FUNCTIONAL CHARACTERISATION OF A NOVEL IMMUNE MODULATORY MOLECULE FROM FASCIOLA HEPATICA

by

Raquel Alvarado B.Sc. (Hons)

A Thesis Submitted for the Degree of

Doctor of Philosophy in Science

School of Medical and Molecular Biosciences, Faculty of Science,
University of Technology Sydney, Australia.

2014

CERTIFICATE OF AUTHORSHIP/ORIGINALITY

I certify that the work in this thesis has not previously been submitted for a degree nor

has it been submitted as part of requirements for a degree except as fully acknowledged

within the text.

I also certify that the thesis has been written by me. Any help that I have received in my

research work and the preparation of the thesis itself has been acknowledged. In

addition, I certify that all information sources and literature used are indicated in the

thesis.

Production Note:

Signature of Student: Signature removed prior to publication.

Date: ____22nd December, 2014

i

ACKNOWLEDGEMENTS

Science has fascinated me ever since I was a child, therefore I am very grateful to God for not only giving me the opportunity, but also the willingness and strength to pursue my studies in this field that I truly enjoy. I consider myself very fortunate for being able to do this PhD project, which despite being my greatest challenge as a student, has rewarded me with getting to know so many people who have generously shared their knowledge and friendship with me.

To my parents, I thank them for all their love, for listening, encouraging me and always giving me their complete support and to my mum in particular for being my best friend. Without them I would simply not have been able to achieve this goal.

I thank Bronwyn O'Brien, Sheila Donnelly and Mark Robinson, for giving me the privilege of working with them. It was Bronwyn's enthusiastic Immunology lectures that first showed me how interesting this field can be. I really appreciate the direct involvement in the laboratory of Sheila and Mark, who patiently dedicated a lot of their time to teach me new techniques and help me with experiments. I want to thank the three of them for giving me such an interesting project, for teaching me so much, checking my drafts, giving their valuable feedback, for all their guidance and overall for being such excellent supervisors. I'm also very grateful to Joyce To for helping me so much in this project and for teaching me her clever modifications of experimental techniques. To Maria Lund, Andrew Hutchinson and Paddy McCauley-Winter, I thank all their assistance in flow cytometry and their valuable advice. I am very proud to have been part of this research team, whose members were so kind to me and I wish them all very successful futures with the hope that our friendship endures a very long time.

In these years I have also met fellow postgraduate students with whom I have acquired what I would love to be lifelong friendships, and with whom I have shared unforgettable moments. I want to thank them for being so wonderful, especially to Maria Lund, Alex Gale and Rita Rapa for letting me count on them in good and difficult moments.

I would also like to express my gratitude to the UTS technical staff, especially Mike Johnson and Lynne Turnbull for their assistance in microscopy. Also, I thank Ashley Mansell, Anita Pinar and Suat Dervish for their contribution to my project and the science faculty for granting me a postgraduate research scholarship.

PUBLICATIONS ASSOCIATED WITH THIS THESIS

Journal Publications

Mark W. Robinson, **Raquel Alvarado**, Joyce To, Andrew Hutchinson, Stephanie N. Dowdell, Maria Lund, Lynne Turnbull, Cynthia B. Witchurch, Bronwyn A. O'Brien, John P. Dalton and Sheila Donnelly, 2012; A helminth cathelicidin-like protein suppresses antigen processing and presentation in macrophages via inhibition of lysosomal vATPase. FASEB Journal. 26, 4614-4627.

Raquel Alvarado, Bronwyn O'Brien, Akane Tanaka, John P. Dalton, Sheila Donnelly, 2014; A parasitic helminth-derived peptide that targets the macrophage lysosome is a novel therapeutic option for autoimmune disease. Immunobiology. (In press, DOI 10.1016/j.imbio.2014.11.008).

Raquel Alvarado, Joyce To, Anita Pinar, Ashley Mansell, Mark Robinson, Bronwyn O'Brien, John Dalton, Sheila Donnelly, 2014; The *F. hepatica* cathelicidin like peptide (FhHDM-1) modulates the activation of the NLRP3 inflammasome in macrophages. (Manuscript in preparation).

Conference presentations

Raquel Alvarado, Maria Lund, Mark W. Robinson, Andrew Hutchinson, Joyce To, John P. Dalton, Bronwyn O'Brien and Sheila Donnelly, 2012; Presentation entitled: A novel molecule secreted by *Fasciola hepatica* modulates the activity of innate immune cells. 42nd Annual Scientific Meeting of the Australasian Society for Immunology.

Raquel Alvarado, Joyce To, Maria Lund, Mark W. Robinson, Andrew Hutchinson, Bronwyn O'Brien, John P. Dalton and Sheila Donnelly, 2013; Presentation entitled: A novel molecule secreted by the parasite *F. hepatica* modulates the response of macrophage NLRP3 inflammasome. 27th Annual Conference of the European Macrophage & Dendritic Cell Society.

TABLE OF CONTENTS

Certif	icate of authorship/originality	i
Ackno	owledgements	ii
Public	cations associated with this thesis	iii
Table	of contents	iv
List o	f illustrations	ix
List o	f tables	xi
Abbre	eviations	xii
Abstra	act	xvi
Chapt	er 1 General Introduction	1
1.1.	Versatility of the mammalian immune system	1
1.2.	Helminth parasites are master regulators of the mammalian immune system	n2
1.3.	Characterisation of immune modulatory components of helminth	
	excretory/secretory products	5
1.3.1.	Proteases	6
1.3.2.	Protease inhibitors	7
1.3.3.	IPSE/alpha-1, Omega-1 and other helminth glycans	9
1.3.4.	Cytokine homologues	10
1.3.5.	Antioxidants	11
1.3.6.	ES-62 a phosphorylcholine (PC) containing glycoprotein	12
1.3.7.	Additional ES products that are immune modulators	13
1.4.	Fasciola hepatica: A model of helminth-induced immune-modulation	18
1.4.1.	FhCL1	20
1 4 2	FhPry	21

1.4.3.	FhHDM-1	22
1.5.	FhHDM-1 is a helminth-derived peptide with homology to the mammalian cathelicidin defence peptides	24
1.5.1.	A proposed mechanism of action for FhHDM-1	27
Chapte	er 2 General Materials & Methods	29
2.1.	Production of RecFhHDM-1, sFhHDM-1 and anti-FhHDM-1 antibody	30
2.2.	Cell culture	31
2.2.1.	Sterility	31
2.2.2.	Cell lines	31
2.2.2.1	. RAW264.7 macrophages	31
2.2.2.2	2. ASC macrophages	31
2.2.3.	Primary cells	32
2.2.3.1	. Murine Bone Marrow-Derived Macrophages (BMDMs)	32
2.2.3.1	.1. Isolation of bone marrow cells	32
2.2.3.1	.2. Differentiation of Bone Marrow-Derived Macrophages	32
2.2.3.2	2. Human Monocyte Isolation and Macrophage Differentiation	32
2.3.	Immunofluorescence Confocal Microscopy	33
2.3.1.	NPG Antifade Mounting Media preparation	35
2.4.	Flow Cytometry	35
2.5.	ELISAs	35
2.6.	Statistical Analysis	36
Chapte	er 3 Characterisation of the interaction between FhHDM-1 and macrophages .	37
3.1.	Introduction	37
3.2.	Specific methods	39
3.2.1.	Cholesterol binding assay	39
3.2.2.	Immunofluorescent Confocal Microscopy	39
3.2.2.1	Co-localisation of FhHDM-1 with lipid rafts	39

3.2.2.2. Inhibitor Studies	40
3.2.2.3. Co-localisation of FhHDM-1 with organelle markers	40
3.2.2.3.1. Localisation of FhHDM-1 with caveolae, Golgi and mitochondria	40
3.2.2.3.2. Temporal localisation of FhHDM-1 with early endosomes	41
3.2.2.3.3. Temporal localisation of FhHDM-1 with late endosomes/lysosomes	41
3.3. Results	43
3.3.1. FhHDM-1 binds to cholesterol and interacts with lipid rafts	43
3.3.2. FhHDM-1 is actively endocytosed by a cytoskeletal-dependent mechanism	45
3.3.3. Endocytosis of FhHDM-1 involves early endosomal and lysosomal colocalisation	50
3.4. Discussion	58
Chapter 4 FhHDM-1 modulates the processing of antigens by macrophages	62
4.1. Introduction	62
4.2. Specific methods	65
4.2.1. Endocytosis and vesicular acidification studies	65
4.2.1.1. Dextran endocytosis studies	65
4.2.1.1.1. Flow Cytometry	65
4.2.1.1.2. Confocal microscopy	66
4.2.2. Effects of FhHDM-1 on antigen processing and presentation	66
4.2.2.1. Effects of sFhHDM-1 on DQ Ovalbumin processing	66
4.2.2.2. Effects of FhHDM-1 on antigen presentation to transgenic murine cells	67
4.2.2.2.1. Antigen processing studies	67
4.2.2.2.2. T cell isolation	67
4.2.2.2.3. Antigen presentation studies	68
4.2.3. Effects of FhHDM-1 on MHCII surface expression by BMDMs	
4.3. Results	69

4.3.1. FhHDM-1 reduces vesicular acidification and enhances endocytosis by	7
macrophages	69
4.3.2. FhHDM-1 reduces antigen processing by macrophages	76
4.3.3. Effects of FhHDM-1 on antigen presentation by macrophages	78
4.4. Discussion	82
Chapter 5 FhHDM-1 reduces NLRP3 inflammasome activation in macrophage	es87
5.1. Introduction	87
5.2. Specific methods	91
5.2.1. FhHDM-1 effects on NLRP3 Inflammasome Activation	91
5.2.1.1. NLRP3 activation	91
5.2.1.2. Cytokine detection by ELISA	91
5.2.1.3. Detection of activated IL-1β and Caspase-1 by Western Blot	91
5.2.1.3.1. TCA supernatant protein precipitation	92
5.2.1.3.2. Gel Electrophoresis	92
5.2.1.3.3. Western Blotting	92
5.2.2. ASC speck formation	93
5.2.3. FhHDM-1 effects on lysosomal integrity	93
5.2.3.1. Lysosomal stability	93
5.2.3.1.1. DQ Ova compartmentalisation	93
5.2.3.2. Cathepsin B activity	94
5.3. Results	95
5.3.1. FhHDM-1 inhibits lysosomal-dependent NLRP3 activation	95
5.3.1.1. FhHDM-1 reduces NLRP3 inflammasome activation by ALUM	95
5.3.1.2. FhHDM-1 reduces NLRP3 inflammasome activation by Nano-SiO ₂	96
5.3.2. FhHDM-1 prevents NLRP3 dependent ASC oligomerisation	98
5.3.3. Effects of FhHDM-1 on lysosomal integrity and stability after ALUM-	induced
inflammasome activation	101

5.3.4.	Effects of FhHDM-1 on LPS priming.	. 105
5.4.	Discussion	. 107
Chapte	er 6 General Discussion	.111
Refere	ences	.114

LIST OF ILLUSTRATIONS

Figure 1.1 Characterization of FhHDM-1 and structural homology to LL-37	.23
Figure 3.1 FhHDM-1 binds to cholesterol and co-localises with lipid rafts in the plasm membranes of macrophages.	
Figure 3.2 FhHDM-1 is internalised by macrophages and localises in close proximity cytoskeletal networks.	
Figure 3.3 Internalisation of FhHDM-1 by macrophages is an active process dependent upon cholesterol, as well as actin and microtubule networks.	
Figure 3.4 FhHDM-1 endocytosis was not mediated by caveolae	.51
Figure 3.5 FhHDM-1 co-localised with early endosomes.	.52
Figure 3.6 FhHDM-1 co-localised with endolysosomes of fixed RAW264.7 macrophages.	. 54
Figure 3.7 FhHDM-1 co-localised with endolysosomes of live BALBc BMDMs	. 55
Figure 3.8 FhHDM-1 did not co-localise with the Golgi apparatus or mitochondria in RAW264.7 macrophages.	
Figure 4.1 Macrophage pre-treatment with FhHDM-1 decreased dextran detection in acidified vesicles but not its endocytosis.	.72
Figure 4.2 Simultaneous incubation of macrophages with FhHDM-1 did not affect the detection of dextran in acidified vesicles, but it did increase the uptake of dextran	
Figure 4.3 Macrophages treated with FhHDM-1 can endocytose dextran and simultaneous incubation with both molecules enhanced the co-localisation of FhHDM and dextran.	
Figure 4.4 Simultaneous incubation with FhHDM-1 and ovalbumin reduces the ability of macrophages to process antigen.	-
Figure 4.5 FhHDM-1 reduces the ability of macrophages to process antigenic peptide for their presentation to T cells.	
Figure 4.6 FhHDM-1 did not alter MHCII expression levels by macrophages	.81
Figure 5.1 NLRP3 activation pathway	89

Figure 5.2 FhHDM-1 reduced NLRP3 inflammasome activation induced by ly	/sosomal
destabilising agents.	97
Figure 5.3 FhHDM-1 reduced ASC speck formation.	100
Figure 5.4 FhHDM-1 does not prevent the lysosomal destabilisation induced by	y ALUM.
	103
Figure 5.5 FhHDM-1 treatment of macrophages reduced cathepsin B activity.	104
Figure 5.6 FhHDM-1 reduced LPS priming efficiency in macrophages	106

LIST OF TABLES

Table 1.1 Classification of identified helminth ES components and their immune	
modulatory/evasive effects.	16
Table 2. 1 General materials and reagents.	29
Table 2.2 List of antibodies and dyes used for confocal microscopy experiments	34

ABBREVIATIONS

ACEC Animal Care and Ethic Committee

ADCC Antibody dependent cell mediated cytotoxicity

AIM2 Absent in melanoma 2

ALUM Aluminium salts

AMPs Antimicrobial peptides
APC(s) Antigen presenting cell(s)
ARC Animal Resources Centre

ASC Apoptosis associated speck like protein

ATCC American Type Culture Collection

Az Azide

BcR B cell receptor

BMDMs Bone marrow derived macrophages

BSA Bovine serum albumin

CAT Catalases
Cav-1 Caveolin-1

ChTx Cholera toxin subunit B

CLIC Clathrin-independent non-caveolar pathway

CO₂ Carbon dioxide

DAMPs Damage-associated molecular patterns

4'6-diamidino-2 phenylindole, dilactate

DMSO Dimethyl sulfoxide

DC(s) Dendritic cell(s)

E. coli Escherichia coli

ELISA Enzyme-linked immunosorbent assay

ES Excretory secretory products

FACS Fluorescence-activated cell sorting (flow cytometry)

FBS Foetal bovine serum

FhCL1 Fasciola hepatica cathepsin L-1

FhES Fasciola hepatica excretory/secretory products **FhHDM-1** Fasciola hepatica helminth defence molecule-1

FhHDM-1p2 FhHDM-1 peptide 2

FhPrx Fasciola hepatica peroxiredoxin

Geo Mean Geometric mean

GPx Glutathione peroxidase

hCAP18 Human cationic antimicrobial protein 18kDa

HDMs Helminth defence molecules

HDPs Host defence peptides

HEPES 4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid

HVS Herpesvirus saimiri

IAPP Islet amyloid polypeptide

IFNγ Interferon gammaIg Immunoglobulin

IL Interleukin

IMDM Iscove's modified Dulbecco's medium

IPSE IL-4 inducing principle of schistosome eggs

kDa Kilo daltons

LBP LPS-binding protein

LNFPIII Lacto-N-fucopentaose III

LPS Lipopolysaccharide

MAPKs Mitogen-activated protein kinases

MCD Methyl-β-cyclodextrin

M-CSF Macrophage colony stimulating factor

MHC Major histocompatibility complex

MIC Minimal concentration capable of inhibiting visible microbial

growth

MIF Migration inhibitory factor
MPR Mannose phosphate receptor
mRNA Messenger ribonucleic acid

MS Multiple sclerosis
MSU Mono sodium urate
MW Molecular weight

MyD88 Myeloid differentiation factor 88

N/A Not applicable

Na₂CO₃ Sodium carbonate

Nano-SiO₂ Silicon dioxide (nanoparticles)

NH₄Cl Ammonium chloride

NPG N-propyl gallate microscopy mounting media

NLRs NACHT-leucine-rich repeat receptors

(NOD)-like receptors

Nucleotide-binding oligomerisation domain protein like

receptors

OD Optical density

O/N Overnight

OPep Ovalbumin peptide

Ova Ovalbumin

PAMPS Pathogen-associated molecular patterns

PBMCs Peripheral blood mononuclear cells

PBS Phosphate buffered saline

PC Phosphorylcholine
PFA Paraformaldehyde
PGE₂ Prostaglandin E2

PI Peak I Peak II

PI 3-K Phosphoinositide 3 kinase

Prx Peroxiredoxin

RecFhHDM-1 Recombinant *Fasciola hepatica* helminth defence molecule 1

RELM-α Resistin-like molecule-alpha

RIPA Radioimmunoprecipitation Assay Buffer

ROS Reactive oxygen species

RPMI Roswell Park Memorial Institute 1640 medium

RP-HPLC Reversed-phase high performance liquid chromatography

RT Room temperature

SmCB1 Schistosoma mansoni cathepsin B

SDS-PAGE Sodium dodecyl sulphate- polyacrylamide gel electrophoresis

SEA Soluble egg antigens

SEMs Standard errors of the means

sFhHDM-1 Synthetic *Fasciola hepatica* helminth defence molecule-1

Silicon dioxide (nanoparticles)

SOD Superoxide dismutases

T1D Type I diabetes
TBS Tris buffer saline
TcR T cell receptor

TGF-β Transforming growth factor beta

Trans-Golgi network **TGN**

T helper cells Th

Tyrosine kinase interacting protein Tip

Toll like receptor TLR

TNF Tumour necrosis factor

3,3',5,5'-Tetramethylbenzidine liquid substrate system for **TMB**

ELISA

T regulatory cells Treg

TIR domain-containing adaptor inducing IFN- β **TRIF**

Triton X 100 **TX100**

Vacuolar adenosine triphosphatase vATPase

 \mathbf{v}/\mathbf{v} Volume / volume Weight / volume \mathbf{w}/\mathbf{v}

ABSTRACT

The ability of tissue dwelling helminth parasites to induce chronic long term infections, is enabled by the establishment of T helper 2/ regulatory T cell (Th2/Treg) immune responses within their mammalian hosts. Such responses prevent the expulsion of the parasites, whilst simultaneously avoiding excessive inflammation/fibrosis arising within the host, as a consequence of tissue damage induced by helminth migration. Importantly, helminths excrete and secrete a series of molecules (collectively known as ES products), which not only play major roles in parasite biology, but also exert direct immune modulatory functions, promoting the establishment of Th2/Treg immunity. The trematode, *Fasciola hepatica*, is an excellent model of helminth-mediated immune modulation, because it induces a very rapid switch towards Th2 responses in its mammalian hosts and inhibits Th1 immunity. Fractionation of the ES products of *F. hepatica* has identified three major immune modulatory components: the protease cathepsin L1, the antioxidant peroxiredoxin, and a previously uncharacterised peptide, FhHDM-1.

Structural analysis of FhHDM-1 revealed a close resemblance to the cathelicidin, LL-37, a well characterised mammalian immune-modulating peptide. Therefore, a putative immune modulatory role for FhHDM-1 was explored in this project. Immunofluorescent confocal microscopy demonstrated that FhHDM-1 interacted with macrophage lipid rafts, prior to being actively internalised by cholesterol- and cytoskeletal network-dependent endocytosis, with progressive compartmentalisation of the peptide into early endosomes and endolysosomal vesicles. Flow cytometry studies indicated that, once internalised, FhHDM-1 enhanced the rate of endocytosis of dextran by macrophages. Despite this, FhHDM-1 was found to impair the acidification of macrophage endolysosomes and as a consequence, the efficient processing and subsequent presentation of ovalbumin to T cells was prevented, as assessed by decreased detection of digested fluorescent ovalbumin and reduced IL-2 secretion by transgenic CD4⁺ T cells. Additionally, FhHDM-1 impaired NLRP3 inflammasome activation by lysosomal disruptive agents in macrophages. This was found to be a consequence of reduced cathepsin B activity (due to FhHDM-1 induced suboptimal lysosomal acidification), which was incapable of stimulating inflammasome complex formation, thus avoiding IL-1 β and caspase-1 cleavage.

These findings suggest that by targeting endolysosomal activity, FhHDM-1 limits macrophage function. Therefore, the current study is the first to demonstrate that FhHDM-1 possesses immune modulatory properties, which are directed by a mechanism not previously described for a helminth-secreted cathelicidin-like peptide.