative lack of protection against COVID-19. High SARS-CoV-2 seroprevalence has been observed internationally; many estimates in the general population to mid-2021 have been over 20% and as high as 73% [21]. As of July 2021, approximately 14% of people globally are fully vaccinated, and several countries have reached over 60% vaccination coverage [22]. By comparison at the same time point, only 13% of the Australian population have been fully vaccinated, and our study demonstrates that Australia has had extremely limited acquisition of any potential “natural” protection [22]. With the exception of the second epidemic in the state of Victoria and the current outbreak in Sydney, COVID-19 has been successfully suppressed nationally since the first wave and after collection of our sample [3]. As such, even as of July 2021, it is likely that seroprevalence estimates obtained in this study approximate seroprevalence for the majority of Australian jurisdictions before the commencement of COVID-19 vaccinations in February 2021. Our results make clear Australia’s high susceptibility to further waves of infection and near-complete reliance on a successful, widespread vaccination program to forge a path out of the COVID-19 pandemic.

Table 3. Number and Proportion of Seropositive Specimens on the Wantai Total SARS-CoV-2 Antibody Assay by Collection, Age Group, Sex, and Jurisdiction

Demographic Group		General Pathology			Antenatal Screening			Blood Donors		
		Specimens Tested	Specimens Positive		Specimens Tested	Specimens Positive		Specimens Tested	Specimens Positive	
		(n)	(n)	(%)	(n)	(n)	(%)	(n)	(n)	(%)
Age group (years)	0 to <5	424	2	0.5						
	5 to <10	423	2	0.5						
	10 to <20	584	4	0.7						
	20 to <30	598	4	0.7	1434	5	0.3	643	5	0.8
	30 to <40	609	3	0.5	1538	10	0.7	658	4	0.6
	40 to <50	571	3	0.5				643	7	1.1
	50 to <60	552	4	0.7				639	6	0.9
	60 to <70	526	1	0.2				630	5	0.8
	70 to <80	467	4	0.9						
	80+	378	2	0.5						
Sex	Female	2787	15	0.5	2972	15	0.5	1576	15	1.0
	Male	2345	14	0.6				1637	12	0.7
Jurisdiction	Australia	5132	29	0.6	2972	15	0.5	3213	27	0.8
	ACT	439	1	0.2	311	1	0.3	351	4	1.1
	NSW	1285	9	0.7	732	8	1.1	581	6	1.0
	NT	319	5	1.6	134	1	0.7	368	2	0.5
	QLD	871	3	0.3	397	2	0.5	386	4	1.0
	SA	354	1	0.3	352	0	0.0	343	0	0.0
	TAS	426	2	0.5	287	1	0.3	334	3	0.9
	VIC	1070	5	0.5	389	2	0.5	503	3	0.6
	WA	368	3	0.8	370	0	0.0	347	5	1.4
Total		5132	29	0.6	2972	15	0.5	3213	27	0.8

Abbreviations: ACT, Australian Capital Territory; NSW, New South Wales; NT, Northern Territory; QLD, Queensland; SA, South Australia; SARS-CoV-2, severe acute respiratory syndrome coronavirus 2; TAS, Tasmania; VIC, Victoria; WA, Western Australia.

Of note, no neutralizing antibodies were detected among our ELISA-positive specimens. Evidence suggests that neutralizing antibodies can wane over time [23], with more rapid decline in those with mild or asymptomatic infections [24, 25]. In addition, some SARS-CoV-2 infections do not result in any detectable neutralizing antibodies [24]. Given that false positivity is already accounted for in our analysis, waning is the most likely explanation for the lack of detectable neutralizing antibody in our sample. Although the correlates and duration of protection after SARS-CoV-2 infection remain unknown, substantial waning of neutralizing antibody may have important implications for any infection-derived immunity in the Australian population.

Population-based SARS-CoV-2 serosurveys have consistently shown that COVID-19 case numbers underrepresent SARS-CoV-2 infections to a degree that is primarily dependent on testing rates [11, 21]. Although infection-to-case ratios should be cautiously interpreted, our estimation of some undetected SARS-CoV-2 transmission during the first wave is consistent with previous Australian serosurveys in more limited geographic settings [20, 26]. Some undetected transmission during this time is plausible, based on initially restricted testing policies [27], the continuation of international travel until mid-March 2020 [7], and the dispersion of community transmission

in every jurisdiction [8], allowing for mild disease to spread undetected. However, our serosurvey cannot differentiate between locally versus overseas-acquired infections, and as such it is unlikely to solely represent infections based on local transmission. Until mid-2021, any notable undetected transmission is likely to be isolated to the first wave, given the subsequent introduction of strict public health measures and widespread testing [1, 5, 7, 10].

Although we found no difference in seroprevalence estimates between age groups, it is possible that there was some age-related variation in the levels of undetected SARS-CoV-2 infection during the first wave. The rate of notified COVID-19 cases in those <18 years was approximately 10 times lower than in the adult population in early 2020 [6]. By contrast, seroprevalence estimates in those <20 years overlapped substantially with estimates in adult age groups (noting wide credible intervals and challenges in making comparisons in low prevalence settings). Children and adolescents have been shown to have higher rates of asymptomatic and mild infection than adults [28], which may contribute to a difference in detection of infections. Although patterns vary internationally [29], serosurveys in the United Kingdom [30] and China [31] have also demonstrated similar infection rates in children and adults despite lower rates of notified disease in younger groups.

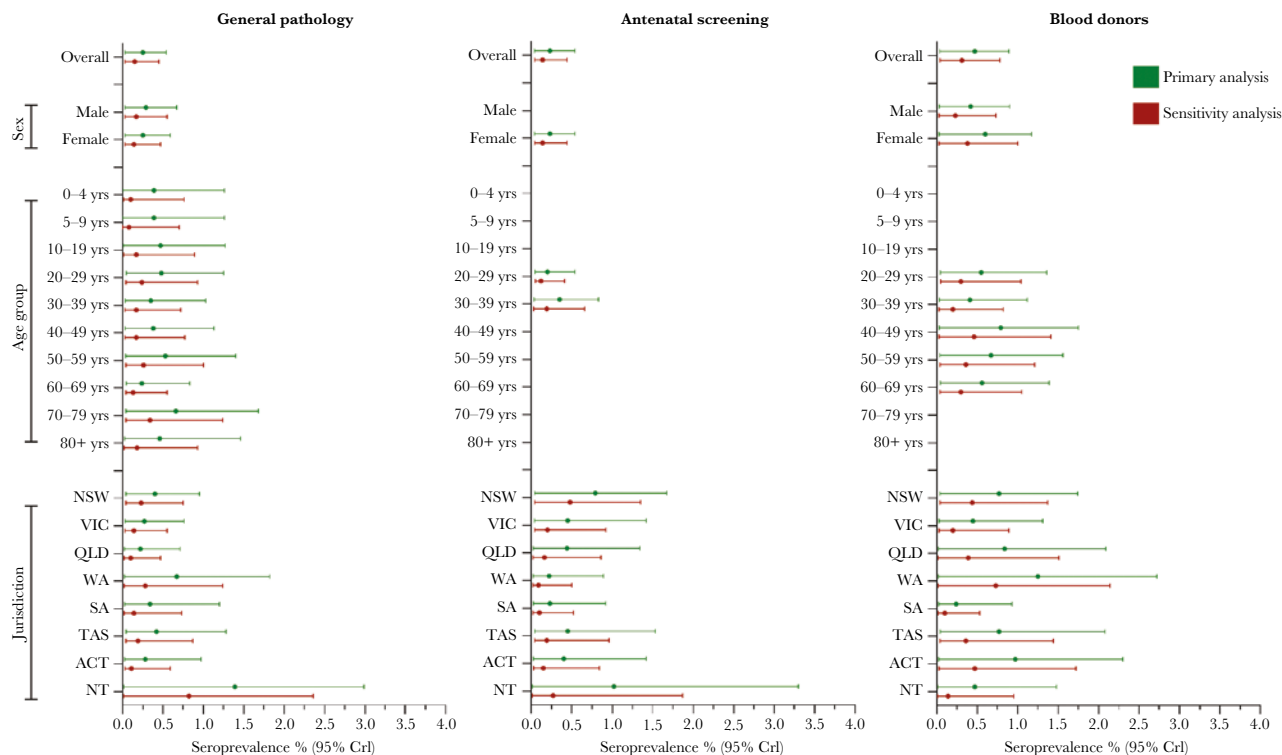


Figure 3. Estimated seroprevalence and 95% credible intervals by jurisdiction, age group, sex, and collection for Bayesian primary and sensitivity analyses. ACT, Australian Capital Territory; CrI, credible interval; NSW, New South Wales; NT, Northern Territory; QLD, Queensland; SA, South Australia; TAS, Tasmania; VIC, Victoria; WA, Western Australia.

Internationally, a wide variety of sampling methodologies have been used to conduct population SARS-CoV-2 serosurveys [11, 21]. Although probability sampling of the general population would be ideal to estimate seroprevalence [32], it is often not feasible and can introduce its own biases [33]. For this survey, collecting residual specimens was practical in the context of minimizing unnecessary close contact during the pandemic and facilitating rapid sampling at a significant time-point without lengthy consent-based procedures. To our knowledge, no national-level SARS-CoV-2 serosurveys have used more than 2 different residual specimen sources, and the vast majority use 1 [21]. Although residual specimens can only be representative of their source population, sampling 3 populations increased confidence in the plausible range of seroprevalence in the broader Australian community. The 3 populations have complementary characteristics. Blood donors are a “healthy”, well defined population frequently used in serosurveys [21], but they are limited to adults and may underrepresent sociodemographic groups, particularly those from culturally and linguistically diverse populations [34]. The general pathology population includes the full age spectrum, but it likely overrepresents those with high health system contact (eg, for chronic conditions), whose testing patterns may have changed during the pandemic. The antenatal screening population is limited in age range and sex, but it is likely to include broader sociodemographic groups [35] and be more stable despite the pandemic [36]. Taken together,

these populations provide a convenient and complementary cross section of the Australian community.

Published SARS-CoV-2 serosurveys have also used a variety of SARS-CoV-2 serological assays, with a diversity of performance characteristics [11]. Detecting SARS-CoV-2-specific antibody is dependent on the time since infection, disease severity, and the type and antigenic targets of the test used [17, 23, 25, 37, 38]. We chose to use the Wantai total antibody assay, recommended by the World Health Organization for population serosurveys, because it could be reliably run by all participating reference laboratories and it performed well in the literature as well as in local validation, with both high sensitivity and specificity compared with other commercially available assays [15, 39, 40]. A high specificity test is especially important in a low prevalence setting, as even with a specificity of 99.6%, we would expect approximately 45 of our 71 ELISA-positive specimens to be false positives.

A key contribution of this study was the establishment of a national network to enable further serosurveys for SARS-CoV-2 and potentially other communicable diseases. The network provides (1) a range of geographically diverse collection sites across the 3 populations, (2) strong links to jurisdictional health departments, and (3) laboratory expertise to guide testing algorithms. Serosurveys using networks such as ours provide a practical option to monitor evolving SARS-CoV-2 epidemiology, including vaccine impacts. The network also provides a potential mechanism to characterize the epidemiology and spread of future novel pathogens.

Our study has several limitations. Low seropositivity meant wide credible intervals and prevented population-weighted adjustments in analysis. The noninformative prior distribution was used to estimate seroprevalence in the primary analysis. While this is an objective and commonly preferred method, it's a conservative approach that may inflate infection estimates and related ratios. Further limitations exist related to our sampled populations. Truly random samples within these populations were not possible. Specimens were selected at random or sequentially depending on feasibility at each collection site, which may have introduced different selection biases. In South Australia, antenatal screening specimens were only available from women undergoing nuchal translucency testing (which incurs a cost to patients, while other antenatal screening is publicly funded) and may therefore represent a higher socioeconomic status group, at lower risk of COVID-19, than in other jurisdictions. However, the impact of this potential bias on the overall seroprevalence estimate among antenatal women was likely minimal, given that the group contributed to only 12% of the total sample. Finally, epidemiological investigation based on detecting COVID-19 antibodies alone may not capture individuals with recent infections who did not have time to mount detectable antibody responses. The impact of this on our estimates is likely to be negligible because specimen collection occurred during a time of very low COVID-19 activity. Only 1738 cases were notified during the study period, for all jurisdictions excluding Victoria. There was some overlap with our collection and the beginning of the Victorian second wave. This would have captured some recent infections resulting in an overestimate of seroprevalence for Victoria. However, we did not observe any discernible differences in estimates by jurisdiction, and it is likely that there was limited time for antibody development among those exposed at the time [14].

CONCLUSIONS

Our study provides evidence to further characterize the early and most geographically dispersed phase of the COVID-19 pandemic in Australia beyond findings obtained from case notifications alone. That there was some undetected virus transmission during Australia's first, and so far only, national COVID-19 wave is relatively unsurprising, but it reinforces the need to maintain strict control measures while a suppression strategy is being pursued. More importantly, even at the upper estimate of prevalence, Australia's SARS-CoV-2 antibody seroprevalence at mid-2020 was extremely low, highlighting the urgency of an effective vaccination program to increase protection from future outbreaks and severe outcomes.

Supplementary Data

Supplementary materials are available at *Open Forum Infectious Diseases* online. Consisting of data provided by the authors to benefit the reader, the posted materials are not copyedited and are the sole responsibility of the authors, so questions or comments should be addressed to the corresponding author.

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References

1. Australian Health Protection Principal Committee. Coronavirus (COVID-19) in Australia—Pandemic Health Intelligence Plan. Canberra: Australian Government Department of Health; 2020.
2. Australian Bureau of Statistics. Estimated resident population (ERP) by state/territory, sex and age to March 2020. Available at: http://stat.data.abs.gov.au/index.aspx?DatasetCode=ERP_QUARTERLY. Accessed 17 December 2020.
3. Australian Government Department of Health. Coronavirus (COVID-19) current situation and case numbers. Available at: <https://www.health.gov.au/news/health-alerts/novel-coronavirus-2019-ncov-health-alert/coronavirus-covid-19-current-situation-and-case-numbers>. Accessed 30 July 2021.
4. World Health Organization. WHO coronavirus (COVID-19) dashboard: vaccinations and cases. Available at: <https://covid19.who.int/>. Accessed 27 July 2021.

5. Hasell J, Mathieu E, Beltekian D, et al. A cross-country database of COVID-19 testing. *Sci Data* **2020**; 7:345.
6. COVID-19 National Incident Room Surveillance Team. COVID-19 Australia: epidemiology report 17—fortnightly reporting period ending 24 May 2020. *Commun Dis Intell* **2020**; 44:6.
7. COVID-19 National Incident Room Surveillance Team. COVID-19 Australia: epidemiology report 31—fortnightly reporting period ending 6 December 2020. *Commun Dis Intell* **2020**; 44:11–13.
8. O'Brien J. COVID-19 data Australia. Available at: <https://www.covid19data.com.au/>. Accessed 25 March 2021.
9. Victorian Department of Health and Human Services. Victorian coronavirus (COVID-19) data. Available at: <https://www.dhhs.vic.gov.au/victorian-coronavirus-covid-19-data>. Accessed 23 June 2021.
10. COVID-19 National Incident Room Surveillance Team. COVID-19 Australia: epidemiology report 43—reporting period ending 6 June 2021. *Commun Dis Intell* **2021**; 45:4.
11. Chen X, Chen Z, Azman AS, et al. Serological evidence of human infection with SARS-CoV-2: a systematic review and meta-analysis. *Lancet Glob Health* **2021**; 9:e598–609.
12. Oran DP, Topol EJ. The proportion of SARS-CoV-2 infections that are asymptomatic: a systematic review. *Ann Intern Med* **2021**; 174:655–62.
13. FluTracking. Weekly interim Report Australia: week ending 29 March 2020. Available at: <https://info.flutracking.net/reports-2/australia-reports/>. Accessed 20 April 2021.
14. Post N, Eddy D, Huntley C, et al. Antibody response to SARS-CoV-2 infection in humans: a systematic review. *PLoS One* **2020**; 15:e0244126.
15. World Health Organization. Population-based age-stratified seroepidemiological investigation protocol for coronavirus 2019 (COVID-19) infection (Version 2.0). Available at: <https://www.who.int/publications/i/item/WHO-2019-nCoV-Seroepidemiology-2020.2>. Accessed 17 January 2021.
16. Australian Bureau of Statistics. 1270.0.55.001—Australian Statistical Geography Standard (ASGS): volume 1—main structure and greater capital city statistical areas, July 2016 2021. Available at: <https://www.abs.gov.au/AUSSTATS/abs@nsf/DetailsPage/1270.0.55.001July%202016?OpenDocument>. Accessed 22 April 2021.
17. Hueston L, Kok J, Guibone A, et al. The antibody response to SARS-CoV-2 Infection. *Open Forum Infect Dis* **2020**; 7:ofaa387.
18. Australian Bureau of Statistics. Births by age and state to 2019. Available at: http://stat.data.abs.gov.au/index.aspx?DatasetCode=FERTILITY_AGE_STATE. Accessed 17 December 2020.
19. Australian Government Department of Health. Introduction to the national notifiable diseases surveillance system. Available at: <https://www1.health.gov.au/internet/main/publishing.nsf/Content/cda-surveil-nndss-nndssintro.htm>. Accessed 9 July 2021.
20. Gidding HF, Machalek DA, Hendry AJ, et al. Seroprevalence of SARS-CoV-2-specific antibodies in Sydney after the first epidemic wave of 2020. *Med J Aust* **2021**; 214:179–85.
21. Arora RK, Joseph A, Van Wyk J, et al. SeroTracker: a global SARS-CoV-2 seroprevalence dashboard. *Lancet Infect Dis* **2021**; 21:e75–6.
22. Mathieu E, Ritchie H, Ortiz-Ospina E, et al. A global database of COVID-19 vaccinations. *Nat Hum Behav* **2021**. doi: [10.1038/s41562-021-01122-8](https://doi.org/10.1038/s41562-021-01122-8).
23. Seow J, Graham C, Merrick B, et al. Longitudinal observation and decline of neutralizing antibody responses in the three months following SARS-CoV-2 infection in humans. *Nat Microbiol* **2020**; 5:1598–607.
24. Chia WN, Zhu F, Ong SWX, et al. Dynamics of SARS-CoV-2 neutralising antibody responses and duration of immunity: a longitudinal study. *Lancet Microbe* **2021**. doi: [10.1016/S2666-5247\(21\)00025-2](https://doi.org/10.1016/S2666-5247(21)00025-2).
25. Röltgen K, Powell AE, Wirz OF, et al. Defining the features and duration of antibody responses to SARS-CoV-2 infection associated with disease severity and outcome. *Sci Immunol* **2020**; 5:eabe0240. doi: [10.1126/sciimmunol.abe0240](https://doi.org/10.1126/sciimmunol.abe0240).
26. Hicks SM, Pohl K, Neeman T, et al. A dual-antigen enzyme-linked immunosorbent assay allows the assessment of severe acute respiratory syndrome coronavirus 2 antibody seroprevalence in a low-transmission setting. *J Infect Dis* **2021**; 223:10–4.
27. Murphy B. Chief Medical Officer update on coronavirus testing. Available at: <https://www1.racgp.org.au/newsgp/professional/chief-medical-officer-update-on-coronavirus-testin>. Accessed 24 February 2021.
28. Riccardo F, Ajelli M, Andrianou XD, et al. Epidemiological characteristics of COVID-19 cases and estimates of the reproductive numbers 1 month into the epidemic, Italy, 28 January to 31 March 2020. *Euro Surveill* **2020**; 25:2000790. doi: [10.2807/1560-7917.ES.2020.25.49.2000790](https://doi.org/10.2807/1560-7917.ES.2020.25.49.2000790).
29. Viner RM, Mytton OT, Bonell C, et al. Susceptibility to SARS-CoV-2 infection among children and adolescents compared with adults: a systematic review and meta-analysis. *JAMA Pediatr* **2021**; 175:143–56.
30. Ladhani SN, Baawuah F, Beckmann J, et al. SARS-CoV-2 infection and transmission in primary schools in England in June–December, 2020 (SKIDS): an active, prospective surveillance study. *Lancet Child Adolesc Health* **2021**. doi: [10.1016/S2352-4642\(21\)00061-4](https://doi.org/10.1016/S2352-4642(21)00061-4).
31. Bi Q, Wu Y, Mei S, et al. Epidemiology and transmission of COVID-19 in 391 cases and 1286 of their close contacts in Shenzhen, China: a retrospective cohort study. *Lancet Infect Dis* **2020**; 20:911–9.
32. Gronvall G, Connell N, Farley J, et al. Developing a National Strategy for SARS-CoV-2 Serosurveys in the United States. Baltimore, United States: Johns Hopkins Center for Health Security; **2020**.
33. Gidding H. Australia's national serosurveillance program. *New South Wales Public Health Bull* **2003**; 14:90–3.
34. Davison T, Daly J, Flower R. Australia's ethnic face is changing, and so are our blood types. Available at: <https://theconversation.com/australias-ethnic-face-is-changing-and-so-are-our-blood-types-113454>. Accessed 1 April 2021.
35. Australian Institute of Health and Welfare. Health of mothers and babies. Available at: <https://www.aihw.gov.au/reports/australias-health/health-of-mothers-and-babies>. Accessed 1 April 2021.
36. Australian Institute of Health and Welfare. Antenatal care during COVID-19, 2020. Available at: <https://www.aihw.gov.au/reports/mothers-babies/antenatal-care-during-covid-19/contents/did-access-to-antenatal-care-change-during-the-covid-19-pandemic>. Accessed 22 April 2021.
37. Orth-Höller D, Eigentler A, Stiasny K, Weseslindtner L, Möst J. Kinetics of SARS-CoV-2 specific antibodies (IgM, IgA, IgG) in non-hospitalized patients four months following infection. *J Infect* **2021**; 82:282–327.
38. Long Q-X, Tang X-J, Shi Q-L, et al. Clinical and immunological assessment of asymptomatic SARS-CoV-2 infections. *Nat Med* **2020**; 26:1200–4.
39. GeurtsvanKessel CH, Okba NMA, Igloi Z, et al. An evaluation of COVID-19 serological assays informs future diagnostics and exposure assessment. *Nat Commun* **2020**; 11:3436.
40. Nicholson S, Karapanagiotidis T, Khvorov A, et al. Evaluation of six commercial SARS-CoV-2 serology assays detecting different antibodies for clinical testing and serosurveillance. *Open Forum Infect Dis* **2021**. doi: [10.1093/ofid/ofab239](https://doi.org/10.1093/ofid/ofab239).