Development of hydroculture plants for the improvement of indoor air quality



Peter Irga
University of Technology, Sydney
Honours Thesis 2012

Submitted in partial fulfilment of the requirements for the Degree of Bachelor of Science (Honours) in Environmental Science at the University of Technology Sydney

Student Declaration

I certify that this thesis has not already been submitted for any other degree and is not being submitted as part of the candidature for any other degree.

I also certify that the thesis has been written by me, and that any help I have received in preparing this thesis, and all sources used, have been acknowledged in this thesis. I also certify that I am aware that plagiarism software may be used in the examination of this thesis.

I certify that the word count is: 15,885

Student signature:

Date: 18th June 2012

Abstract

Tissue-cultured plants (*Syngonium podophyllum*) planted into conventional potting mix and hydroculture were investigated for their capacities to bring about reductions of the two major types of indoor air pollution; volatile organic compounds (VOCs) and CO₂.

The results confirm that, with a moderate increase in indoor light intensity, the species used could be developed and used to remove significant amounts of indoor CO₂. The results also indicate that hydroculture as a growth medium makes for greater efficacy of CO₂ removal than potting mix. Furthermore, the VOC removing potential of hydroculture plants was demonstrated. Whilst the rate of VOC removal was somewhat slower than plants grown in traditional potting mix, the simultaneous capacity of the system for effective CO₂ removal is evidence that hydroculture is a more effective system for functional indoor plants than the potting mix systems that are used now.

An examination was also made of the possibility of bioaugmentation with both rhizosphere bacteria and arbuscular mycorrhizal fungi to improve the performance of the hydroculture plants to improve growth and remove more VOCs, while maintaining a lower microbial load than potting mix, so as to reduce soil CO₂ emissions, however, the efforts trialled here in this species were unsuccessful.

The effect of benzene on the community level physiological profiles of rhizospheric bacteria of hydroculture plants was assessed. Whilst the bacterial community present in hydroculture was reduced in diversity compared to potting mix, the species present encompassed at least some of those involved with VOC removal, thus indicating that hydroculture plants should still be an effective means of reducing indoor VOC concentrations.

A qualitative screen for pathogenic fungal spores from plants grown both in hydroculture and potting mix showed the nutrient solution and supporting media did not harbour any pathogenic fungi and are thus unlikely to pose a major health risk.

Table of Contents

Student Declaration	i
Abstract	ii
List of Figures	vi
List of Tables	vii
List of Abbreviations	viii
Acknowledgements	x
1. INTRODUCTION	1
1.1 General aims	1
1.2 Health impacts of air pollution	1
1.3 Association between VOC & CO ₂ concentration and Sick Building Syndrome	2
1.4 Ventilation and indoor air quality	3
1.5 Indoor plants can improve indoor air quality	4
1.6 Potential of indoor plants to reduce air-conditioning costs	5
1.7 CO ₂ reduction by indoor plants	6
1.8 Hydroculture for ornamental plants	6
1.9 Hydroculture plants' capacity to improve air quality.	7
1.10 Bioaugmentation of benzene degrading rhizospheric organisms	8
1.11 Tissue culture acclimatization and the potential for AMF to assist in hardening	9
1.12 EXPERIMENTAL AIMS	10
1.13 Outcomes	11
2. MATERIAL AND METHODS	12
2.1 Hydroculture pot design and concept.	12
2.2 Plant Materials	13
2.3 Substrate media and growth conditions	13
2.4 Selection of hydroculture growth medium	14
2.5 Measurement of physiological health on transplanting from tissue culture	15
2.6. Inoculation to enhance VOC degrading capabilities.	15
2.6.1 Hydroponic inoculation – METHOD 1	16
2.6.2 Hydroponic inoculation – METHOD 2	17
2.6.3 Screening of inocula	18
2.6.3.1 Testing for VOC removal capacity	18

	2.6.3.2 Testing roots for arbuscular mycorrhizas	20
	Treatments summary	21
	2.7 Indoor <i>S. podophyllum</i> CO ₂ reduction capacity	22
	2.7.1 Estimating leaf based light compensation points	22
	2.7.2 Measurement of whole-potted-plant CO2 fluxes	23
	2.8 VOC Removal	24
	2.8.1 Benzene chamber control	25
	2.9 VOC effects on bacterial community level physiological profiles (CLPPs)	26
	2.10 Hazardous Mould risk analysis.	27
	2.11 Data analysis	28
3.	RESULTS AND DISCUSSION	30
	3.1 Transplant success of tissue cultured plantlets to potting mix and hydroculture treatments.	30
	3.2 Inoculation treatments and success of plants after bioaugmentation	32
	3.3 Light compensation point for <i>Syngonium podophyllum</i> grown in lab conditions	36
	3.4 Whole potted plant CO ₂ fluxes.	37
	3.4.1 Under normal indoor light intensity	37
	3.3.2 Under maximum indoor light intensity	39
	3.4 Benzene removal	41
	3.4.1 Comparison of rates among treatments	41
	Why is potting mix more efficient than the other treatments?	42
	Failure of improved VOC removal with bioaugmentation	43
	3.5. Community Level Physiological Profiles of the rhizosphere of plants in potting mix and hydroculture before and after exposure to benzene	45
	3.6 Qualitative screen for pathogenic fungal spores from indoor <i>S. podophyllum</i> grown in hydroculture and potting mix.	48
4.	SIGNIFICANCE OF FINDINGS	51
	4.1 Comparisons of the performance of plants in hydroculture and potting mix	51
	4.1.1 Survival and growth	51
	4.1.2 Responses to hydroculture bioaugmentation treatments	51
	4.1.3 CO ₂ removal	52
	4.1.4 VOC removal	52
	4.1.5 Effects of benzene exposure on substrate bacterial communities	53
	4.1.6 Hydroculture and hazardous fungi	54
	4.2 Further investigations	54

4.3 Concluding remarks	55
REFERENCES.	56
APPENDIX	62

List of Figures

and second PCs are plotted (n=5).

Figure 2.1 Cross section of the 'self-watering' hydroculture pots.	11
Figure 2.2 Growth chambers and experimental treatments.	13
Figure 2.3 One gram fresh weight samples of <i>Spathiphyllum wallsii</i> roots used inoculation method 2.	16
Figure 2.4 Removal from test-jar air of two doses of 5 ppm, and one dose of 25 benzene, with TSB/vermiculite bacterial cultures developed from the potting mi <i>Spathiphyllum wallisii</i> compared to sterile jars of the same contents (Data are means ± n=4).	x of
Figure 2.5 Removal from test-jar air of two doses of 5 ppm, and one dose of 25 benzene, with 2 gram wet weight excised root suspended in moist vermiculite devel from the potting mix of <i>Spathiphyllum wallisii</i> compared to sterile jars of the same con (Data are means \pm SE; n=4).	oped
Figure 2.6 Benzene leakage rates, averaged across all 15 L chambers (Data are means \pm n=5).	E SE; 25
Figure 3.1 Changes in plant height for hydroculture & potting mix plants over 47 days (Means \pm SEM; Final n = 31 potting mix, 79 hydroculture).	30
Figure 3.2 Leaf number of plants in hydroculture & potting mix plants over 47 days grow (means \pm SEM; Final n = 31 potting mix, 79 hydroculture).	wth 30
Figure 3.3 Changes in leaf number for all treatments over 133 days (Means \pm SE; n = 15 hydroculture control; 35 inoculation method 1; 32 inoculation method 2; 15 inoculation method 2 control; 15 standard potting mix).	32
Figure 3.5 Condition of plants in all treatments at final harvest (Day 133).	35
Figure 3.6 Light response curve of <i>Syngonium podophyllum</i> grown in laboratory condition for 133 days (Means \pm SE; n=4).	ons 36
Figure 3.7 CO ₂ changes under a 10 μ mol PAR m ⁻² s ⁻¹ for <i>S. podophyllum</i> grown in pomix, hydroculture, and inoculated hydroculture (Means \pm SE; n=4).	otting 37
Figure 3.8 Displays all treatments (Potting mix, hydroculture, and inoculated hydroculture) CO ₂ changes under 350 μ mol PAR m ⁻² s ⁻¹ light regime (Means \pm SE; n=4).	ılture 39
Figure 3.9 Removal rates of a 25 ppm dose of benzene, for the four treatments, correcte chamber leakage (Means \pm SE; n=4).	ed for 42
Figure 3.10 Ordination biplot produced from PCA of CLPP data of pre- and post-ben	izene

exposure of hydroculture and potting mix treatments. Scores of each CLPP data for the first

45

List of Tables

- **Table 2.1** Colonisation of *Spathiphyllum wallisii* roots by arbuscular mycorrhizal fungi, subsequently used as an inoculum.
- Table 2.2 Gas chromatography parameters for benzene vapour estimation
 24
- **Table 3.1** Mortality rates of hydroculture and potting mix plants over 47 days 30
- Table 3.2 Plant mortality rates across all treatments. At Day 133 (86 days after inoculation).

31

- **Table 3.3** Leaf areas (m²), shoot fresh weight (FWT; g) and dry weight (DWT; g); root FWT and DWT for five treatments: potting mix; hydroculture control; hydroculture with inoculation method 1; hydroculture with inoculation method 2; and hydroculture inoculation method 2 control. (Means ± SE; n=4), and GLM ANOVA p-values amongst treatments for each data variable.
- **Table 3.4** GLM ANOVA comparing leaf number between five treatments at Day 133. (n= 15 hydroculture control, 35 hydroculture with inoculation method 1, 32 hydroculture with inoculation method 2; 15 hydroculture inoculation method 2 control; 15 standard potting mix).
- **Table 3.5** GLM ANOVA comparing plant height of five treatments at Day 133. (n= 15 hydroculture control, 35 hydroculture with inoculation method 1, 32 hydroculture with inoculation method 2; 15 hydroculture inoculation method 2 control; 15 standard potting mix).
- **Table 3.6** Arbuscular mycorrhizal fungal (AMF) colonization as percentage of root length colonized in *Syngonium podophyllum* in inoculated hydroculture (method 2),; hydroculture inoculated with sterile roots (method 2, control); and potting mix treatments. (Means \pm SE; n=3).
- **Table 3.7** GLM ANOVA comparing CO_2 removal among the five treatments, after a 40 minute test period at a light intensity of 10 μ mol PAR m⁻² s⁻¹ (n = 4).
- **Table 3.8** GLM ANOVA comparing the mean rates of CO_2 removal among the five treatments, after a 40 minute test period at a light intensity of 350 μ mol PAR m⁻² s⁻¹ (n = 4).
- **Table 3.9** Percent removal of benzene from 15 L chambers at 24, 48 and 72 hours and time taken to obtain 2/3 removal for all treatments, with significance tests at the times indicated (GLM ANOVA) (Means \pm SE; n=4). (Times taken to remove 65% of benzene were interpolated from Figure 3.9).
- **Table 3.10** Number of morphological species found per fungal genera, frequency of occurrence and mean counts expressed in CFU/mL found in potting-mix and hydroculture medium (n=3).*Values are of samples found only once and hence not true means. Total CFU/mL is the average of total colonies found, estimated from the spread plate method and is not the total of the mean CFU/mL of the genera found.

List of Abbreviations

ANOSIM Analysis of Similarity

ANOVA Analysis of Variation

AMF Arbuscular Mycorrhizal Fungi

CFU Colony Forming Units

CO₂ Carbon Dioxide

CO Carbon Monoxide

CLPP Community Level Physiological Profiles

GC Gas Chromatography

HP Hypersensitivity Pneumonitis

HVAC Heating, Ventilation and Air Conditioning

IAQ Indoor Air Quality

LCP Light Compensation Point

LRC Light Response Curve

NO_x Oxides of Nitrogen

OD Optical Density

PAH Polycyclic aromatic Hydrocarbons

PAR Photosynthetically Active Radiation

PCA Principal Components Analysis

ppmv Parts Per Million by volume

RBC Rose Bengal Chloramphenicol Agar

SBS Sick Building Syndrome

SO_x Oxides of Sulfur

SDX Sabouraud Dextrose Agar

TSB Tryptic Soy Broth

TVOC Total Volatile Organic Compound

UAP Urban Air Pollution

UTS University of Technology, Sydney

VOC Volatile Organic Compound

± SE Standard Error of the Mean

Acknowledgements

First and foremost I wish to thank my remarkable supervisors; Dr Fraser Torpy and Prof. Margaret Burchett. Words cannot express the gratitude and admiration I have for you both. From the blind faith you had in me when you brought me to the research group, to the continued guidance direction you gave me. No matter how busy you both were in your work and personal lives, I always felt that you went over and beyond the amount of effort any other honours supervisor gave their students. My experience with the UTS Plants and Indoor Environmental Quality Group will be one that I never forget, and I am sincerely grateful to have you guys in my life.

I would like to acknowledge the National Indoor Plants Association (NIPA) for the generous research grant used to conduct the experiments. NIPA is an industry peak body, promoting the use and appreciation of interior plants, including supporting research such as this project into potential benefits to health and wellbeing of indoor plant use.

Thank you to Lou de Filippis for the continuous advice and instruction.

Thank you to Jason Brennan for your continual offers for assistance and interest in the research.

Thank you to my fellow honours candidates. You guys are awesome and inspire me to strive harder simply because you were all so passionate and skilled in your chosen fields.

Thank you to my lab assistants and anyone who helped with watering my plants.

Thank you to all my family and friends who continually gave me support throughout the year. Thank you to my parents who have always been there for me. Thank you to my grandmother who dealt with my whinging when I had a broken wrist.